

## ANNEX REPORT IPI2022

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## ANNEX I: Form 1 return slip and checklist



**IPI2022-OCHABS-1  
FORM 1: CHECKLIST CONFIRMATION**

Please ensure to complete the table below upon receipt of samples, then scan and e-mail to [rsalas@observatoriocanariohabs.com](mailto:rsalas@observatoriocanariohabs.com)  
Tel: +34 623428236

Analyst Name:		
Laboratory Name:		
Analyst Code Assigned :		
Contact Tel. No. / e-mail		
<b>CHECKLIST OF ITEMS RECEIVED (Please circle the relevant answer)</b>		
Sample code Numbers: _____	YES	NO
Set of Instructions	YES	NO
Envelope containing 4 x ampoules, droppers, lugols iodine and 4 x 50ml sterilin tubes	YES	NO
Enumeration and identification result log sheet (Form 2)	YES	NO

I confirm that I have received the items as detailed above and that the materials were received in good working conditions.

(If any of the above items are missing, please contact [rsalas@observatoriocanariohabs.com](mailto:rsalas@observatoriocanariohabs.com)) or tel: + 34 623428236

**SIGNED:** \_\_\_\_\_

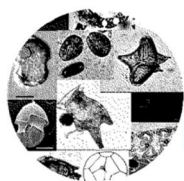
**DATE:** \_\_\_\_\_

Form 1: Checklist confirmation

**ANNEX II: Form 2 Enumeration and identification results log sheet**

IPI2022-OCHABS Phytoplankton Intercomparison Exercise									
Analyst Name:									
Laboratory Code:									
Analyst Code :									
Settlement date:									
Volume Chamber (ml)									
Analysis date:									
Sample No:									
Organism	Cell count	Cell count	Cell count	Multiplication factor	Number cells/L	Number cells/L	Number cells/L	Average	
								#DIV/0!	
								#DIV/0!	
								#DIV/0!	
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Comments:									
Form 2: Results logsheet									

## ANNEX III: Test instructions



I P I

INTERNATIONAL PHYTOPLANKTON  
INTERCOMPARISON



Observatorio Canario de algas nocivas



OceanTeacher Global Academy

### IPI Phytoplankton Proficiency Test 2022 Instructions

- 1. Introduction**
- 2. Deadlines, checklists and forms**
- 3. Test method**
- 4. Equipment**
- 5. Sedimentation chambers and sample preparation**
- 6. Counting procedure and strategy**
- 7. Samples**



## **1. Introduction**

From 2021 to 2025, the IPI Proficiency Testing scheme in abundance and composition of marine microalgae programme moves to the Canary Islands Harmful Algal Observatory (OCHABs) in the University of Las Palmas de Gran Canaria (ULPGC), Spain. The programme continues with the regular collaboration of the IOC UNESCO Centre for Science and Communication of Harmful algae in Denmark (<http://hab.ioc-unesco.org>). This collaboration involves the use of algal cultures from the Scandinavian Culture Collection of Algae and Protozoa in Copenhagen, the elaboration of a marine phytoplankton taxonomy assessment (online HAB quiz) using the online platform 'Ocean Teacher' Global academy (OTGA), <http://classroom.oceanteacher.org/> hosted by the IODE (International Oceanographic Data and information Exchange) [www.iode.org/](http://www.iode.org/) office based in Oostende, Belgium, a project office of the IOC.

The collaboration also involves the organisation of a training workshop which is held annually to discuss the results of the intercomparison exercise and to provide training on phytoplankton taxonomy. This workshop has become an important forum for phytoplankton taxonomists working on phytoplankton monitoring programmes from around the world to convene and be able to discuss taxonomical matters related to monitoring, new advances and finds, taxonomical nomenclature changes, as well as looking at samples from different geographical areas and listening to relevant stories from other laboratories about harmful algal events in their regions of relevant ecological importance. This workshop has taken the format of a full 3 days training workshop with at least 2 days dedicated to lectures on algal groups in rooms equipped with microscopes and using live cultures and preserved samples from participants and locations across the globe.

The purpose of this exercise is to compare and evaluate the performance of testing laboratories and to monitor the laboratories continuing performance over time on the composition and abundance of marine microalgae in preserved marine samples. We work mainly with laboratories engaged in national official/non-official phytoplankton monitoring programmes, water framework directive, marine strategy framework directive and others (environmental agencies, consultancies, private companies) working in the area of analysis of water samples for marine phytoplankton abundance and composition. The Canary Islands Harmful Algal Observatory (OCHABs) recognises that regular quality control assessments are crucial to ensure a high quality output of phytoplankton data. All our work is carried out following the technical and managerial requirements for PT schemes (ISO17043) and the

data is statistically analysed using the statistical methods as laid out in ISO13528. We use the statistical database software ProLab Plus from QuoData to do the statistical evaluation of the participant's data.

The web platform [www.iphyi.org](http://www.iphyi.org) was created to be a single point source of information about the IPI scheme. Registration to the exercise must be completed through this website and all the information required is contained there.

Information about this scheme can also be found through our partners, the IOC (<http://hab.ioc-unesco.org>) under the heading 'activities and training courses') and associates in the NMBAQC website ([www.nmbaqcs.org](http://www.nmbaqcs.org)) under scheme components and phytoplankton, you'll find information on the current timetable schedule for the exercise, the list of participants, previous reports and the workshop agenda from the previous exercises to give you an idea of the range of activities within this intercomparison exercise. There is also information of the other NMBAQC schemes.

Please adhere to the following instructions strictly and note that these instructions are specific to this ring test only.

## **2. Deadlines, checklists and forms**

Upon sample receipt, analysts should ensure that they received everything listed in form 1; checklist confirmation. Make sure that all the samples are intact and sealed properly and check that you have received Form 2; Enumeration and identification results log sheet (Excel workbook).

Please complete Form 1: checklist confirmation form, scan it as a pdf file and send it to me via e-mail to [rsalas@observatoriocanariohabs.com](mailto:rsalas@observatoriocanariohabs.com) . Please name the file as Form 1 followed by the exercise code and your full name **i.e. Form 1: IPI22 Rafael Salas**. This validates the traceability of the samples from origin to the laboratories and ensures that the materials arrive to the performing laboratories in good working conditions.

Analysts must complete and send their test results before or on 30/09/2022 via e-mail to [rsalas@observatoriocanariohabs.com](mailto:rsalas@observatoriocanariohabs.com) ***Please note: Results received after this date will not be included in the final report.***

Form 2 is an Excel workbook named 'Enumeration and identification logsheet' for analysts to input their results. At the top of the form, first fill in your name, analyst and laboratory code. Fill in all the information relevant to the analysis of your samples, for example the settlement date, chamber volume used in 'mls', the analysis date and the sample number in the corresponding cells.

Under the column 'organism' a drop down menu appears with a list of possible species names. You must choose from this list your answers. The list of species is a reduced list and is designed to have more entries than species are in the samples, you must choose which ones you think have been inoculated in the samples and provide a cell count. If is not in the list, is not in the sample.

The number of rows under the column name 'organism' is arbitrary and independent of the number of species in the samples. There are 14 rows but this doesn't necessarily mean that you need to enter 14 names or that there are 14 species in the samples. The number and type of species inoculated in the samples is different from year to year.

In the comments box, you can write information about the test method you used, any deviation from the Utermöhl test method and how you performed your calculations if you think is necessary.

Once you have completed your samples and have reviewed your calculations in form 2, please send your form 2 via e-mail to [rsalas@observatoriocanariohabs.com](mailto:rsalas@observatoriocanariohabs.com). Please name the file as Form 2 followed by the exercise code and your full name **i.e. Form 2: IPI22 Rafael Salas**

**In order to pass the test you must have at least 80% of your results correct. This is, 80% of your identification results and 80% of your enumeration results.**

### **3. Test method**

The Utermöhl cell counting method (Utermöhl 1931, 1958) is the standard test method used. We advise the use of 25ml sedimentation chambers for the purpose of this intercomparison exercise if these are available. If not, other sub-sample volumes and/or

chambers may be used. Do not use a different method to the Utermöhl method, please state all this information in your results.

#### **4. Equipment**

The following are the equipment requirements to complete this exercise:

Sedimentation chambers 25ml volume if possible but other volume chambers (50 ml or 10ml) can be used.

Inverted Microscope: This should be equipped with long distance working lenses up to 40 x objective or higher and condenser of Numerical Aperture (NA) of 0.3 or similar and capable for bright field microscopy. Other types of reflected or transmitted light capabilities may be helpful depending on the type of organisms in the samples and can be used if required.

Tally counters

#### **5. Sedimentation chambers and sample preparation**

Sedimentation chambers consist of a clear plastic cylinder, a metal plate, a glass disposable cover-slip base plate and a glass cover plate. Three sedimentation chambers are required.

5.1 **Storage of ampoules:** If you are not analysing the samples straightaway or if you are analysing them in different dates, please ensure the samples are kept in a fridge at 2-5°C away from direct sunlight and in an upright position.

5.2 **Temperature adaptation:** Vials must be adapted to room temperature before aliquoting and sedimentation takes place. This reduces the risk of air bubbles inside the sedimentation chambers due to temperature differences between room and sample.

5.3 **Preparation of samples for analysis from ampoules:**

5.3.1 Please follow the link here to watch a video on how to prepare your samples for analysis from an ampoule. <https://youtu.be/2WgRNGDn4MU>



Figure 1: Sample set per participant including sealed vials, lugol's iodine, plastic droppers and 50ml sterilin tubes.

**5.3.2 Do not analyse the samples directly from the ampoule. This invalidates the Utermöhl method. The sample must be made to a 50ml volume, homogenised, settled and the cell density calculations must be made from this 50 ml volume.**

5.3.3 The sterilin tubes should be prepared in advance of opening the ampoule.

5.3.4 Measure accurately 47ml of sterile seawater containing a few drops of lugol's iodine. The ampoules are already preserved in lugols, but when the sample is aliquot into the tube, it is going to be diluted and pale in colour, so if you wish your sample to have a slightly darker coloration you can add a few drops of lugols iodine to the sterile seawater before you inoculate your ampoule.

5.3.5 The volume can be measured in different ways, using an accurate pipette is one way to do it. However, you can use a gravimetric method also by weighing the amount using a balance. If you use a gravimetric method, remember that the density of Seawater at 33-35ppt is roughly 1.025g, so that  $47\text{ml} = 48.175\text{g}$  in weight.

5.3.6 The seawater used should be of a salinity of 33-35ppt

5.3.7 Once the sterilin tubes containing 47ml seawater are ready you can start working with the ampoules.

- 5.3.8 First adapt the ampoule and test tube to room temperature, before aliquoting.
- 5.3.9 Make sure the ampoule contents are at the bottom of the ampoule. If some contents are trapped on the top, flick the ampoule using your fingers to dislodge any liquid.
- 5.3.10 Break the ampoule by the neck pre-marked break line using gloves and a wad of paper to avoid cuts and grazes. Avoid losing any sample content. If you think some content is lost, you have an extra sample to work with and if this fails, ask for another set.
- 5.3.11 Use one dropper per sample, do not mix or use the same dropper. Using the dropper, aspirate the contents from the ampoule into the tube.
- 5.3.12 Once all the sample has been aliquoted into the tube, using the same dropper, take the 3ml sample from the tube itself and rinse the ampoule with it once, collect the liquid again back into the tube.
- 5.3.13 Close the lid of the tube, invert the sample 50 times minimum and pour into a sedimentation chamber of your choice.
- 5.3.14 Once the sample has been taken out of the ampoule into the tube, the sample should be settled and analysed. **Do not keep the sample in the tube for several days as this will invalidate your analysis.**

#### 5.4 **Chamber preparation:**

- 5.4.1 All sedimentation chambers should be cleaned before you start

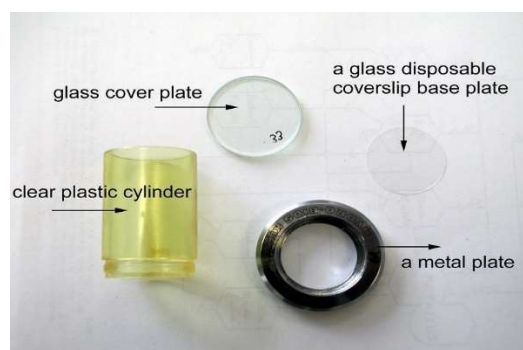


Fig 2: Sedimentation counting chamber

- 5.4.2 Place a new disposable cover slip base plate inside a cleaned metal plate.
- 5.4.3 Screw the plastic cylinder into the metal plate until tight. Extra care should be taken when setting up chambers. Disposable cover slip base plates are fragile and break easily causing cuts and grazes.
- 5.4.4 Once the chamber is set up, it should be tested for the possibility of leaks by filling the completed chamber with sterile filtered seawater and allowing it to rest for a few minutes. If no leakage occurs, pour out the water, dry out completely and proceed with the next step.

## 5.5 **Sample homogenisation and filling:**

- 5.5.1 To set up a sample for analysis, firmly invert the sample at least 50 times before pouring the sample to ensure that the contents are homogenised properly. Avoid hard shaking of the samples
- 5.5.2 Place the chamber in a flat horizontal surface protected from vibration and strong sunlight and gently pour the sample into the counting chamber to the top. Cover the chamber with the glass plate to complete the vacuum, making sure that there are no air bubbles or pockets between the sample and the cover glass.
- 5.5.3 Label the sedimentation chamber with the sample number from the ampoule.

## 5.6 **Sedimentation time:**

- 5.6.1 Settling time is dependent on the height of the chamber. 10ml chambers should be allowed to settle for a minimum of 8 hours, 25ml chambers for a minimum of 12 hours and 50ml chamber for a minimum of 24 hours.

5.6.2 Set the chamber on the inverted microscope and start the analysis.

## **6. Counting Procedure and strategy**

- a. Scan the entire chamber at low magnification first to get an initial overview of the density, distribution and composition of phytoplankton in the samples.
- b. Assess the random distribution pattern of the organisms in the sample before starting the analysis. Larger organisms tend to sediment towards the edges and smaller ones towards the centre if the temperature of the chamber is higher than the sample and vice-versa if the temperature of the chamber is lower than the sample. A visual inspection is enough to assess these patterns.
- c. If the sample is not randomly distributed, then the sample will have to be returned to its original container and settled again after a period of acclimatization. This is particularly important if other counting strategies are to be used in some organisms other than the whole chamber count, in which case, the sample count wouldn't be affected.
- d. Make a preliminary list of species and densities to help you choose the best counting strategy for the sample.
- e. Choose the correct organism/s from the dropdown species list in the Excel worksheet Form 2.
- f. Start at the lower magnification to count the larger species if present, depending on size even x 4 or x 10 objectives could be used. Then, go over the sample again at higher magnifications to count the rest of the species.
- g. The smaller species should be counted at a higher objective magnification (x 20) or x 40 if necessary.
- h. Each analyst should carry out a whole chamber cell count (WC) where possible.
- i. Other counting strategies can also be used where the cell density in the sample for a particular organism is high. Show your calculations if using a half chamber (HC), field of view (FV) or transect (Tr) counting strategy.
- j. If half of the chamber is to be counted, analyse every second transect.



- k. If a transect counting strategy is used for one or several organisms, count at least three transects and average your results. Be consistent as to which cells lying on which borders are to be counted or omitted.
- l. Fields of view should be avoided if possible but if you need to use this counting strategy, count at least ten different randomly selected fields and average your results.

## **7. Samples**

Analysts must analyse three samples in total to complete this part of the exercise. The samples are replicates. A fourth sample is additional and should be used as a replacement in case of one sample leaking or breaking. All the samples are made up in sterile filtered Seawater and spiked with culture material consisting of several species. Participants are asked to carry out a whole sedimentation chamber cell counts (where possible ; see section 6.) on each organism and sample.

The Master mix, have been made up with different aliquots of cell cultures at different concentrations and estimates have been carried out in 1ml lugol's preserved samples and counted in Sedgewick-Rafter chambers for each species. This is done to check the condition and the densities of the cultures prior to inoculating into the Master mix.

Once the master mix have been made up in a 2L brown schott glass bottle with the target species at the required concentrations, this mixture has been homogenised using an automated tumbler mixer (Inversina 2L) that uses the Paul-Schatz movement for 4 minutes at 60 rpm approximately and divided in 4 x 500ml batches. These in turn have been homogenised again at the same speed and time. 3mls of the Master mix have been inoculated using an automated multi-pipette eppendorf into a batch of 10ml brown glass ampoules, containing 100µl of neutral lugols iodine.

The ampoules have been purged using nitrogen gas and sealed using a torch. The ampoules have been checked for leaks by submerging on a water bath and then stored at 2-5°C in the dark. The ampoules have been assigned a random batch number.

Each analyst must **count and identify all phytoplankton species** found in the samples.

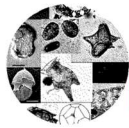
## ANNEX IV: Participating Laboratories



Ministry of Ocean Economy, Marine Resources, Fisheries, Shipping and Outer Island



## ANNEX V: Statement of performance certificate



IPI

INTERNATIONAL PHYTOPLANKTON  
INTERCOMPARISON



OCHABS

Observatorio Canario de algas nocivas

### International Phytoplankton Intercomparison (IPI) In collaboration with IOC of UNESCO & OCHABS

#### STATEMENT OF PERFORMANCE Phytoplankton Component of Community Analysis Year 2022 The participant was successful

**Participant details:**

**Name of organisation:**

**Country:**

**Participant:**

**Year of joining:**

**Years of participation:**

**Statement Issue Date:** 06/01/2023

**Statement Number:** OCHABS-IPI-22-0

**Summary of results:** At least 80% of the analytes in this certificate must be **measured and identified** successfully. Z-scores must be between -2 and +2 STDev. OceanTeacher test pass mark is 70%.

Component Name	Subcontracted	Results		Identification
		Z-score (+/- 2 Sigma limits)		
Phytoplankton abundance and composition IPI-OCHABS-2022	IOC Science and communication Centre on Harmful algae and OCHABS	<i>Actinopterychus splendens</i>		
		<i>Synedropsis sp.</i>		
		<i>Lampriscus sp.</i>		
		<i>Guinardia striata</i>		
		<i>Chaetoceros peruvianus</i>		
		<i>Prorocentrum rhathymum</i>		
		<i>Coolia sp.</i>		
Phytoplankton Taxonomy quiz IPI-OCHABS-2022	IOC Science and communication Centre on Harmful algae and OCHABS	<i>Alexandrium pacificum</i>		

np: Participant did not return any results for this component; not detected items are given a -3 score.

**Details certified by:**

Rafael Gallardo Salas

International Phytoplankton Intercomparison (IPI), Programme Manager, Research Scientist  
Observatorio Canario de Algas Nocivas (OCHABS)

## ANNEX V

### Description of Scheme components and associated performance standards

In the table overleaf, for those components on which a standard has been set, ‘Proficient’, ‘Good’, and ‘Pass’ flags indicate that the participants results met or exceeded the standards set by the IPI scheme; ‘Participated’ flag indicates that the candidate participated in the exercise but did not reach these standards. The Scheme standards are under continuous review.

Component	Annual exercise	Purpose	Description	Standard
Phytoplankton abundance and composition Exercise IPI2022	1	To assess the performance of participants using the Utermöhl cell counting technique on the analysis of prepared sample/s of Seawater preserved in Lugol’s iodine spiked using biological or synthetic materials.	Prepared marine water sample/s distributed to participants for abundance and composition of marine phytoplankton species	At least 80% of the analytes in this certificate must be measured and identified successfully to pass the test. Z-scores must be between -2 and +2 STDev.  The STDev is calculated from the consensus values using PROLab Plus software Q/Huber Algorithm and following the assessment criteria as set out in ISO13528:2015  Participants are also required to identify the organisms found in the samples correctly to the required taxon. Flags will be given as correct, incorrect or not detected
Phytoplankton Ocean Teacher online Taxonomic Assessment IPI2022	1	This online assessment allows us to assess participants’ marine phytoplankton taxonomic ability and compare those skills across laboratories.	The online taxonomic assessment is produced from scratch in the web platform Oceanteacher and designed to entice participants to study Phytoplankton taxonomic literature. The level of taxonomic proficiency required to perform well is high.	The pass mark for the Phytoplankton taxonomic assessment exercise is 70%.  Results above 90% are deemed proficient, results above 80% are deemed good, results above 70% are deemed acceptable, results below 70% are reported as “needs improvement”.

# ANNEX VI: Homogeneity and stability test using ProLab plus

## *Actinoptychus splendens* homogeneity test

IPI2022

### Survey of homogeneity test results



Sample: Homogeneity Date: 07/11/2022  
Measurand: Actinoptychus splendens

Mean: 1154  
Analytical standard deviation: 254  
Heterogeneity standard deviation s(samples): 412  
Standard deviation for proficiency assessment: 0 (Manual)

#### Results of homogeneity analysis (with statistical background)

For the homogeneity test, 10 of the prepared proficiency test items of sample Homogeneity and stability test 22 were randomly selected, and the measurand Actinoptychus splendens was analyzed 2 times. The mean across all 10 proficiency test items is 1154. The standard deviation within proficiency test items s(analytical) (=analytical precision) is 254, and the standard deviation between proficiency test items s(sample) is 412.

#### F test

According to the F test, the heterogeneity standard deviation is significantly different from 0 (significance level 5 %), therefore the proficiency test items should be considered heterogeneous according to this criterion.

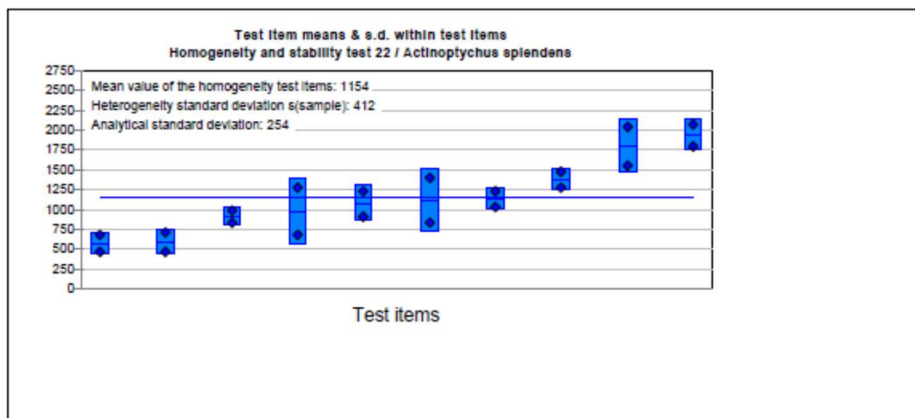
#### ISO 13528:2015 - Test for adequate homogeneity

According to ISO 13528:2015, the heterogeneity standard deviation s(sample) between the proficiency test items should not exceed 30 % of the standard deviation for proficiency assessment.

The heterogeneity standard deviation is greater than 30 % of the standard deviation for proficiency assessment 644 (Manual), therefore the proficiency test items cannot be considered as adequately homogeneous, i.e. they have to be considered heterogeneous.

#### ISO 13528:2015 - Test for significant heterogeneity

The proficiency test items exhibit significant heterogeneity (5 % significance level). The specified standard deviation for proficiency assessment is 644 (Manual). The heterogeneity standard deviation s(sample) equals 412 and is significantly too high.



# ANNEX VI: *Actinoptychus splendens* stability test

IPI2022

## Survey of stability test results



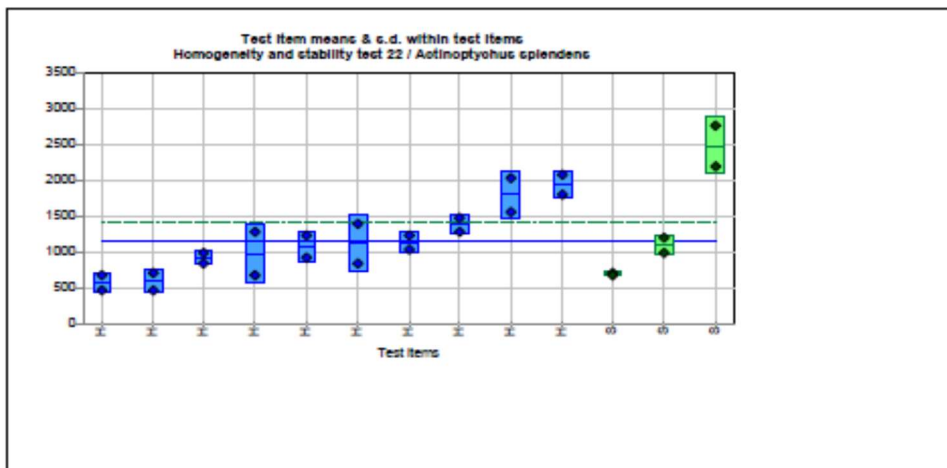
Sample:	Homogeneity	Date:	07/11/2022
Measurand:	Actinoptychus splendens		
Mean of homogeneity:	1154		
Mean of stability:	1427		
Uncertainty of mean for homogeneity measurement:	142		
Uncertainty of mean for stability measurement:	539		
Standard deviation for proficiency assessment:	0 (Manual)		

### Results of Stability Test

For the test for stability, 3 of the proficiency test items of sample Homogeneity and stability test 22 have been selected randomly and the measurand *Actinoptychus splendens* has been analyzed 2 times. The mean value across all proficiency test items of the homogeneity analysis equals 1154, the mean value across all proficiency test items of the stability analysis equals 1427. Therefore, the mean value of the stability analysis lies 23.6 % above the mean value of the homogeneity analysis.

According to ISO 13528:2015, the absolute difference between the mean values of the homogeneity analysis and the stability analysis should not exceed 30 % of the standard deviation for proficiency assessment. Although for the given standard deviation for proficiency assessment of 644, the proficiency test items may not be considered as adequately stable, the expanded acceptance criterion by adding the uncertainty of the difference to the standard deviation for proficiency assessment is fulfilled. Hence, stability of the proficiency test items is given only according to the expanded criterion of ISO 13528:2015.

By means of the t test it is checked whether the mean values of the homogeneity analysis and the stability analysis differ significantly (level of significance 5 %). The difference of the mean values is not statistically significant. Therefore the proficiency test items can be considered stable according to the t test.



quo data

07/11/2022

PROLab  
Page 1



## ANNEX VI: *Alexandrium pacificum* homogeneity test

IPI2022

### Survey of homogeneity test results



Sample: Homogeneity Date: 07/11/2022  
Measurand: *Alexandrium pacificum*

Mean: 4022  
Analytical standard deviation: 636  
Heterogeneity standard deviation s(samples): 0  
Standard deviation for proficiency assessment: 0 (Manual)

#### Results of homogeneity analysis (with statistical background)

For the homogeneity test, 10 of the prepared proficiency test items of sample Homogeneity and stability test 22 were randomly selected, and the measurand *Alexandrium pacificum* was analyzed 2 times. The mean across all 10 proficiency test items is 4022. The standard deviation within proficiency test items s(analytical) (=analytical precision) is 636, and the standard deviation between proficiency test items s(sample) is 0.

#### F test

The heterogeneity standard deviation s(sample) is 0, and hence no statistically significant difference to 0 can be detected by the F test.

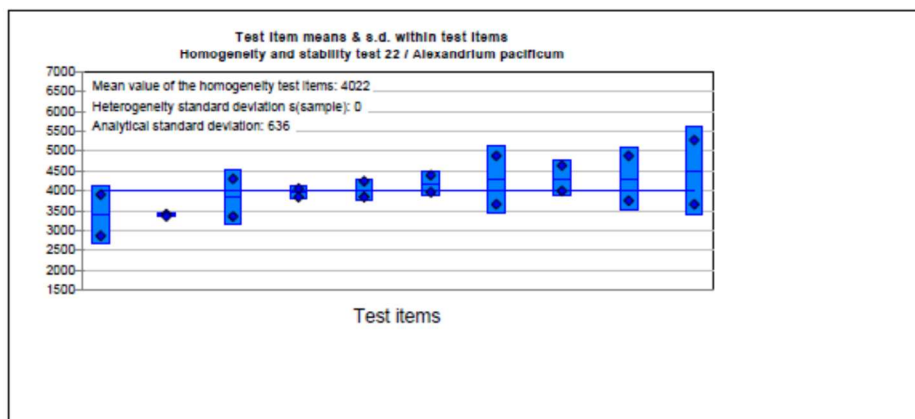
#### ISO 13528:2015 - Test for adequate homogeneity

According to ISO 13528:2015, the heterogeneity standard deviation s(sample) between the proficiency test items should not exceed 30 % of the standard deviation for proficiency assessment.

The heterogeneity standard deviation is less than 30 % of the standard deviation for proficiency assessment 725 (Manual), therefore the proficiency test items can be considered adequately homogeneous according to ISO 13528:2015.

#### ISO 13528:2015 - Test for significant heterogeneity

For the proficiency test items, no significant heterogeneity can be identified, therefore they can be considered homogeneous.



# ANNEX VI: *Alexandrium pacificum* stability test

IP12022



## Survey of stability test results

Sample: Homogeneity  
Measurand: *Alexandrium pacificum*

Date: 07/11/2022

Mean of homogeneity: 4022  
Mean of stability: 4320  
Uncertainty of mean for homogeneity measurement: 142  
Uncertainty of mean for stability measurement: 231  
Standard deviation for proficiency assessment: 0 (Manual)

### Results of Stability Test

For the test for stability, 3 of the proficiency test items of sample Homogeneity and stability test 22 have been selected randomly and the measurand *Alexandrium pacificum* has been analyzed 2 times.

The mean value across all proficiency test items of the homogeneity analysis equals 4022, the mean value across all proficiency test items of the stability analysis equals 4320.

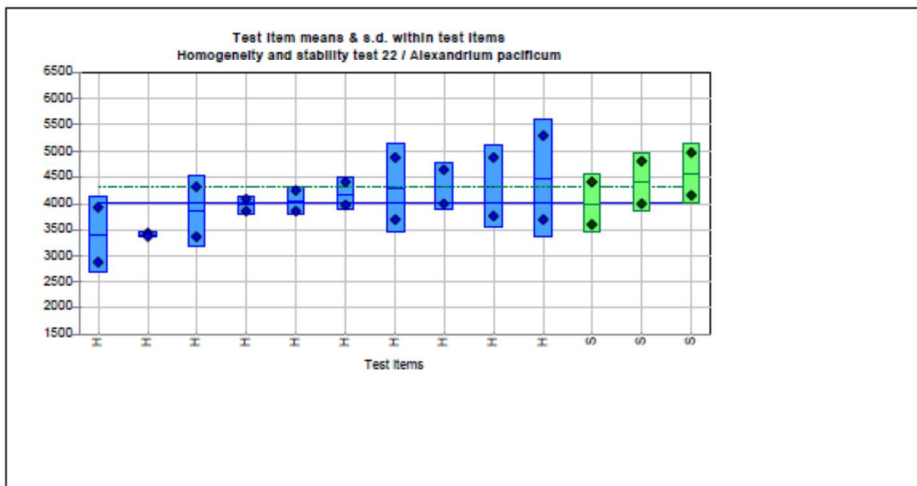
Therefore, the mean value of the stability analysis lies 7.4 % above the mean value of the homogeneity analysis.

According to ISO 13528:2015, the absolute difference between the mean values of the homogeneity analysis and the stability analysis should not exceed 30 % of the standard deviation for proficiency assessment.

Although for the given standard deviation for proficiency assessment of 725, the proficiency test items may not be considered as adequately stable, the expanded acceptance criterion by adding the uncertainty of the difference to the standard deviation for proficiency assessment is fulfilled. Hence, stability of the proficiency test items is given only according to the expanded criterion of ISO 13528:2015.

By means of the t test it is checked whether the mean values of the homogeneity analysis and the stability analysis differ significantly (level of significance 5 %).

The difference of the mean values is not statistically significant. Therefore the proficiency test items can be considered stable according to the t test.





## ANNEX VI: *Coolia monotis* homogeneity test

IPI2022

### Survey of homogeneity test results



Sample: Homogeneity  
Measurand: *Coolia monotis*

Date: 07/11/2022

Mean: 6308  
Analytical standard deviation: 966  
Heterogeneity standard deviation  $s(\text{samples})$ : 0  
Standard deviation for proficiency assessment: 0 (Manual)

#### Results of homogeneity analysis (with statistical background)

For the homogeneity test, 10 of the prepared proficiency test items of sample Homogeneity and stability test 22 were randomly selected, and the measurand *Coolia monotis* was analyzed 2 times. The mean across all 10 proficiency test items is 6308. The standard deviation within proficiency test items  $s(\text{analytical})$  (=analytical precision) is 966, and the standard deviation between proficiency test items  $s(\text{sample})$  is 0.

#### F test

The heterogeneity standard deviation  $s(\text{sample})$  is 0, and hence no statistically significant difference to 0 can be detected by the F test.

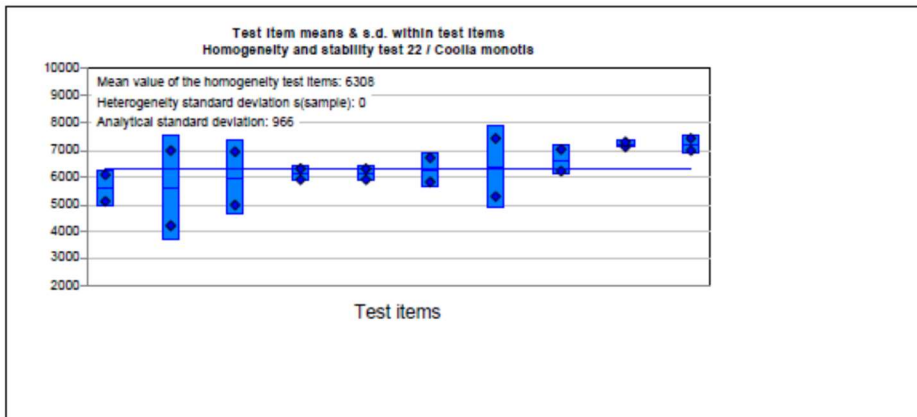
#### ISO 13528:2015 - Test for adequate homogeneity

According to ISO 13528:2015, the heterogeneity standard deviation  $s(\text{sample})$  between the proficiency test items should not exceed 30 % of the standard deviation for proficiency assessment.

The heterogeneity standard deviation is less than 30 % of the standard deviation for proficiency assessment 1115 (Manual), therefore the proficiency test items can be considered adequately homogeneous according to ISO 13528:2015.

#### ISO 13528:2015 - Test for significant heterogeneity

For the proficiency test items, no significant heterogeneity can be identified, therefore they can be considered homogeneous.



# ANNEX VI: *Coolia monotis* stability test

IPI2022

## Survey of stability test results



Sample: Homogeneity  
Measurand: *Coolia monotis*

Date: 07/11/2022

Mean of homogeneity: 6308  
Mean of stability: 6853  
Uncertainty of mean for homogeneity measurement: 216  
Uncertainty of mean for stability measurement: 430  
Standard deviation for proficiency assessment: 0 (Manual)

### Results of Stability Test

For the test for stability, 3 of the proficiency test items of sample Homogeneity and stability test 22 have been selected randomly and the measurand *Coolia monotis* has been analyzed 2 times.

The mean value across all proficiency test items of the homogeneity analysis equals 6308, the mean value across all proficiency test items of the stability analysis equals 6853.

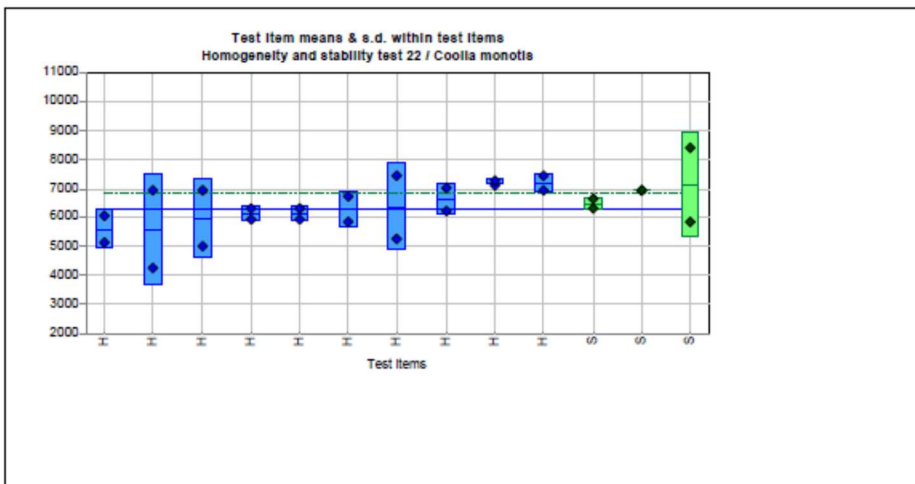
Therefore, the mean value of the stability analysis lies 8.6 % above the mean value of the homogeneity analysis.

According to ISO 13528:2015, the absolute difference between the mean values of the homogeneity analysis and the stability analysis should not exceed 30 % of the standard deviation for proficiency assessment.

Although for the given standard deviation for proficiency assessment of 1115, the proficiency test items may not be considered as adequately stable, the expanded acceptance criterion by adding the uncertainty of the difference to the standard deviation for proficiency assessment is fulfilled. Hence, stability of the proficiency test items is given only according to the expanded criterion of ISO 13528:2015.

By means of the t test it is checked whether the mean values of the homogeneity analysis and the stability analysis differ significantly (level of significance 5 %).

The difference of the mean values is not statistically significant. Therefore the proficiency test items can be considered stable according to the t test.



## ANNEX VI: *Chaetoceros peruvianus* homogeneity test

IPI2022

### Survey of homogeneity test results



Sample: Homogeneity  
Measurand: *Chaetoceros peruvianus*

Date: 07/11/2022

Mean: 15920  
Analytical standard deviation: 1771  
Heterogeneity standard deviation s(samples): 1395  
Standard deviation for proficiency assessment: 0 (Manual)

#### Results of homogeneity analysis (with statistical background)

For the homogeneity test, 10 of the prepared proficiency test items of sample Homogeneity and stability test 22 were randomly selected, and the measurand *Chaetoceros peruvianus* was analyzed 2 times. The mean across all 10 proficiency test items is 15920. The standard deviation within proficiency test items s(analytical) (=analytical precision) is 1771, and the standard deviation between proficiency test items s(sample) is 1395.

#### F test

According to the F test, the heterogeneity standard deviation is not significantly different from 0 (significance level 5%), therefore the proficiency test items can be considered sufficiently homogeneous according to this criterion.

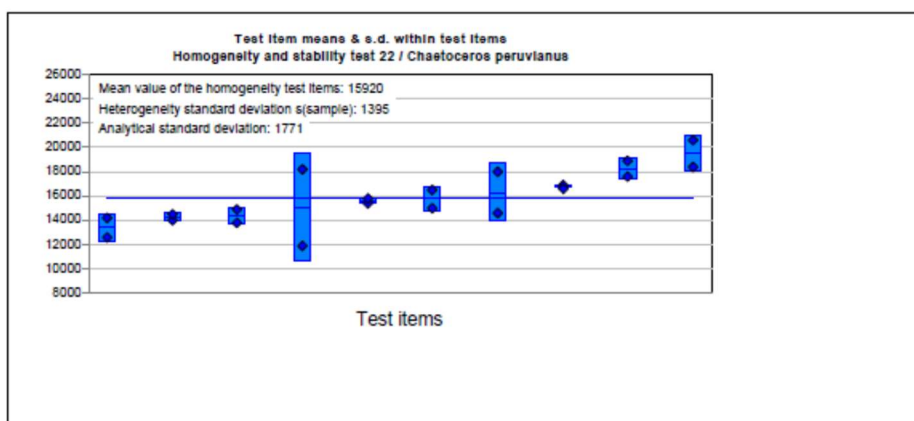
#### ISO 13528:2015 - Test for adequate homogeneity

According to ISO 13528:2015, the heterogeneity standard deviation s(sample) between the proficiency test items should not exceed 30% of the standard deviation for proficiency assessment.

The heterogeneity standard deviation is less than 30% of the standard deviation for proficiency assessment 5312 (Manual), therefore the proficiency test items can be considered adequately homogeneous according to ISO 13528:2015.

#### ISO 13528:2015 - Test for significant heterogeneity

For the proficiency test items, no significant heterogeneity can be identified, therefore they can be considered homogeneous.



# ANNEX VI: *Chaetoceros peruvianus* stability test

IPI2022

## Survey of stability test results



Sample: Homogeneity  
Measurand: *Chaetoceros peruvianus*

Date: 07/11/2022

Mean of homogeneity: 15920  
Mean of stability: 17087  
Uncertainty of mean for homogeneity measurement: 593  
Uncertainty of mean for stability measurement: 590  
Standard deviation for proficiency assessment: 0 (Manual)

### Results of Stability Test

For the test for stability, 3 of the proficiency test items of sample Homogeneity and stability test 22 have been selected randomly and the measurand *Chaetoceros peruvianus* has been analyzed 2 times.

The mean value across all proficiency test items of the homogeneity analysis equals 15920, the mean value across all proficiency test items of the stability analysis equals 17087.

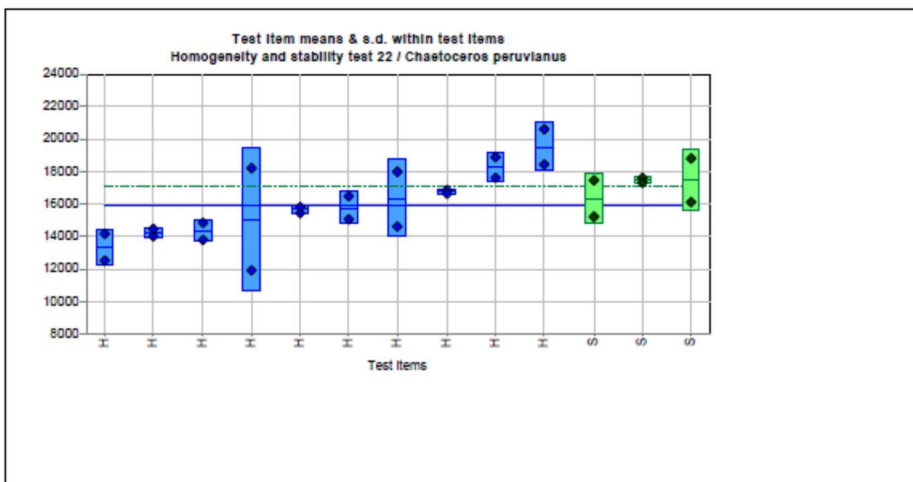
Therefore, the mean value of the stability analysis lies 7.2 % above the mean value of the homogeneity analysis.

According to ISO 13528:2015, the absolute difference between the mean values of the homogeneity analysis and the stability analysis should not exceed 30 % of the standard deviation for proficiency assessment.

Therefore, given the standard deviation for proficiency assessment of 5312, the proficiency test items may be considered as adequately stable.

By means of the t test it is checked whether the mean values of the homogeneity analysis and the stability analysis differ significantly (level of significance 5 %).

The difference of the mean values is not statistically significant. Therefore the proficiency test items can be considered stable according to the t test.



## ANNEX VI: *Guinardia striata* homogeneity test

IPI2022

### Survey of homogeneity test results



Sample: Homogeneity  
Measurand: *Guinardia striata*

Date: 07/11/2022

Mean: 7962  
Analytical standard deviation: 1523  
Heterogeneity standard deviation  $s(\text{samples})$ : 746  
Standard deviation for proficiency assessment: 0 (Manual)

#### Results of homogeneity analysis (with statistical background)

For the homogeneity test, 10 of the prepared proficiency test items of sample Homogeneity and stability test 22 were randomly selected, and the measurand *Guinardia striata* was analyzed 2 times. The mean across all 10 proficiency test items is 7962. The standard deviation within proficiency test items  $s(\text{analytical})$  (=analytical precision) is 1523, and the standard deviation between proficiency test items  $s(\text{sample})$  is 746.

#### F test

According to the F test, the heterogeneity standard deviation is not significantly different from 0 (significance level 5%), therefore the proficiency test items can be considered sufficiently homogeneous according to this criterion.

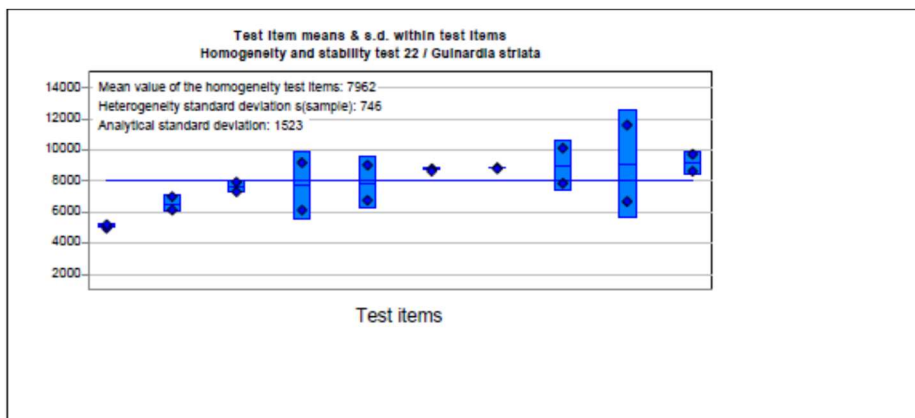
#### ISO 13528:2015 - Test for adequate homogeneity

According to ISO 13528:2015, the heterogeneity standard deviation  $s(\text{sample})$  between the proficiency test items should not exceed 30% of the standard deviation for proficiency assessment.

The heterogeneity standard deviation is greater than 30% of the standard deviation for proficiency assessment 2112 (Manual), therefore the proficiency test items cannot be considered as adequately homogeneous, i.e. they have to be considered heterogeneous.

#### ISO 13528:2015 - Test for significant heterogeneity

For the proficiency test items, no significant heterogeneity can be identified, although the heterogeneity standard deviation is greater than 30% of the standard deviation for proficiency assessment. Hence, the proficiency test items can be considered homogeneous.





# ANNEX VI: *Guinardia striata* stability test

IPI2022

## Survey of stability test results



Sample: Homogeneity  
Measurand: *Guinardia striata*

Date: 07/11/2022

Mean of homogeneity: 7962  
Mean of stability: 9960  
Uncertainty of mean for homogeneity measurement: 414  
Uncertainty of mean for stability measurement: 805  
Standard deviation for proficiency assessment: 0 (Manual)

### Results of Stability Test

For the test for stability, 3 of the proficiency test items of sample Homogeneity and stability test 22 have been selected randomly and the measurand *Guinardia striata* has been analyzed 2 times.

The mean value across all proficiency test items of the homogeneity analysis equals 7962, the mean value across all proficiency test items of the stability analysis equals 9960.

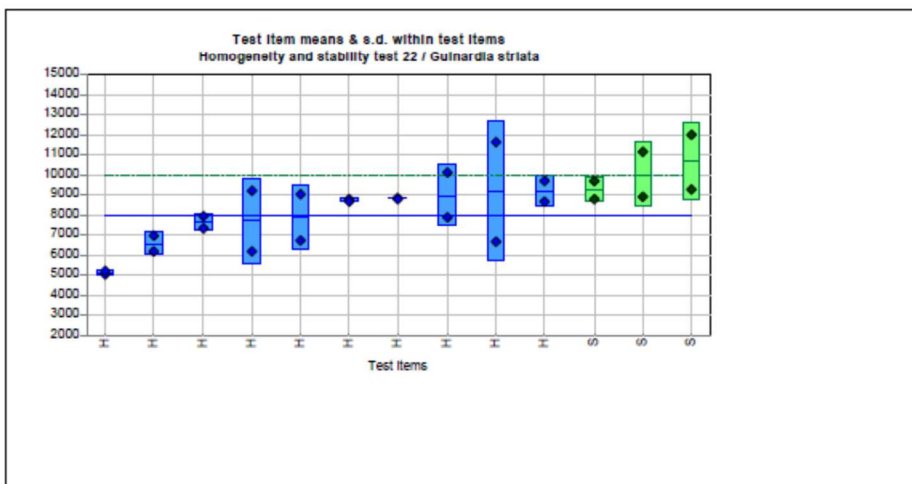
Therefore, the mean value of the stability analysis lies 25.1 % above the mean value of the homogeneity analysis.

According to ISO 13528:2015, the absolute difference between the mean values of the homogeneity analysis and the stability analysis should not exceed 30 % of the standard deviation for proficiency assessment.

Although for the given standard deviation for proficiency assessment of 2112, the proficiency test items may not be considered as adequately stable, the expanded acceptance criterion by adding the uncertainty of the difference to the standard deviation for proficiency assessment is fulfilled. Hence, stability of the proficiency test items is given only according to the expanded criterion of ISO 13528:2015.

By means of the t test it is checked whether the mean values of the homogeneity analysis and the stability analysis differ significantly (level of significance 5 %).

There is a statistically significant difference between the mean values. Therefore the proficiency test items cannot be considered stable according to the t test.



## ANNEX VI: *Lampriscus sp.* homogeneity test

IPI2022

### Survey of homogeneity test results



Sample: Homogeneity Date: 07/11/2022  
Measurand: Lampriscus sp.

Mean: 1358  
Analytical standard deviation: 315  
Heterogeneity standard deviation s(samples): 178  
Standard deviation for proficiency assessment: 0 (Manual)

#### Results of homogeneity analysis (with statistical background)

For the homogeneity test, 10 of the prepared proficiency test items of sample Homogeneity and stability test 22 were randomly selected, and the measurand *Lampriscus sp.* was analyzed 2 times. The mean across all 10 proficiency test items is 1358. The standard deviation within proficiency test items s(analytical) (=analytical precision) is 315, and the standard deviation between proficiency test items s(sample) is 178.

#### F test

According to the F test, the heterogeneity standard deviation is not significantly different from 0 (significance level 5%), therefore the proficiency test items can be considered sufficiently homogeneous according to this criterion.

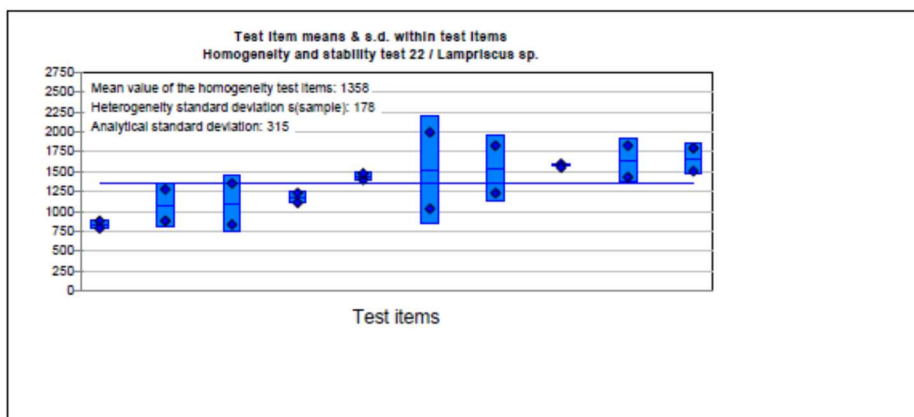
#### ISO 13528:2015 - Test for adequate homogeneity

According to ISO 13528:2015, the heterogeneity standard deviation s(sample) between the proficiency test items should not exceed 30% of the standard deviation for proficiency assessment.

The heterogeneity standard deviation is greater than 30% of the standard deviation for proficiency assessment 395 (Manual), therefore the proficiency test items cannot be considered as adequately homogeneous, i.e. they have to be considered heterogeneous.

#### ISO 13528:2015 - Test for significant heterogeneity

For the proficiency test items, no significant heterogeneity can be identified, although the heterogeneity standard deviation is greater than 30% of the standard deviation for proficiency assessment. Hence, the proficiency test items can be considered homogeneous.



# ANNEX VI: *Lampriscus sp.* stability test

IPI2022

## Survey of stability test results



Sample: Homogeneity  
Measurand: Lampriscus sp.

Date: 07/11/2022

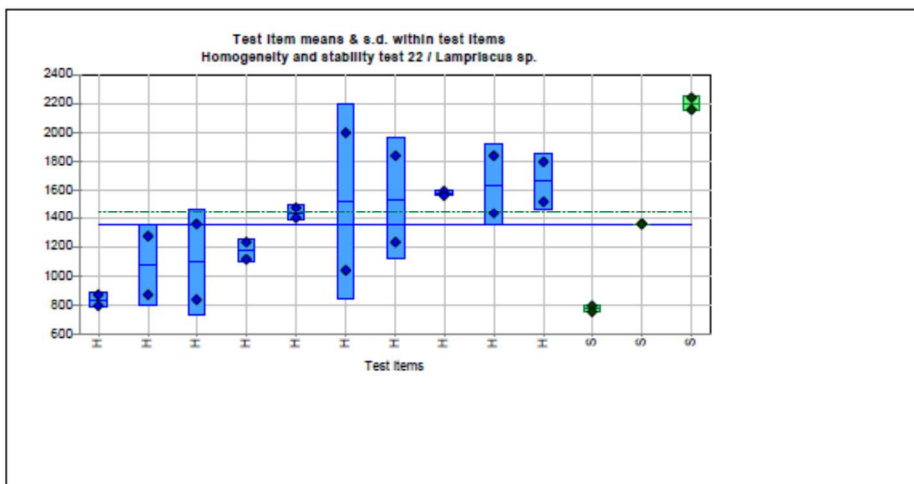
Mean of homogeneity: 1358  
Mean of stability: 1447  
Uncertainty of mean for homogeneity measurement: 90  
Uncertainty of mean for stability measurement: 412  
Standard deviation for proficiency assessment: 0 (Manual)

### Results of Stability Test

For the test for stability, 3 of the proficiency test items of sample Homogeneity and stability test 22 have been selected randomly and the measurand *Lampriscus sp.* has been analyzed 2 times. The mean value across all proficiency test items of the homogeneity analysis equals 1358, the mean value across all proficiency test items of the stability analysis equals 1447. Therefore, the mean value of the stability analysis lies 6.5 % above the mean value of the homogeneity analysis.

According to ISO 13528:2015, the absolute difference between the mean values of the homogeneity analysis and the stability analysis should not exceed 30 % of the standard deviation for proficiency assessment. Therefore, given the standard deviation for proficiency assessment of 395, the proficiency test items may be considered as adequately stable.

By means of the t test it is checked whether the mean values of the homogeneity analysis and the stability analysis differ significantly (level of significance 5 %). The difference of the mean values is not statistically significant. Therefore the proficiency test items can be considered stable according to the t test.





## ANNEX VI: *Prorocentrum rathymum* homogeneity test

IPI2022

### Survey of homogeneity test results



Sample: Homogeneity  
Measurand: Prorocentrum rathymum

Date: 07/11/2022

Mean: 7570  
Analytical standard deviation: 685  
Heterogeneity standard deviation s(samples): 543  
Standard deviation for proficiency assessment: 0 (Manual)

#### Results of homogeneity analysis (with statistical background)

For the homogeneity test, 10 of the prepared proficiency test items of sample Homogeneity and stability test 22 were randomly selected, and the measurand Prorocentrum rathymum was analyzed 2 times. The mean across all 10 proficiency test items is 7570. The standard deviation within proficiency test items s(analytical) (=analytical precision) is 685, and the standard deviation between proficiency test items s(sample) is 543.

#### F test

According to the F test, the heterogeneity standard deviation is not significantly different from 0 (significance level 5%), therefore the proficiency test items can be considered sufficiently homogeneous according to this criterion.

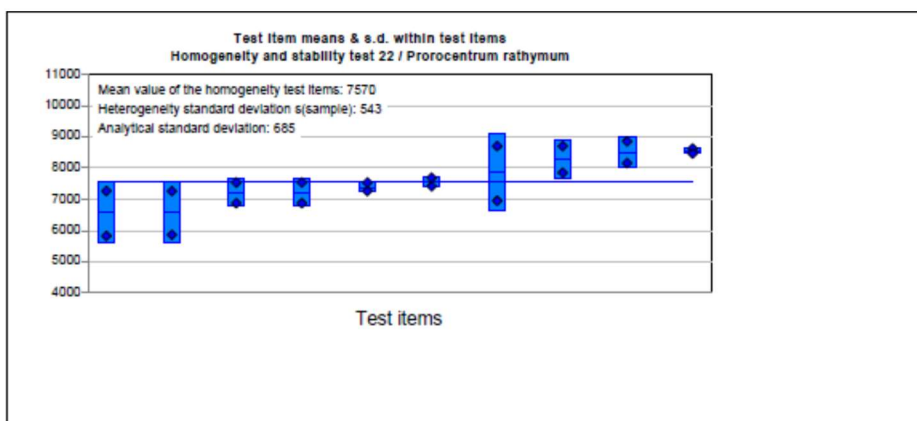
#### ISO 13528:2015 - Test for adequate homogeneity

According to ISO 13528:2015, the heterogeneity standard deviation s(sample) between the proficiency test items should not exceed 30% of the standard deviation for proficiency assessment.

The heterogeneity standard deviation is greater than 30% of the standard deviation for proficiency assessment 1574 (Manual), therefore the proficiency test items cannot be considered as adequately homogeneous, i.e. they have to be considered heterogeneous.

#### ISO 13528:2015 - Test for significant heterogeneity

For the proficiency test items, no significant heterogeneity can be identified, although the heterogeneity standard deviation is greater than 30% of the standard deviation for proficiency assessment. Hence, the proficiency test items can be considered homogeneous.



# ANNEX VI: *Prorocentrum rathymum* stability test

IPI2022

## Survey of stability test results



Sample: Homogeneity Date: 07/11/2022  
Measurand: Prorocentrum rathymum

Mean of homogeneity: 7570  
Mean of stability: 8400  
Uncertainty of mean for homogeneity measurement: 230  
Uncertainty of mean for stability measurement: 895  
Standard deviation for proficiency assessment: 0 (Manual)

### Results of Stability Test

For the test for stability, 3 of the proficiency test items of sample Homogeneity and stability test 22 have been selected randomly and the measurand Prorocentrum rathymum has been analyzed 2 times.

The mean value across all proficiency test items of the homogeneity analysis equals 7570, the mean value across all proficiency test items of the stability analysis equals 8400.

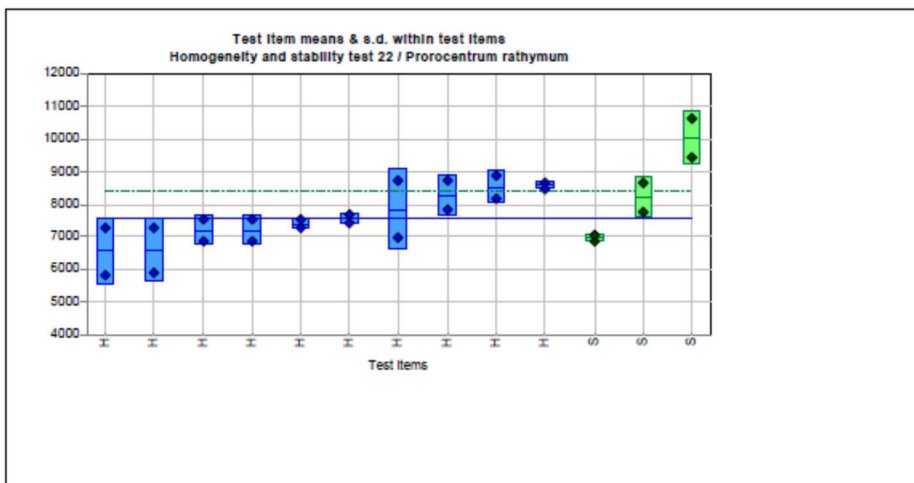
Therefore, the mean value of the stability analysis lies 11.0 % above the mean value of the homogeneity analysis.

According to ISO 13528:2015, the absolute difference between the mean values of the homogeneity analysis and the stability analysis should not exceed 30 % of the standard deviation for proficiency assessment.

Although for the given standard deviation for proficiency assessment of 1574, the proficiency test items may not be considered as adequately stable, the expanded acceptance criterion by adding the uncertainty of the difference to the standard deviation for proficiency assessment is fulfilled. Hence, stability of the proficiency test items is given only according to the expanded criterion of ISO 13528:2015.

By means of the t test it is checked whether the mean values of the homogeneity analysis and the stability analysis differ significantly (level of significance 5 %).

The difference of the mean values is not statistically significant. Therefore the proficiency test items can be considered stable according to the t test.



# ANNEX VI: *Synedropsis sp.* homogeneity test

IPI2022

## Survey of homogeneity test results



Sample: Homogeneity Date: 07/11/2022  
Measurand: *Synedropsis sp.*

Mean: 3062  
Analytical standard deviation: 361  
Heterogeneity standard deviation s(samples): 331  
Standard deviation for proficiency assessment: 0 (Manual)

### Results of homogeneity analysis (with statistical background)

For the homogeneity test, 10 of the prepared proficiency test items of sample Homogeneity and stability test 22 were randomly selected, and the measurand *Synedropsis sp.* was analyzed 2 times. The mean across all 10 proficiency test items is 3062. The standard deviation within proficiency test items s(analytical) (=analytical precision) is 361, and the standard deviation between proficiency test items s(sample) is 331.

### F test

According to the F test, the heterogeneity standard deviation is not significantly different from 0 (significance level 5%), therefore the proficiency test items can be considered sufficiently homogeneous according to this criterion.

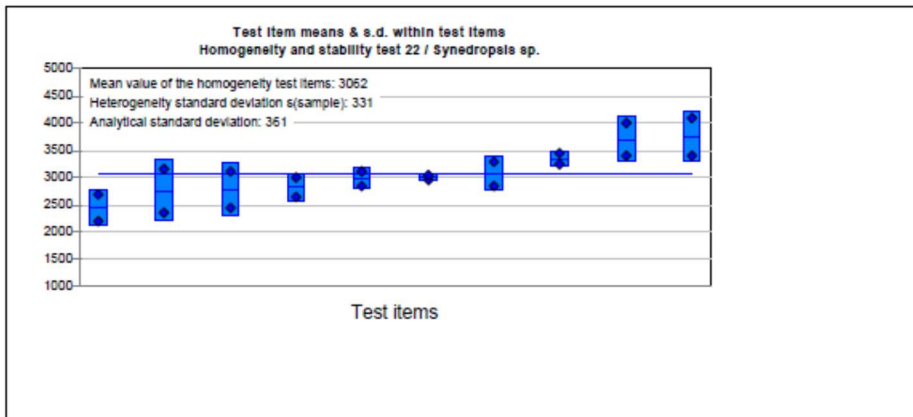
### ISO 13528:2015 - Test for adequate homogeneity

According to ISO 13528:2015, the heterogeneity standard deviation s(sample) between the proficiency test items should not exceed 30% of the standard deviation for proficiency assessment.

The heterogeneity standard deviation is greater than 30% of the standard deviation for proficiency assessment 620 (Manual), therefore the proficiency test items cannot be considered as adequately homogeneous, i.e. they have to be considered heterogeneous.

### ISO 13528:2015 - Test for significant heterogeneity

For the proficiency test items, no significant heterogeneity can be identified, although the heterogeneity standard deviation is greater than 30% of the standard deviation for proficiency assessment. Hence, the proficiency test items can be considered homogeneous.



# ANNEX VI: *Synedropsis sp.* stability test

IPI2022



## Survey of stability test results

Sample: Homogeneity Date: 07/11/2022  
Measurand: *Synedropsis sp.*

Mean of homogeneity: 3062  
Mean of stability: 3500  
Uncertainty of mean for homogeneity measurement: 132  
Uncertainty of mean for stability measurement: 424  
Standard deviation for proficiency assessment: 0 (Manual)

### Results of Stability Test

For the test for stability, 3 of the proficiency test items of sample Homogeneity and stability test 22 have been selected randomly and the measurand *Synedropsis sp.* has been analyzed 2 times.

The mean value across all proficiency test items of the homogeneity analysis equals 3062, the mean value across all proficiency test items of the stability analysis equals 3500.

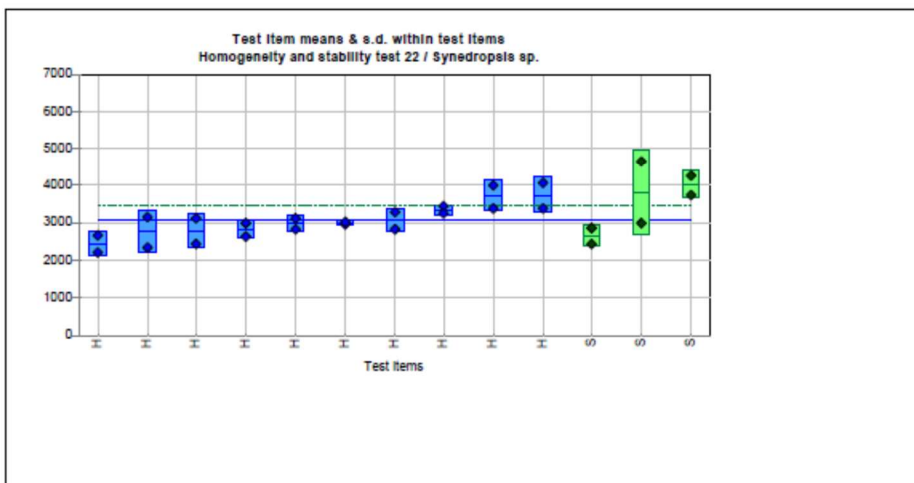
Therefore, the mean value of the stability analysis lies 14.3 % above the mean value of the homogeneity analysis.

According to ISO 13528:2015, the absolute difference between the mean values of the homogeneity analysis and the stability analysis should not exceed 30 % of the standard deviation for proficiency assessment.

Although for the given standard deviation for proficiency assessment of 626, the proficiency test items may not be considered as adequately stable, the expanded acceptance criterion by adding the uncertainty of the difference to the standard deviation for proficiency assessment is fulfilled. Hence, stability of the proficiency test items is given only according to the expanded criterion of ISO 13528:2015.

By means of the t test it is checked whether the mean values of the homogeneity analysis and the stability analysis differ significantly (level of significance 5 %).

The difference of the mean values is not statistically significant. Therefore the proficiency test items can be considered stable according to the t test.









**Annex VIII: Robust mean and Standard deviation calculation according  
to algorithm A annex C ISO13528 *Alexandrium pacificum* iteration**

IPI2022 Iteration								
<b>Alexandrium pacificum</b>								
CELLS / L								
Date	Sample	M1	M2	sample average	*2	Batch #	Label	
13.09.2021	water22	2880	3920	3400	1040	1081600	A	106
13.09.2021	water22	3960	4400	4180	440	193600	A	166
13.09.2021	water22	4080	3840	3960	240	57600	A	211
13.09.2021	water22	5280	3680	4480	1600	2560000	B	360
13.09.2021	water22	3440	3360	3400	80	6400	B	365
13.09.2021	water22	4240	3840	4040	400	160000	C	590
13.09.2021	water22	3360	4320	3840	960	921600	C	546
13.09.2021	water22	4640	4000	4320	640	409600	C	563
13.09.2021	water22	3760	4880	4320	1120	1254400	D	737
13.09.2021	water22	4880	3680	4280	1200	1440000	D	792
			Average:	4022	Sum	3899200		
			SD	379	P=	10		
			SD within samples:	442				
			SD between samples:	214				
Date	Sample number	Test portion 1	Test portion 2	sample average	Between test portion range	*2	Batch #	Label
13.10.2021	water22	4400	3600	4000	800	640000	A	154
13.10.2021	water22	4160	4960	4560	800	640000	B	326
13.10.2021	water22	4000	4800	4400	800	640000	D	716
			Average:	4320	Sum	1920000		
			SD	288	P=	3		
			SD within samples:	566				
			SD between samples:	277				
homogeneity criteria			214	114				
stability check criteria			4022	4320	298			

**Analysts' iteration for *Alexandrium pacificum***

Average X	3092		3010	3011	3011	3011
SD S	1122		613	611	611	611
robust average X*	2963	new X*	3010	3011	3011	3011
robust stdev S*	672	new S*	695	693	692	692
$\delta = 1.5S^*$	1008		1042	1039	1039	1039
$X^* - \delta$	1955		1967	1971	1972	1972
$X^* + \delta$	3972		4052	4050	4049	4049
no of analysts P	74		74	74	74	74
Between Samples SD	214					
new stdev for APACIF	725					

**Annex VIII: Robust mean and Standard deviation calculation according to algorithm A annex C ISO13528 *Prorocentrum rhathymum* iteration**

IPI2022 Iteration								
<b>Prorocentrum rhathymum</b>								
CELLS / L								
Date	Sample	M1	M2	sample average	*2	Batch #	Label	
13.09.2021	water22	7280	7520	7400	240	57600	A	106
13.09.2021	water22	5880	7280	6580	1400	1960000	A	166
13.09.2021	water22	7520	6880	7200	640	409600	A	211
13.09.2021	water22	8720	6960	7840	1760	3097600	B	360
13.09.2021	water22	5840	7280	6560	1440	2073600	B	365
13.09.2021	water22	8720	7840	8280	880	774400	C	590
13.09.2021	water22	7440	7680	7560	240	57600	C	546
13.09.2021	water22	8640	8480	8560	160	25600	C	563
13.09.2021	water22	8880	8160	8520	720	518400	D	737
13.09.2021	water22	7520	6880	7200	640	409600	D	792
			Average:	7570	Sum	7598400		
			SD	727	P=	10		
			SD within samples:	616				
			SD between samples:	582				
Date	Sample number	Test portion 1	Test portion 2	sample average	Between test portion range	*2	Batch #	Label
13.10.2021	water22	6880	7040	6960	160	25600	A	154
13.10.2021	water22	7760	8640	8200	880	774400	B	326
13.10.2021	water22	9440	10640	10040	1200	1440000	D	716
			Average:	8400	Sum	2240000		
			SD	1550	P=	3		
			SD within samples:	611				
			SD between samples:	1488				
homogeneity criteria			582	218				
stability check criteria			7570	8400	830			

**Analysts' iteration for *Prorocentrum rhathymum***

Average X	5416		5361	5361
SD S	1580		1290	1290
robust average X*	5340	new X*	5361	5361
robust stdev S*	1398	new S*	1463	1463
$\delta = 1.5S^*$	2097		2194	2194
X* - $\delta$	3243		3167	3167
X* + $\delta$	7437		7555	7555
no of analysts P	76		76	76
Between Samples SD	582			
new stdev for PRATH	1574			



**Annex VIII: Robust mean and Standard deviation calculation according to algorithm A annex C ISO13528 *Actinoptychus splendens* iteration**

IPI2022 Iteration									
<b>Actinoptychus</b>									
CELLS / L									
Date	Sample	M1	M2	sample average	*2	Batch #	Label		
13.09.2022	water22	2080	1800	1940	280	78400	A	106	
13.09.2022	water22	480	680	580	200	40000	A	166	
13.09.2022	water22	720	480	600	240	57600	A	211	
13.09.2022	water22	1240	920	1080	320	102400	B	360	
13.09.2022	water22	1280	680	980	600	360000	B	365	
13.09.2022	water22	840	1000	920	160	25600	C	590	
13.09.2022	water22	2040	1560	1800	480	230400	C	546	
13.09.2022	water22	1240	1040	1140	200	40000	C	563	
13.09.2022	water22	1480	1280	1380	200	40000	D	737	
13.09.2022	water22	1400	840	1120	560	313600	D	792	
Average:				1154	Sum	638400			
SD				449	P=	10			
SD within samples:				179					
SD between samples:				431					
Date	Sample number	Test portion 1	Test portion 2	sample average	Between test portion range	*2	Batch #	Label	
13.10.2022	water22	720	680	700	40	1600	A	154	
13.10.2022	water22	2760	2200	2480	560	313600	B	326	
13.10.2022	water22	1000	1200	1100	200	40000	D	716	
Average:				1427	Sum	355200			
SD				934	P=	3			
SD within samples:				243					
SD between samples:				918					
homogeneity criteria			431	135					
stability check criteria			1154	1427	273				

**Analysts' iteration for *Actinoptychus splendens***

Average X	1420		1376	1376
SD S	523		422	422
robust average X*	1333	new X*	1376	1376
robust stdev S*	494	new S*	478	478
$\delta = 1.5S^*$	742		718	717
$X^* - \delta$	592		658	658
$X^* + \delta$	2075		2093	2093
no of analysts P	77		77	77
Between Samples SD	431			
new stdev for ACTINO	644			

**Annex VIII: Robust mean and Standard deviation calculation according to algorithm A annex C ISO13528 *Coolia monotis* iteration**

IPI2022 Iteration								
Coolia sp.								
CELLS / L								
Date	Sample	M1	M2	sample average	*2	Batch #	Label	
13.09.2022	water22	5280	7440	6360	2160	4665600	A	106
13.09.2022	water22	5000	6920	5960	1920	3686400	A	166
13.09.2022	water22	6320	5920	6120	400	160000	A	211
13.09.2022	water22	7120	7280	7200	160	25600	B	360
13.09.2022	water22	6080	5120	5600	960	921600	B	365
13.09.2022	water22	7040	6240	6640	800	640000	C	590
13.09.2022	water22	6960	4240	5600	2720	7398400	C	546
13.09.2022	water22	7440	6960	7200	480	230400	C	563
13.09.2022	water22	5840	6720	6280	880	774400	D	737
13.09.2022	water22	5920	6320	6120	400	160000	D	792
			Average:	6308	Sum	9459200		
			SD	600	P=	10		
			SD within samples:	688				
			SD between samples:	351				
Date	Sample number	Test portion 1	Test portion 2	sample average	Between test portion range	*2	Batch #	Label
13.10.2022	water22	6320	6640	6480	320	102400	A	154
13.10.2022	water22	6960	6960	6960	0	0	B	326
13.10.2022	water22	8400	5840	7120	2560	6553600	D	716
			Average:	6853	Sum	6656000		
			SD	333	P=	3		
			SD within samples:	1053				
			SD between samples:	666				
homogeneity criteria			351	180				
stability check criteria			6308	6853	545			

**Analysts' iteration for *Coolia monotis***

Average X	4792		4797	4797
SD S	1286		933	933
robust average X*	4754	new X*	4797	4797
robust stdev S*	958	new S*	1058	1058
$\delta = 1.5S^*$	1437		1587	1587
X* - $\delta$	3316		3210	3210
X* + $\delta$	6191		6384	6384
no of analysts P	73		73	73
Between Samples SD	351			
new stdev for COOLIA	1115			

**Annex VIII: Robust mean and Standard deviation calculation according to algorithm A annex C ISO13528 *Chaetoceros peruvianus* iteration**

IPI2022 Iteration									
Chaetoceros peruvianus									
CELLS / L									
Date	Sample	M1	M2	sample average		*2	Batch #	Label	
13.09.2021	water22	17600	18880	18240	1280	1638400	A	106	
13.09.2021	water22	20560	18400	19480	2160	4665600	A	166	
13.09.2021	water22	16880	16640	16760	240	57600	A	211	
13.09.2021	water22	15040	16480	15760	1440	2073600	B	360	
13.09.2021	water22	14880	13840	14360	1040	1081600	B	365	
13.09.2021	water22	15440	15840	15640	400	160000	C	590	
13.09.2021	water22	18000	14640	16320	3360	11289600	C	546	
13.09.2021	water22	14000	14480	14240	480	230400	C	563	
13.09.2021	water22	12560	14160	13360	1600	2560000	D	737	
13.09.2021	water22	18160	11920	15040	6240	38937600	D	792	
			Average:	15920	Sum	9516800			
			SD	1874	P=	10			
			SD within samples:	690					
			SD between samples:	1809					
Date	Sample number	Test portion 1	Test portion 2	sample average	Between test portion range	*2	Batch #	Label	
13.10.2021	water22	17600	17280	17440	320	102400	A	154	
13.10.2021	water22	18800	16080	17440	2720	7398400	B	326	
13.10.2021	water22	15200	17440	16320	2240	5017600	D	716	
			Average:	17067	Sum	12518400			
			SD	647	P=	3			
			SD within samples:	1444					
			SD between samples:	791					
homogeneity criteria			1809	562					
stability check criteria			15920	17067	1147				

**Analysts' iteration for *Chaetoceros peruvianus***

Average X	15281		15732	15706	15704	15703	15703
SD S	5412		4456	4409	4405	4404	4404
robust average X*	16493	new X*	15732	15706	15704	15703	15703
robust stdev S*	5246	new S*	5053	5000	4995	4994	4994
$\delta = 1.5S^*$	7870		7580	7500	7492	7491	7491
X* - $\delta$	8624		8152	8206	8211	8212	8212
X* + $\delta$	24363		23312	23207	23196	23195	23195
no of analysts P	77		77	77	77	77	77
Between Samples SD	1809						
new stdev for PERUV	5312						

**Annex VIII: Robust mean and Standard deviation calculation according  
to algorithm A annex C ISO13528 *Guinardia striata* iteration**

IPI2022 Iteration								
<b>Guinardia striata</b>								
CELLS / L								
Date	Sample	M1	M2	sample average	*2	Batch #	Label	
13.09.2022	water22	8840	8800	8820	40	1600	A	106
13.09.2022	water22	8640	9680	9160	1040	1081600	A	166
13.09.2022	water22	5000	5200	5100	200	40000	A	211
13.09.2022	water22	7320	7920	7620	600	360000	B	360
13.09.2022	water22	8800	8640	8720	160	25600	B	365
13.09.2022	water22	10080	7840	8960	2240	5017600	C	590
13.09.2022	water22	9040	6720	7880	2320	5382400	C	546
13.09.2022	water22	6160	6960	6560	800	640000	C	563
13.09.2022	water22	6160	9200	7680	3040	9241600	D	737
13.09.2022	water22	11600	6640	9120	4960	24601600	D	792
			Average:	7962	Sum	1508800		
			SD	1660	P=	10		
			SD within samples:	275				
			SD between samples:	1649				
Date	Sample number	Test portion 1	Test portion 2	sample average	Between test portion range	*2	Batch #	Label
13.10.2022	water22	8880	11120	10000	2240	5017600	A	154
13.10.2022	water22	9280	12000	10640	2720	7398400	B	326
13.10.2022	water22	8800	9680	9240	880	774400	D	716
			Average:	9960	Sum	13190400		
homogeneity criteria			1649	498				
stability check criteria			7962	9960	1998			

**Analysts' iteration for *Guinardia striata***

Average X	6257		6339	6339
SD S	1686		1164	1164
robust average X*	6389	new X*	6339	6339
robust stdev S*	1251	new S*	1320	1320
$\delta = 1.5S^*$	1876		1980	1980
$X^* - \delta$	4513		4359	4359
$X^* + \delta$	8265		8319	8319
no of analysts P	77		77	77
Between Samples SD	1649			
new stdev for GSTRIATA	2112			

**Annex VIII: Robust mean and Standard deviation calculation according  
to algorithm A annex C ISO13528 *Lampriscus sp.* iteration**

IPI2022 Iteration								
<b>lampriscus sp.</b>								
CELLS / L								
Date	Sample	M1	M2	sample average	*2	Batch #	Label	
13.09.2022	water22	1400	1480	1440	80	6400	A	106
13.09.2022	water22	1520	1800	1660	280	78400	A	166
13.09.2022	water22	1240	1120	1180	120	14400	A	211
13.09.2022	water22	1840	1440	1640	400	160000	B	360
13.09.2022	water22	1280	880	1080	400	160000	B	365
13.09.2022	water22	880	800	840	80	6400	C	590
13.09.2022	water22	1840	1240	1540	600	360000	C	546
13.09.2022	water22	1600	1560	1580	40	1600	C	563
13.09.2022	water22	2000	1040	1520	960	921600	D	737
13.09.2022	water22	840	1360	1100	520	270400	D	792
Average:				1358	Sum	1979200		
SD				285	P=	10		
SD within samples:				315				
SD between samples:				178				
Date	Sample number	Test portion 1	Test portion 2	sample average	Between test portion range	*2	Batch #	Label
13.10.2022	water22	800	760	780	40	1600	A	154
13.10.2022	water22	2240	2160	2200	80	6400	B	326
13.10.2022	water22	1360	1360	1360	0	0	D	716
Average:				1447	Sum	8000		
SD				714	P=	3		
SD within samples:				37				
SD between samples:				713				
homogeneity criteria			178	85				
stability check criteria			1358	1447	89			

**Analysts' iteration for *Lampriscus sp.***

Average X	1481		1485	1484	1485	1485	1485
SD S	297		266	260	259	258	258
robust average X*	1502	new X*	1485	1484	1485	1485	1485
robust stdev S*	323	new S*	301	295	293	292	292
$\delta = 1.5S^*$	485		452	443	440	439	438
$X^* - \delta$	1017		1033	1042	1045	1046	1046
$X^* + \delta$	1987		1937	1927	1924	1923	1923
no of analysts P	77		77	77	77	77	77
Between Samples SD	178						
new stdev for LAMPRI	342						

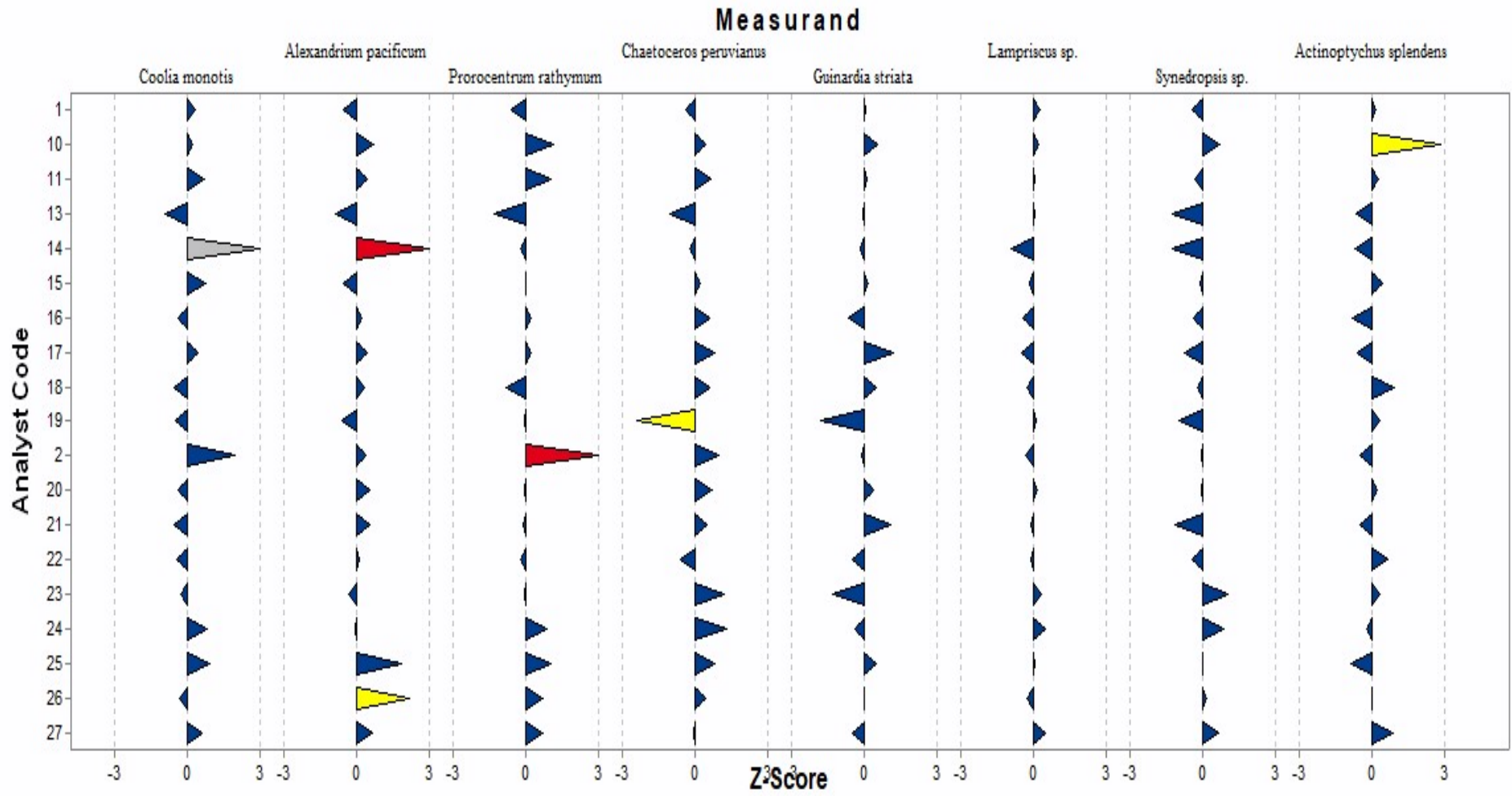
**Annex VIII: Robust mean and Standard deviation calculation according to algorithm A annex C ISO13528 *Synedropsis sp.* iteration**

IPI2022 Iteration								
<b>Synedropsis sp.</b>								
CELLS / L								
Date	Sample	M1	M2	sample average	*2	Batch #	Label	
13.09.202:	water22	3000	2640	2820	360	129600	A	106
13.09.202:	water22	2960	3040	3000	80	6400	A	166
13.09.202:	water22	3160	2360	2760	800	640000	A	211
13.09.202:	water22	3440	3240	3340	200	40000	B	360
13.09.202:	water22	2440	3120	2780	680	462400	B	365
13.09.202:	water22	4080	3400	3740	680	462400	C	590
13.09.202:	water22	3120	2840	2980	280	78400	C	546
13.09.202:	water22	3400	4000	3700	600	360000	C	563
13.09.202:	water22	2840	3280	3060	440	193600	D	737
13.09.202:	water22	2680	2200	2440	480	230400	D	792
			Average:	3062	Sum	1278400		
			SD	243	P=	10		
			SD within samples:	253				
			SD between samples:	164				
Date	Sample number	Test portion 1	Test portion 2	sample average	Between test portion range	*2	Batch #	Label
13.10.202:	water22	2440	2880	2660	440	193600	A	154
13.10.202:	water22	4640	3000	3820	1640	2689600	B	326
13.10.202:	water22	4280	3760	4020	520	270400	D	716
			Average:	3500	Sum	3153600		
homogeneity criteria			164	73				
stability check criteria			3062	3500	438			

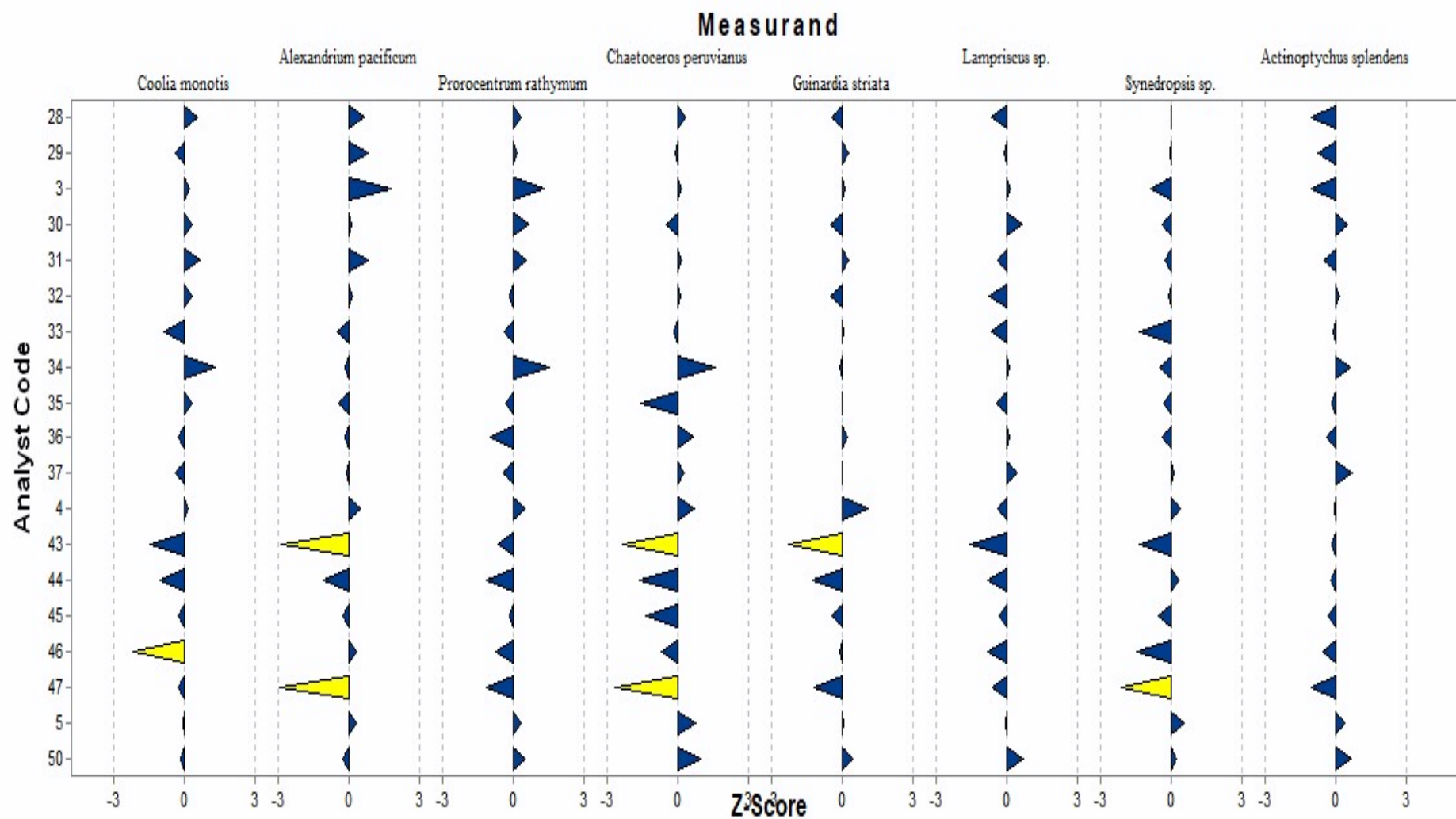
**Analysts' iteration for *Synedropsis sp.***

Average X	3491		3501	3501
SD S	695		533	533
robust average X*	3453	new X*	3501	3501
robust stdev S*	534	new S*	604	604
$\delta = 1.5S^*$	801		906	906
$X^* - \delta$	2653		2595	2595
$X^* + \delta$	4254		4407	4407
no of analysts P	77		77	77
Between Samples SD	164			
new stdev for SYNEDROP	626			

ANNEX IX: Summary of Z-scores IPI202 for all measurands pg1

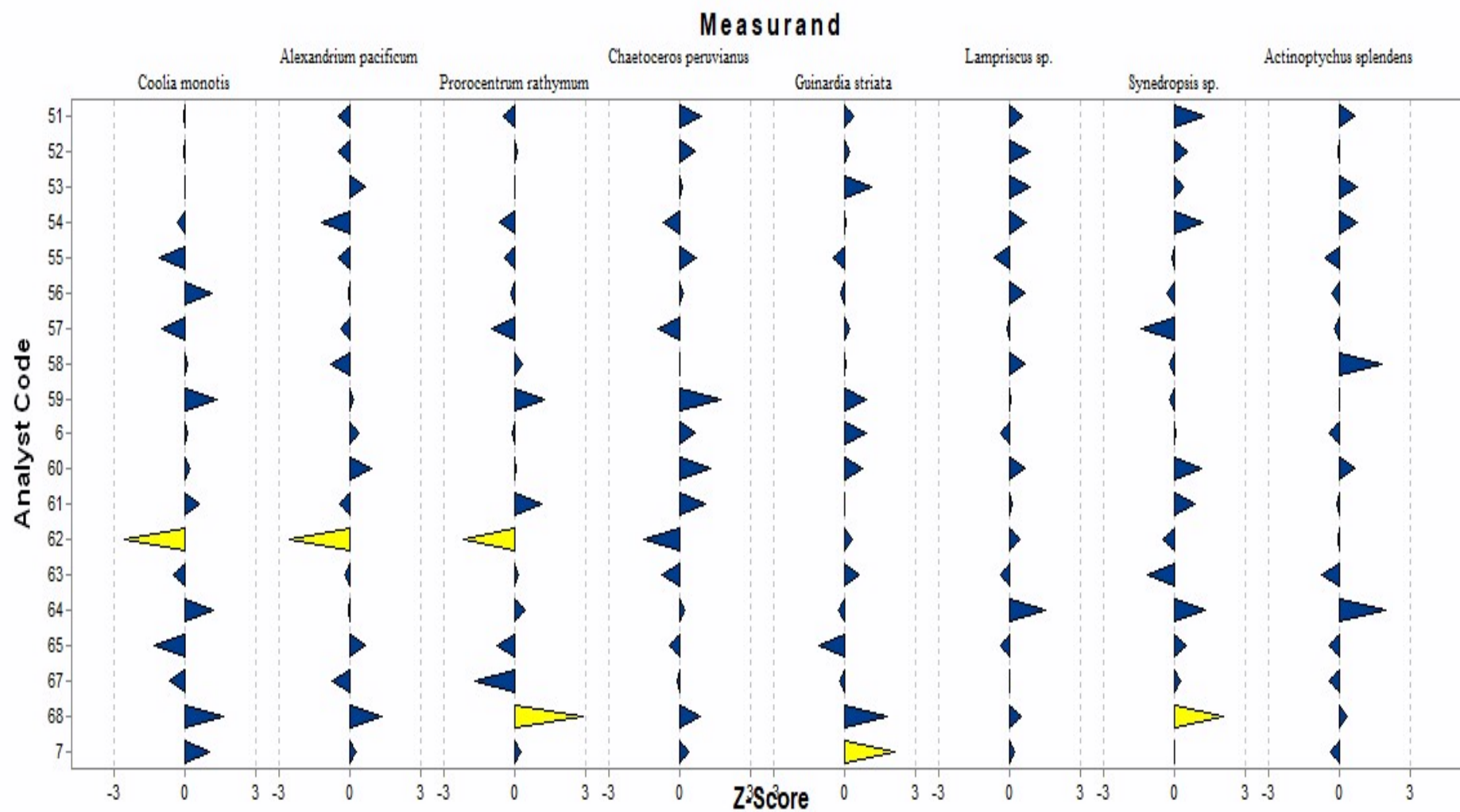


ANNEX IX: Summary of Z-scores IPI202 for all measurands pg2

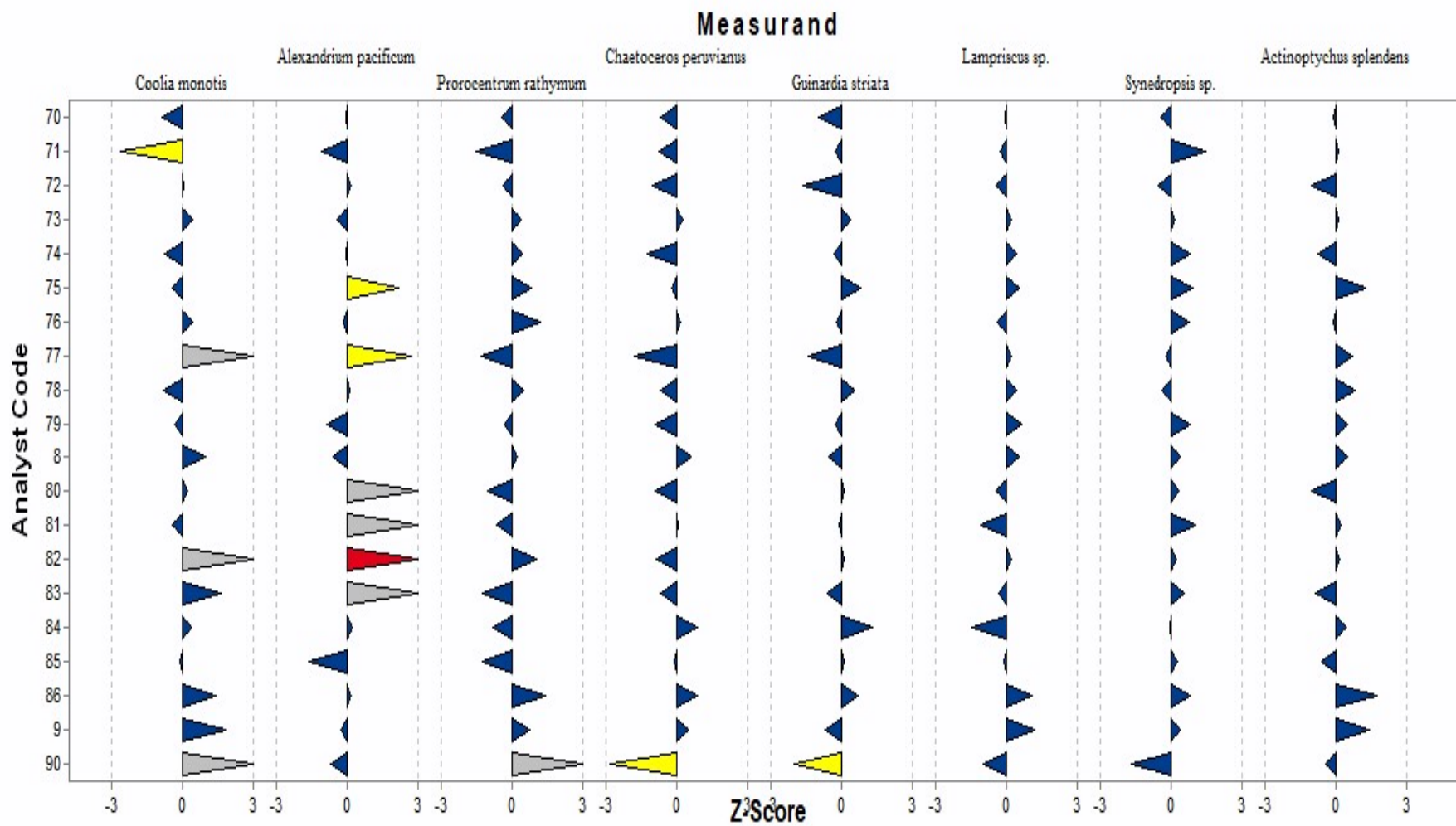




ANNEX IX: Summary of Z-scores IPI202 for all measurands pg3



ANNEX IX: Summary of Z-scores IPI202 for all measurands pg4



**ANNEX X: Performance statistics for the test IPI2022**

Analyst code	Measurands	Within tolerance	%	Successful	Analyst code	Measurands	Within tolerance	%	Successful	Analyst code	Measurands	Within tolerance	%	Successful
1	8	8	100	Yes	44	8	8	100	Yes	78	8	8	100	Yes
11	8	8	100	Yes	45	8	8	100	Yes	79	8	8	100	Yes
13	8	8	100	Yes	5	8	8	100	Yes	8	8	8	100	Yes
15	8	8	100	Yes	50	8	8	100	Yes	84	8	8	100	Yes
16	8	8	100	Yes	51	8	8	100	Yes	85	8	8	100	Yes
17	8	8	100	Yes	52	8	8	100	Yes	86	8	8	100	Yes
18	8	8	100	Yes	53	8	8	100	Yes	9	8	8	100	Yes
20	8	8	100	Yes	54	8	8	100	Yes	10	8	7	87.5	Yes
21	8	8	100	Yes	55	8	8	100	Yes	19	8	7	87.5	Yes
22	8	8	100	Yes	56	8	8	100	Yes	2	8	7	87.5	Yes
23	8	8	100	Yes	57	8	8	100	Yes	26	8	7	87.5	Yes
24	8	8	100	Yes	58	8	8	100	Yes	46	8	7	87.5	Yes
25	8	8	100	Yes						7	8	7	87.5	Yes
27	8	8	100	Yes	59	8	8	100	Yes	71	8	7	87.5	Yes
28	8	8	100	Yes	6	8	8	100	Yes	75	8	7	87.5	Yes
29	8	8	100	Yes	60	8	8	100	Yes	80	8	7	87.5	Yes
3	8	8	100	Yes	61	8	8	100	Yes	81	8	7	87.5	Yes
30	8	8	100	Yes	63	8	8	100	Yes	83	8	7	87.5	Yes
31	8	8	100	Yes	64	8	8	100	Yes	14	8	6	75	Yes
32	8	8	100	Yes	65	8	8	100	Yes	68	8	6	75	Yes
33	8	8	100	Yes	67	8	8	100	Yes	77	8	6	75	Yes
34	8	8	100	Yes	70	8	8	100	Yes	82	8	6	75	Yes
35	8	8	100	Yes	72	8	8	100	Yes	43	8	5	62.5	No
36	8	8	100	Yes	73	8	8	100	Yes	47	8	5	62.5	No
37	8	8	100	Yes	74	8	8	100	Yes	62	8	5	62.5	No
4	8	8	100	Yes	76	8	8	100	Yes	90	8	4	50	No

**ANNEX XI: Qualitative results IPI202 by Analyst and Measurand**

Analyst code	Alexandrium pacificum	Prorocentrum rhathymum	Actinoptychu s splendens	Coolia sp.	Chaetoceros peruvianus	Guinardia striata	Lampriscus sp.	Synedropsis sp.	Overall Flag	Number of Measurands correct
51	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
47	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
37	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
80	not detected	Correct	Correct	Correct	Correct	Correct	Correct	Incorrect	Pass	6
9	Correct	Correct	Correct	Correct	Correct	Correct	Incorrect	Correct	Pass	7
82	Correct	Correct	Correct	not detected	Correct	Correct	Correct	Correct	Pass	7
5	Correct	Correct	Correct	Incorrect	Correct	Correct	Correct	Correct	Pass	7
46	Correct	Correct	Correct	Incorrect	Correct	Correct	Correct	Correct	Pass	7
6	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
65	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
63	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
60	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
20	Correct	Correct	Correct	Incorrect	Correct	Correct	Incorrect	Correct	Pass	6
70	Correct	Correct	Correct	Correct	Correct	Correct	Incorrect	Correct	Pass	8
61	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Incorrect	Pass	8
81	not detected	Correct	Correct	Correct	Correct	Correct	Correct	Incorrect	Pass	6
8	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
84	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
85	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
10	Correct	Correct	incorrect	Correct	Correct	Correct	Correct	Correct	Pass	7
66	nr	nr	nr	nr	nr	nr	nr	nr	nr	nr
77	Correct	Correct	Correct	not detected	Correct	Correct	Correct	Correct	Pass	7
54	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
33	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
72	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
13	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Incorrect	Pass	7
73	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
36	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
45	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
57	Correct	Correct	Correct	Correct	Correct	Correct	Incorrect	Correct	Pass	7
79	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
38	nr	nr	nr	nr	nr	nr	nr	nr	nr	nr
29	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
27	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
4	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
28	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
55	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
21	Correct	Correct	Correct	Incorrect	Correct	Correct	Incorrect	Correct	Pass	6
34	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
83	not detected	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	7
7	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Incorrect	Pass	7
56	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
50	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
71	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
53	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8

## ANNEX XI: Qualitative results IPI2022 by Analyst and Measurand

Analyst code	Alexandrium pacificum	Prorocentrum rhathymum	Actinoptychu splendens	Coolia sp.	Chaetoceros peruvianus	Guinardia striata	Lampriscus sp.	Synedropsis sp.	Overall Flag	Number of Measurands correct
90	Correct	no detected	Correct	not detected	Correct	Correct	Correct	Correct	Pass	6
35	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
62	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
19	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
14	Correct	Correct	incorrect	not detected	Correct	Correct	Correct	Correct	Pass	6
18	Correct	Correct	Correct	Correct	Correct	Correct	Incorrect	Incorrect	Pass	6
24	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
44	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
59	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
2	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
58	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
22	Correct	Correct	Correct	Incorrect	Correct	Correct	Incorrect	Correct	Pass	6
17	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
16	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Incorrect	Pass	7
41	nr	nr	nr	nr	nr	nr	nr	nr	nr	nr
74	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
75	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
76	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
43	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
15	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
52	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
31	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
23	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
67	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
32	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
11	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
86	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
40	nr	nr	nr	nr	nr	nr	nr	nr	nr	nr
69	nr	nr	nr	nr	nr	nr	nr	nr	nr	nr
64	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
30	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
25	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
78	Correct	Correct	Correct	Correct	Correct	Correct	Incorrect	Correct	Pass	7
3	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
26	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
1	Correct	Correct	Correct	Correct	Correct	Incorrect	Correct	Correct	Pass	7
39	nr	nr	nr	nr	nr	nr	nr	nr	nr	nr
42	nr	nr	nr	nr	nr	nr	nr	nr	nr	nr
68	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Correct	Pass	8
<b>Total Correct</b>	74	76	74	68	77	76	43	71	<b>559</b>	
<b>Total Incorrect</b>	0	0	3	5	0	1	34	6	<b>49</b>	
Non-Detected	3	1	0	4	0	0	0	0	<b>8</b>	
No results	7	7	7	7	7	7	7	7	<b>56</b>	

## ANNEX XII: Summary of laboratory means + statistical parameters

A. Code	A. splendens		Synedropsis sp.		Lampriscus sp.		G. striata		C. peruvianus		P. rhathymum		C. monotis		A. pacificum	
	Cells/L	Z-score	Cells/L	Z-score	Cells/L	Z-score	Cells/L	Z-score	Cells/L	Z-score	Cells/L	Z-score	Cells/L	Z-score	Cells/L	Z-score
1	1500	0.2	3133	-0.4	1633	0.3	6433	0	13633	-0.4	4400	-0.6	5233	0.3	2500	-0.5
2	1040	-0.5	3440	-0.1	1320	-0.3	6067	-0.1	21068	1	10772	3.2	7552	2	3400	0.4
3	667	-1	2760	-0.9	1560	0.1	6600	0.1	16560	0.2	7533	1.3	5120	0.2	4747	1.8
4	1320	-0.1	3813	0.4	1280	-0.4	8613	1.1	19560	0.7	6160	0.5	5053	0.2	3480	0.5
5	1653	0.4	3987	0.6	1453	-0.1	6453	0.1	19760	0.7	5947	0.4	4707	-0.1	3307	0.3
6	1067	-0.4	3560	0.1	1280	-0.4	8280	0.9	19240	0.6	5133	-0.1	4933	0.1	3387	0.4
7	1120	-0.4	3507	0	1600	0.2	10813	2.1	17853	0.4	5773	0.2	6280	1.1	3293	0.3
8	1707	0.5	3813	0.4	1773	0.6	5173	-0.6	19000	0.6	5733	0.2	6160	1	2453	-0.6
9	2360	1.4	3840	0.4	2080	1.2	4827	-0.7	18333	0.5	6680	0.8	7413	1.9	2760	-0.3
10	3320	2.8	4133	0.7	1600	0.2	7467	0.5	17973	0.4	7320	1.2	5080	0.2	3707	0.7
11	1553	0.3	3227	-0.3	1507	0	6553	0.1	19187	0.6	7107	1	5760	0.7	3433	0.4
13	933	-0.6	2400	-1.3	1520	0.1	6233	-0.1	10133	-1	3213	-1.3	3473	-0.9	2167	-0.9
14	873	-0.7	2427	-1.2	1013	-0.9	5993	-0.2	14540	-0.2	4987	-0.2		3	6373	3.5
15	1667	0.4	3387	-0.1	1400	-0.2	6720	0.2	16920	0.2	5387	0	5827	0.7	2493	-0.5
16	813	-0.8	3160	-0.4	1267	-0.4	5000	-0.6	19000	0.6	5747	0.2	4253	-0.4	3227	0.2
17	960	-0.6	2853	-0.8	1240	-0.5	8827	1.2	20280	0.8	5733	0.2	5400	0.4	3427	0.4
18	2000	0.9	3333	-0.2	1333	-0.3	7333	0.5	19000	0.6	4000	-0.8	4000	-0.6	3333	0.3
19	1613	0.3	2653	-1	1547	0.1	2493	-1.8	2600	-2.4	5227	-0.1	4080	-0.5	2413	-0.6
20	1520	0.2	3453	-0.1	1573	0.2	7160	0.4	19627	0.7	5307	0	4227	-0.4	3547	0.6
21	1027	-0.5	2493	-1.2	1427	-0.1	8640	1.1	18293	0.5	5173	-0.1	4027	-0.6	3520	0.5
22	1827	0.7	3120	-0.4	1440	-0.1	5347	-0.5	12293	-0.6	5027	-0.2	4173	-0.4	3107	0.1
23	1613	0.3	4387	1	1640	0.3	3533	-1.3	22213	1.2	5267	-0.1	4387	-0.3	2720	-0.3
24	1233	-0.2	4233	0.8	1733	0.5	5567	-0.4	22933	1.3	6833	0.9	5967	0.8	2967	0
25	784	-0.9	3504	0	1502	0	7403	0.5	20177	0.8	7083	1	6110	0.9	4797	1.9
26	1360	0	3653	0.2	1333	-0.3	6320	0	18067	0.4	6560	0.7	4307	-0.4	5093	2.2
27	1973	0.9	4053	0.6	1733	0.5	5267	-0.5	15373	-0.1	6547	0.7	5613	0.6	3653	0.7
28	653	-1.1	3520	0	1160	-0.6	5413	-0.4	17600	0.3	5880	0.3	5587	0.6	3627	0.6
29	840	-0.8	3453	-0.1	1427	-0.1	6973	0.3	15120	-0.1	5653	0.2	4253	-0.4	3813	0.8
30	1714	0.5	3176	-0.4	1809	0.6	5320	-0.5	12956	-0.5	6485	0.7	5242	0.3	3092	0.1
31	1033	-0.5	3279	-0.3	1301	-0.4	6947	0.3	16700	0.2	6285	0.6	5679	0.6	3776	0.8
32	1480	0.2	3427	-0.1	1107	-0.7	5320	-0.5	16307	0.1	5093	-0.2	5253	0.3	3173	0.2
33	1307	-0.1	2320	-1.4	1160	-0.6	6427	0	14840	-0.2	4733	-0.4	3573	-0.9	2520	-0.5
34	1773	0.6	3093	-0.5	1547	0.1	6130	-0.1	24217	1.6	7946	1.6	6660	1.3	2876	-0.1
35	1255	-0.2	3202	-0.3	1262	-0.4	6341	0	7003	-1.6	4848	-0.3	5236	0.3	2582	-0.4
36	1118	-0.4	3160	-0.4	1528	0.1	6857	0.2	19286	0.7	3762	-1	4383	-0.3	2829	-0.2
37	1875	0.7	3583	0.1	1694	0.4	6389	0	17333	0.3	4667	-0.4	4292	-0.4	2889	-0.1
43	1267	-0.2	2307	-1.4	693	-1.6	1533	-2.3	2787	-2.4	4307	-0.6	2760	-1.5	253	-2.9
44	1213	-0.2	3787	0.3	1067	-0.8	3720	-1.2	6907	-1.6	3413	-1.2	3333	-1	1973	-1.1
45	1133	-0.4	3040	-0.5	1320	-0.3	5453	-0.4	8173	-1.4	5107	-0.2	4387	-0.3	2747	-0.3
46	987	-0.6	2213	-1.5	1067	-0.8	6080	-0.1	11920	-0.7	4053	-0.8	1720	-2.2	3307	0.3
47	680	-1	1680	-2.1	1173	-0.6	3840	-1.2	1187	-2.7	3453	-1.1	4440	-0.3	200	-2.9
50	1812	0.6	3681	0.2	1855	0.7	7232	0.4	21146	1	6174	0.5	4594	-0.1	2725	-0.3
51	1841	0.7	4565	1.2	1768	0.6	7131	0.4	20682	0.9	4536	-0.5	4725	-0.1	2536	-0.5
52	1348	0	3971	0.5	1927	0.9	6797	0.2	19189	0.6	5537	0.1	4754	0	2565	-0.5
53	1907	0.8	3813	0.4	1920	0.9	8787	1.2	16307	0.1	5373	0	4773	0	3653	0.7
54	1920	0.8	4533	1.2	1840	0.7	6493	0.1	11867	-0.7	4280	-0.6	4373	-0.3	1880	-1.2
55	960	-0.6	3427	-0.1	1147	-0.7	5307	-0.5	19573	0.7	4600	-0.5	3280	-1.1	2547	-0.5
56	1160	-0.3	3213	-0.3	1827	0.7	6000	-0.2	16493	0.1	5053	-0.2	6387	1.1	2947	-0.1
57	1233	-0.2	2267	-1.4	1433	-0.1	6767	0.2	10667	-0.9	3700	-1	3400	-1	2633	-0.4
58	2613	1.8	3293	-0.2	1813	0.6	6413	0	15793	0	5933	0.3	4973	0.1	2240	-0.8
59	1373	0	3293	-0.2	1520	0.1	8347	1	25224	1.7	7431	1.2	6696	1.4	3145	0.1
60	1813	0.6	4507	1.2	1813	0.6	7987	0.8	22754	1.3	5467	0.1	5080	0.2	3907	0.9
61	1284	-0.1	4272	0.9	1531	0.1	6296	0	21654	1.1	7284	1.2	5654	0.6	2593	-0.4
62	1333	-0.1	3067	-0.5	1700	0.4	7000	0.3	7333	-1.5	1733	-2.2	1233	-2.6	533	-2.6
63	840	-0.8	2507	-1.2	1293	-0.4	7560	0.6	11667	-0.7	5600	0.1	4120	-0.5	2800	-0.2
64	2733	2	4653	1.3	2253	1.5	5773	-0.3	16827	0.2	6067	0.4	6453	1.2	2947	-0.1
65	1067	-0.4	3920	0.5	1280	-0.4	3987	-1.1	13387	-0.4	4080	-0.8	2933	-1.3	3627	0.6
67	1072	-0.4	3760	0.3	1496	0	5847	-0.2	15200	-0.1	2537	-1.7	3920	-0.6	2289	-0.8
68	1609	0.3	5307	2.1	1720	0.5	10148	1.8	20419	0.9	10148	2.9	7054	1.6	4331	1.4
70	1307	-0.1	3120	-0.4	1467	0	4213	-1	11693	-0.7	4613	-0.4	3600	-0.9	2947	-0.1
71	1453	0.1	4787	1.5	1360	-0.2	5773	-0.3	11467	-0.8	2773	-1.6	1120	-2.6	1947	-1.1
72	680	-1	3013	-0.6	1253	-0.5	2880	-1.6	10053	-1	4720	-0.4	4840	0	3187	0.2
73	1467	0.1	3627	0.1	1587	0.2	7107	0.4	17133	0.3	6000	0.4	5400	0.4	2600	-0.4
74	853	-0.8	4227	0.8	1693	0.4	5640	-0.3	8947	-1.2	6120	0.5	3760	-0.7	2960	-0.1
75	2253	1.3	4280	0.9	1760	0.5	8093	0.8	14507	-0.2	6720	0.8	4187	-0.4	5107	2.2
76	1307	-0.1	4147	0.7	1280	-0.4	5853	-0.2	16653	0.2	7373	1.2	5387	0.4	2867	-0.2
77	1880	0.7	3333	-0.2	1587	0.2	3373	-1.4	5920	-1.8	3160	-1.3		3	5600	2.7
78	1947	0.8	3173	-0.4	1693	0.4	7440	0.5	11773	-0.7	6147	0.5	3653	-0.8	3120	0.1
79	1733	0.5	4187	0.8	1827	0.7	5813	-0.2	10787	-0.9	4773	-0.4	4360	-0.3	2200	-0.8
80	667	-1	3800	0.3	1267	-0.4	6533	0.1	10533	-0.9	3667	-1	5067	0.2		3
81	1533	0.2	4400	1	933	-1.1	6100	-0.1	16000	0.1	4300	-0.6	4200	-0.4		3
82	1480	0.2	3693	0.2	1600	0.2	6573	0.1	10907	-0.9	7089	1		3	7246	4.4
83	773	-0.9	3980	0.6	1313	-0.3	5053	-0.6	11820	-0.7	3273	-1.3	7113	1.7		3
84	1667	0.4	3435	-0.1	748	-1.5	9082	1.3	20374	0.9	3980	-0.8	5340	0.4	3197	0.2
85	947	-0.6	3757	0.3	1437	-0.1	6566	0.1	15223	-0.1	3267	-1.3	4671	-0.1	1470	-1.6
86	2573	1.7	4213	0.8	2027	1.1	7873	0.7	20573	0.9	7747	1.4	6800	1.4	3173	0.2
90	1073	-0.4	2053	-1.7	987	-1	2080	-2	167	-2.9		3		3	2307	-0.7

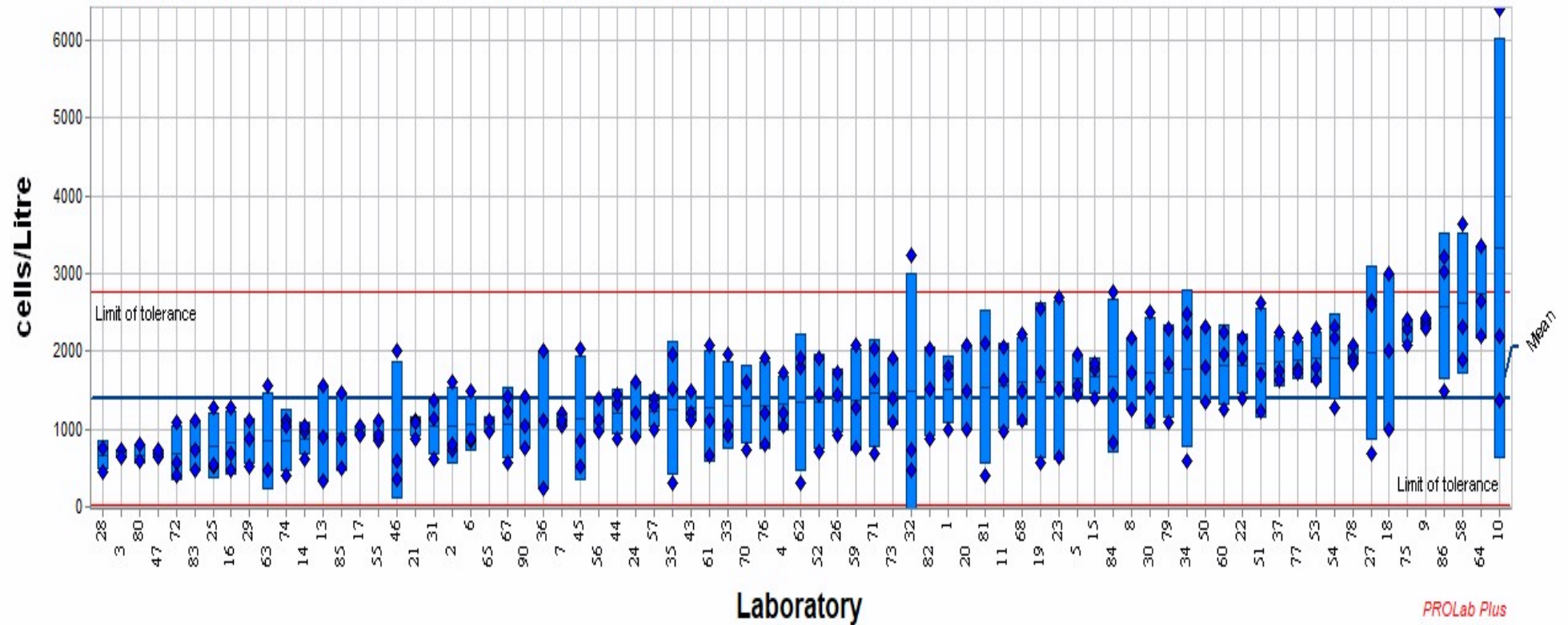
## ANNEX XII: Summary of laboratory means + statistical parameters

Statistical method	Q/Hampel	Q/Hampel	Q/Hampel	Q/Hampel	Q/Hampel	Q/Hampel	Q/Hampel	Q/Hampel	Q/Hampel
Assessment	Z <=2.0	Z <=2.0	Z <=2.0	Z <=2.0	Z <=2.0	Z <=2.0	Z <=2.0	Z <=2.0	Z <=2.0
No. of laboratories that submitted results	77	77	77	77	77	77	76	73	74
No. of participants	77	77	77	77	77	77	77	77	77
Assigned value	1376	3501	1485	6339	15703	5361	4797	3011	3011
Mean	1400	3493	1482	6275	15587	5361	4829	3013	3013
Reference value	1376	3501	1485	6339	15703	5361	4797	3011	3011
SDPA	688	862	505	2109	5445	1666	1394	955	955
Reproducibility s.d.	688	862	505	2109	5445	1666	1394	955	955
Repeatability s.d.	524	612	421	1302	1631	859	711	437	437
Rel. SDPA	50.0 %	24.6 %	34.0 %	33.3 %	34.7 %	31.1 %	29.1 %	31.7 %	31.7 %
Rel. reproducibility s.d.	50.0 %	24.6 %	34.0 %	33.3 %	34.7 %	31.1 %	29.1 %	31.7 %	31.7 %
Rel. repeatability s.d.	38.1 %	17.5 %	28.3 %	20.5 %	10.4 %	16.0 %	14.8 %	14.5 %	14.5 %
Reference s.d.	644	626	395	2112	5312	1574	1115	725	725
Limit of reproducibility, R (2.80 X sR)	1926	2414	1415	5906	15246	4666	3904	2674	2674
Limit of repeatability, r (2.80 X sr)	1468	1714	1178	3646	4567	2406	1990	1224	1224
Rel. limit of reproducibility	140.0 %	68.9 %	95.3 %	93.2 %	97.1 %	87.0 %	81.4 %	88.8 %	88.8 %
Rel. limit of repeatability	106.7 %	48.9 %	79.3 %	57.5 %	29.1 %	44.9 %	41.5 %	40.7 %	40.7 %
HORRAT	74.2	42	51.1	62.1	74.2	56.6	52	53	53
Absolute classical Horwitz s.d.	9	21	10	34	73	29	27	18	18
Relative classical Horwitz s.d.	0.7 %	0.6 %	0.7 %	0.5 %	0.5 %	0.5 %	0.6 %	0.6 %	0.6 %
Lower limit of tolerance	0	1777	475	2120	4813	2028	2008	1101	1101
Upper limit of tolerance	2752	5225	2495	10558	26593	8694	7586	4921	4921
Standard error	61	80	42	208	602	173	148	103	103
No. of laboratories	77	77	77	77	77	76	73	74	74
No. of measurement values and states	231	231	231	231	231	228	219	222	222
No. of measurement values	231	231	231	231	231	228	219	222	222
No. of measurement values without outliers	231	231	231	231	231	228	219	222	222



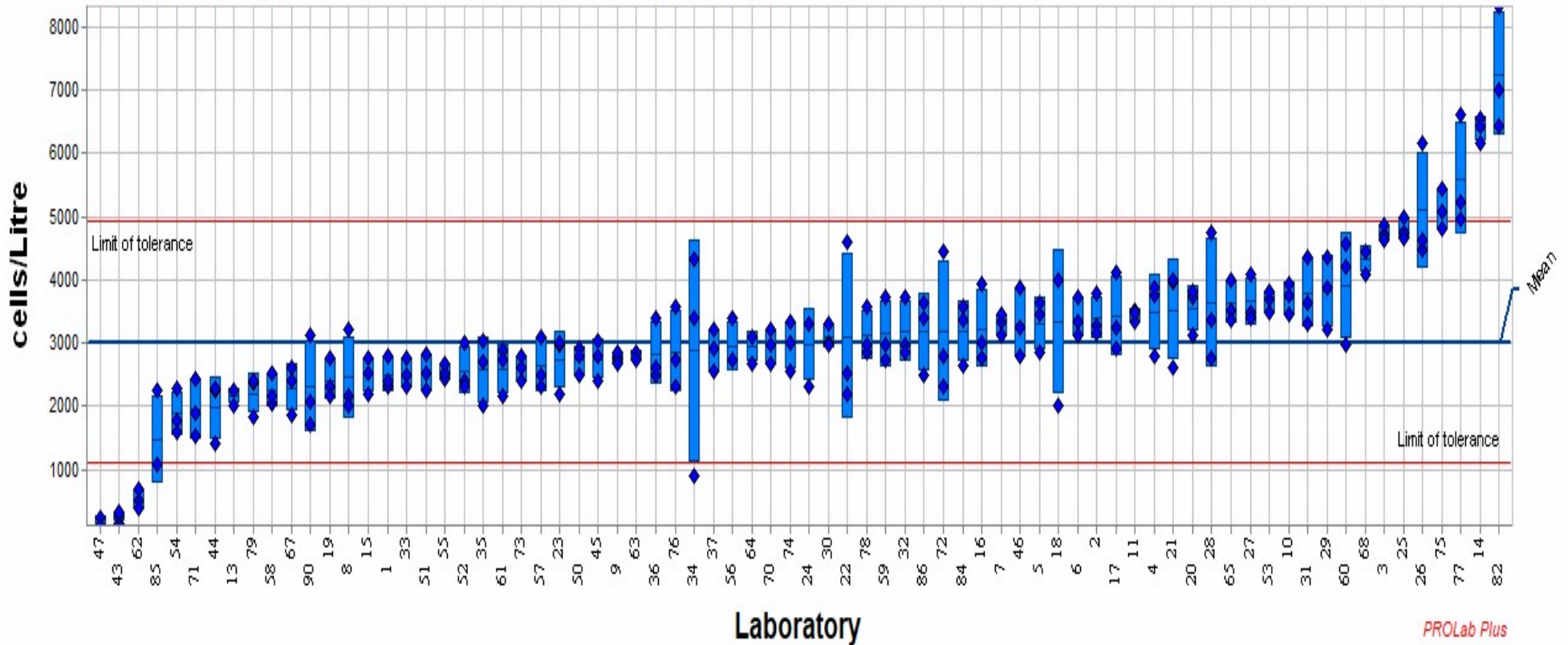
### ANNEX XIII: Graphical summary of *Actinoptychus splendens* results by analyst

Sample: 022  
 Measurand: Actinoptychus splendens  
 Statistical method: Q/Hampel  
 Number of laboratories in calculation: 77  
 Assigned value: 1376 cells/Litre (Reference value)  
 Rel. SDPA: 50.0% (Empirical value)  
 Rel. repeatability s.d.: 38.1%  
 Range of tolerance: 0 - 2752 cells/Litre ( $|Z\text{-Score}| \leq 2.0$ )



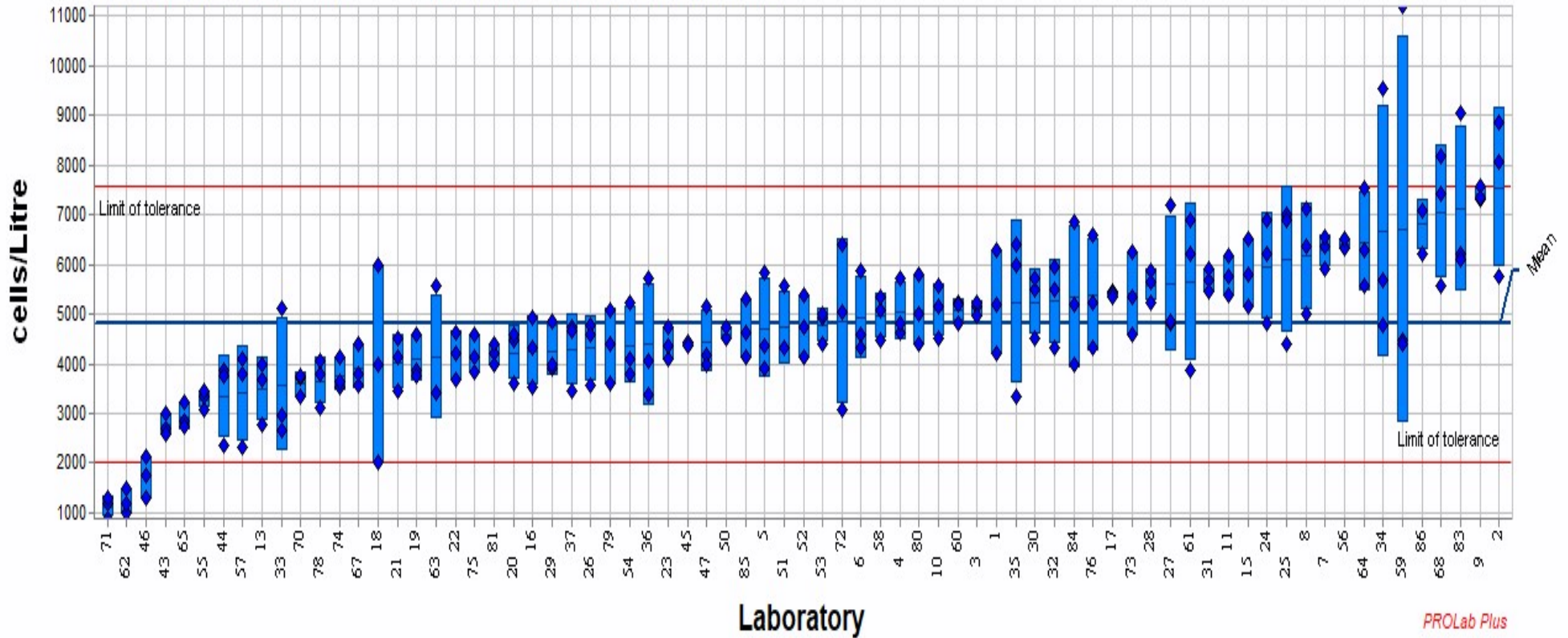
### ANNEX XIII: Graphical summary of *Alexabdrum pacificum* results by analyst

Sample: 022  
 Measurand: Alexandrium pacificum  
 Statistical method: Q/Hampel  
 Number of laboratories in calculation: 74  
 Assigned value: 3011 cells/Litre (Reference value)  
 Rel. SDPA: 31.7% (Empirical value)  
 Rel. repeatability s.d.: 14.5%  
 Range of tolerance: 1101 - 4921 cells/Litre (|Z-Score| <= 2.0)



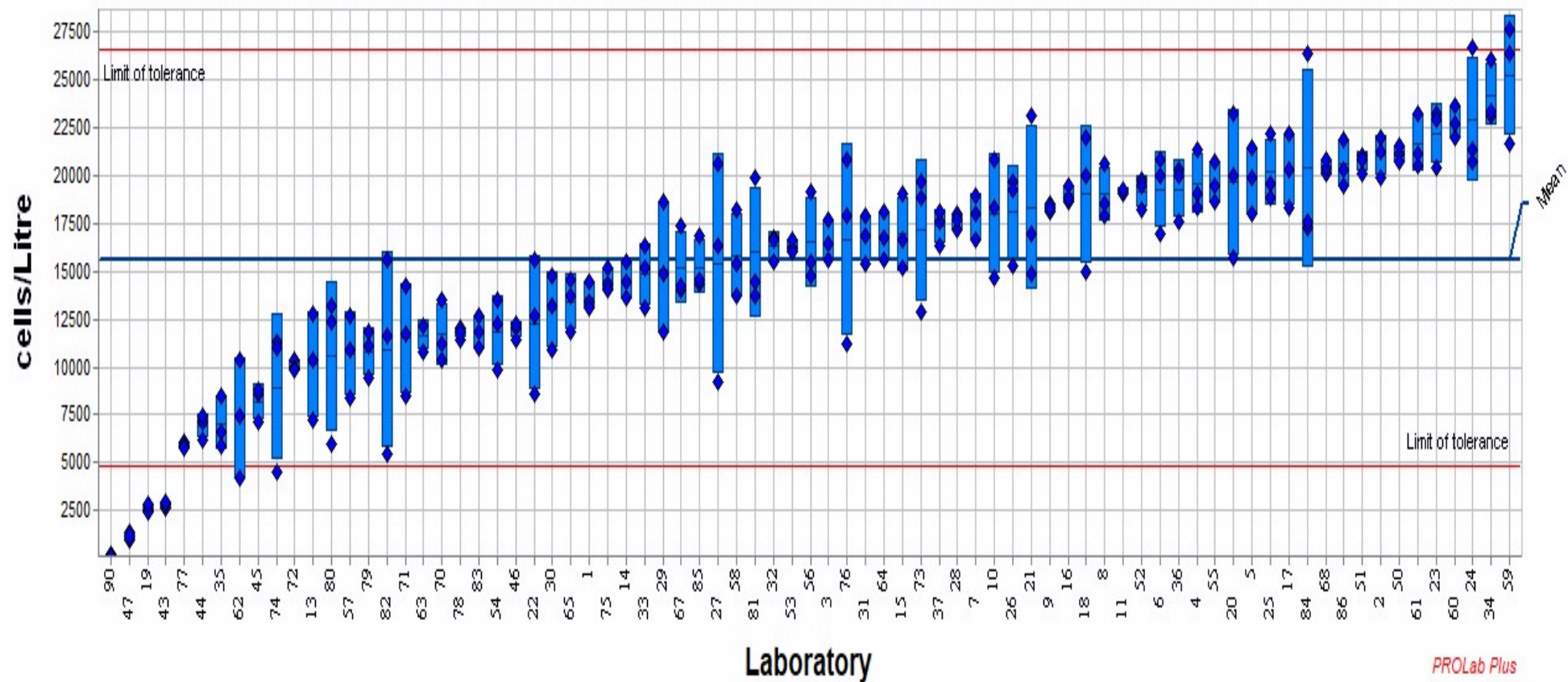
ANNEX XIII: Graphical summary of *Coolia monotis* results by analyst

Sample: 022 Assigned value: 4797 cells/Litre (Reference value)  
 Measurand: *Coolia monotis* Rel. SDPA: 29.1% (Empirical value)  
 Statistical method: Q/Hampel Rel. repeatability s.d.: 14.8%  
 Number of laboratories in calculation: 73 Range of tolerance: 2008 - 7586 cells/Litre ( $|Z\text{-Score}| \leq 2.0$ )



### ANNEX XIII: Graphical summary of *Chaetoceros peruvianus* results by analyst

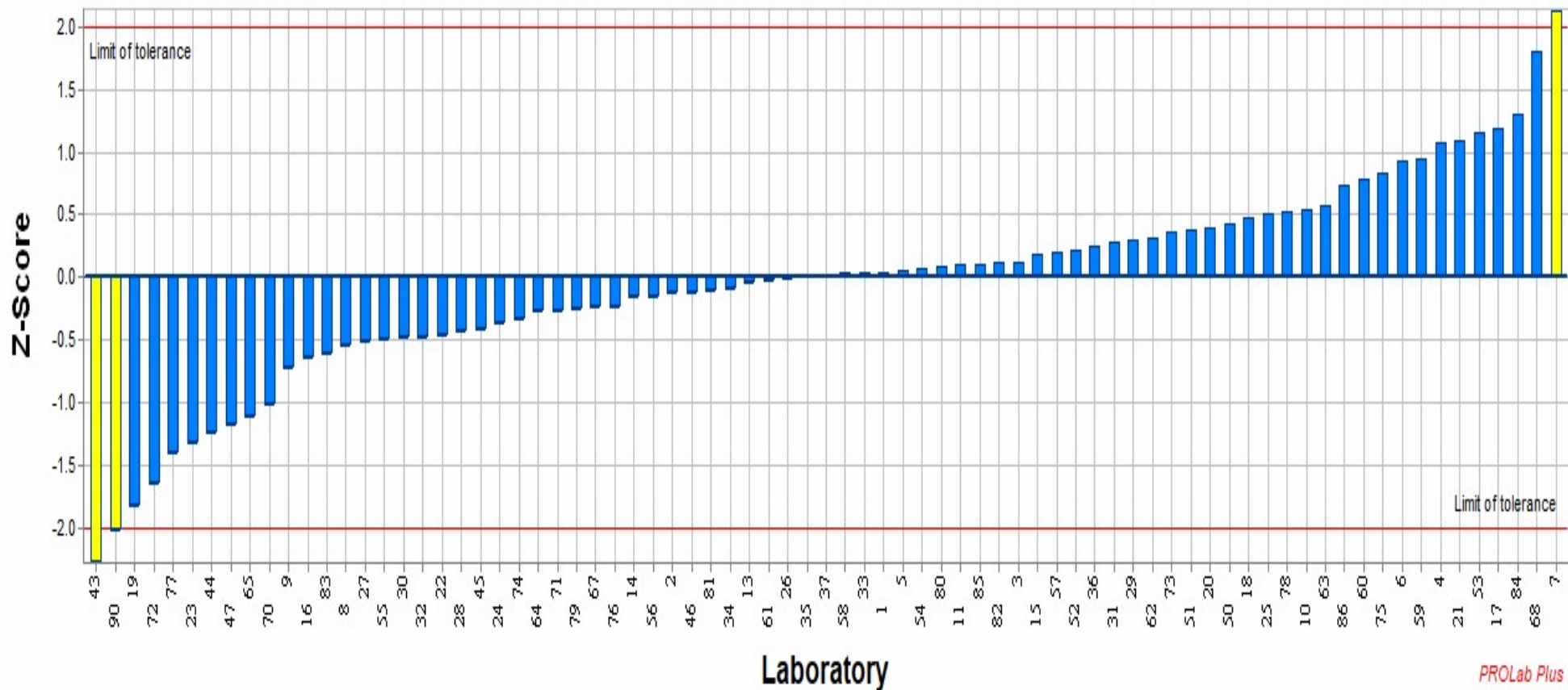
Sample: 022  
 Measurand: *Chaetoceros peruvianus*  
 Statistical method: Q/Hampel  
 Number of laboratories in calculation: 77  
 Assigned value: 15703 cells/Litre (Reference value)  
 Rel. SDPA: 34.7% (Empirical value)  
 Rel. repeatability s.d.: 10.4%  
 Range of tolerance: 4813 - 26593 cells/Litre (|Z-Score| <= 2.0)





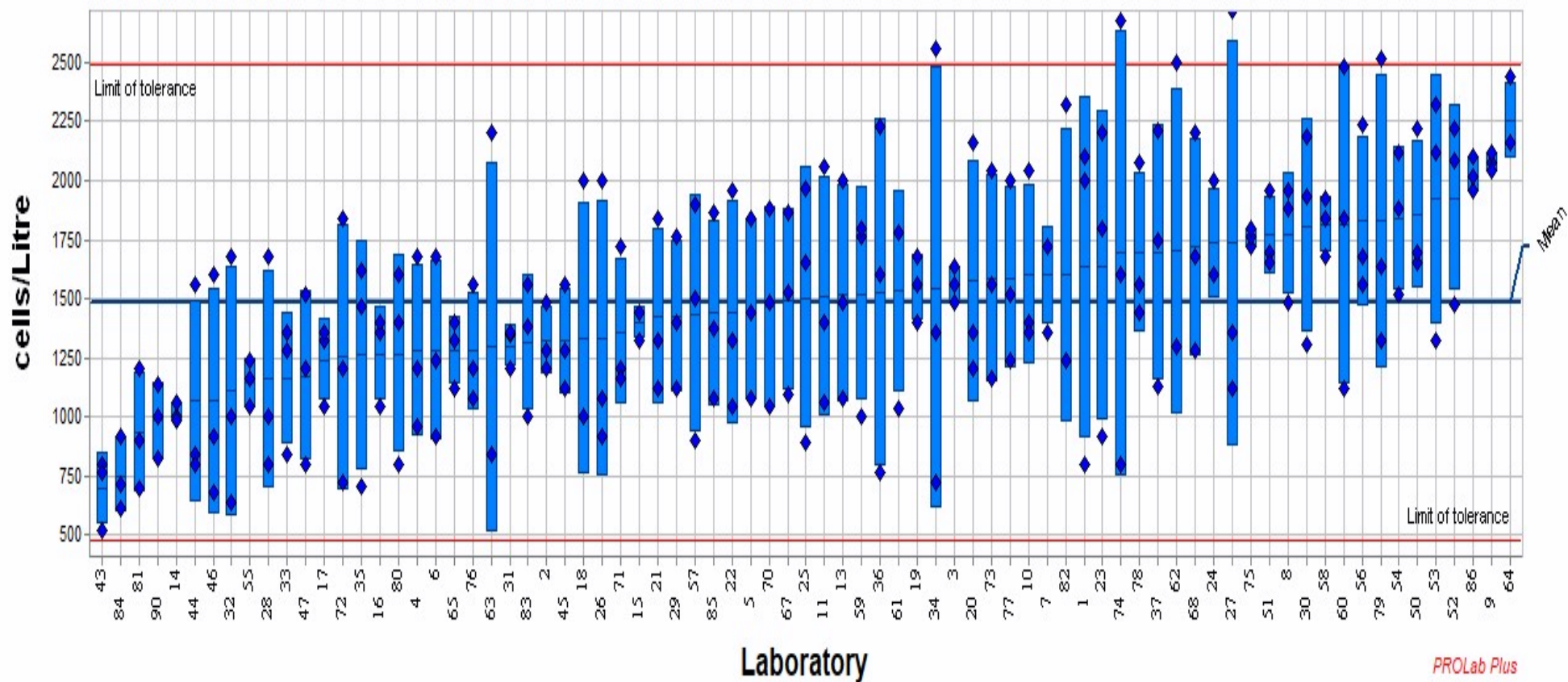
### ANNEX XIII: Graphical summary of *Guinardia striata* results by analyst

Sample: 022 Assigned value: 6339 cells/Litre (Reference value)  
 Measurand: *Guinardia striata* Rel. SDPA: 33.3% (Empirical value)  
 Statistical method: Q/Hampel Rel. repeatability s.d.: 20.5%  
 Number of laboratories in calculation: 77 Range of tolerance: 2120 - 10558 cells/Litre (Z-Score) <= 2.0



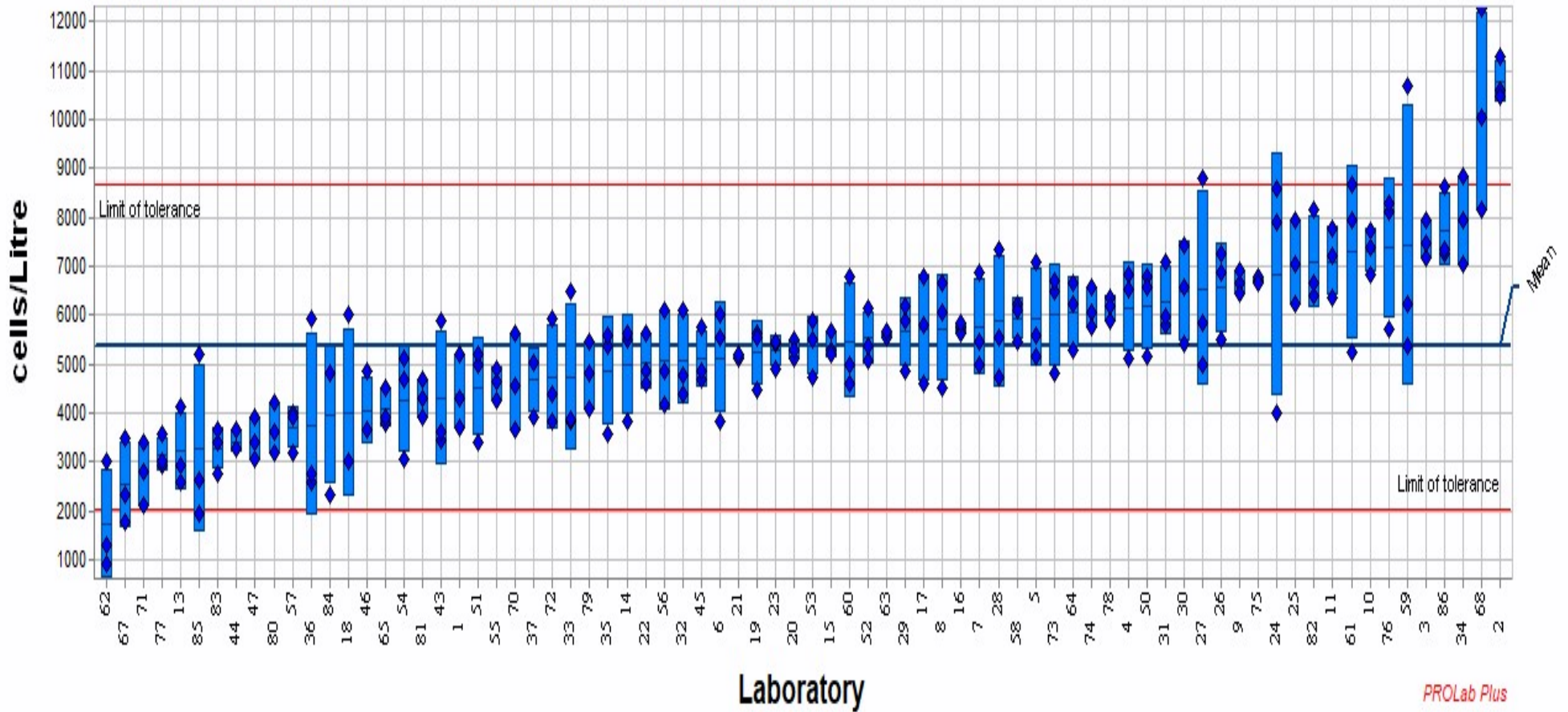
### ANNEX XIII: Graphical summary of *Lampriscus sp.* results by analyst

Sample: 022 Assigned value: 1485 cells/Litre (Reference value)  
 Measurand: Lampriscus sp. Rel. SDPA: 34.0% (Empirical value)  
 Statistical method: Q/Hampel Rel. repeatability s.d.: 28.3%  
 Number of laboratories in calculation: 77 Range of tolerance: 475 - 2495 cells/Litre (|Z-Score| <= 2.0)



ANNEX XIII: Graphical summary of *Prorocentrum rathymum* results by analyst

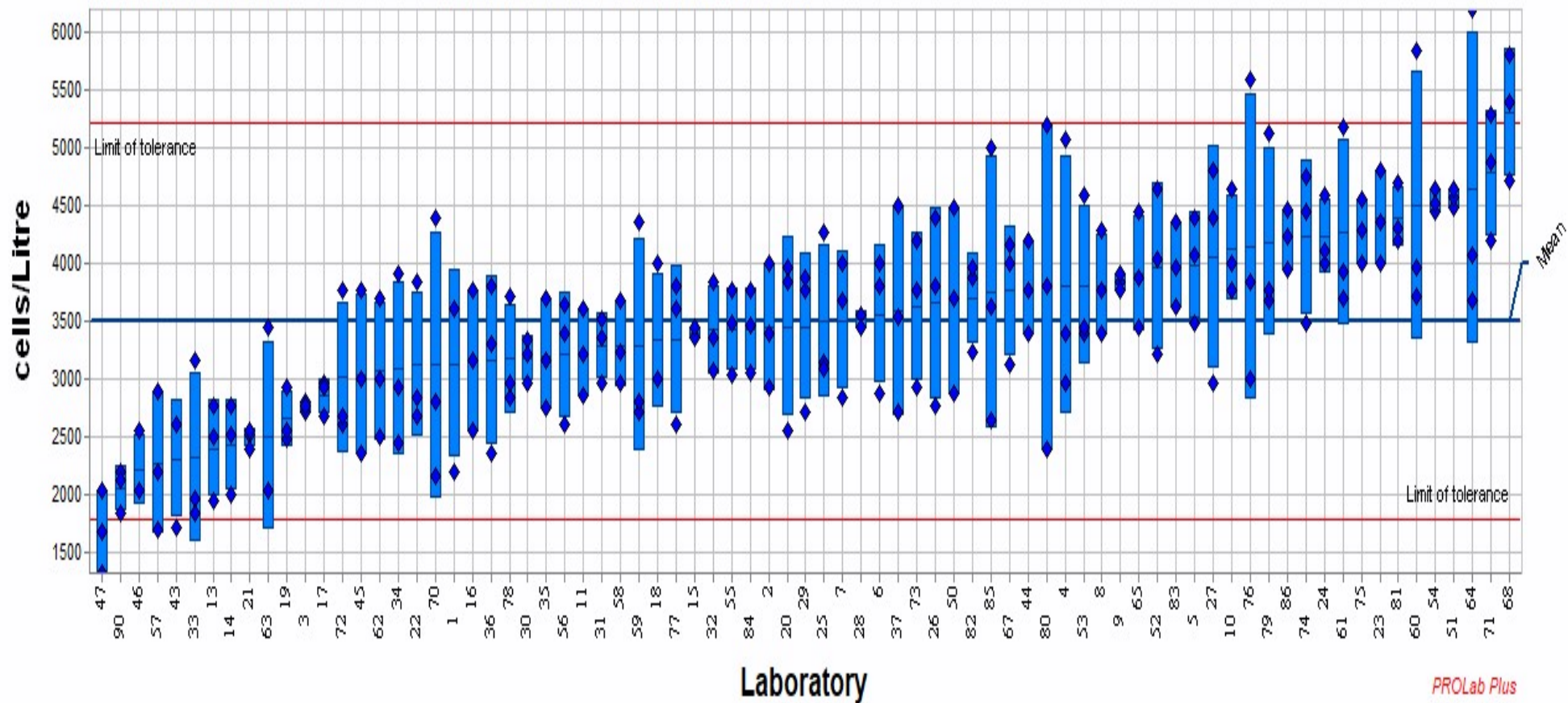
Sample: 022 Assigned value: 5361 cells/Litre (Reference value)  
 Measurand: Prorocentrum rathymum Rel. SDPA: 31.1% (Empirical value)  
 Statistical method: Q/Hampel Rel. repeatability s.d.: 16.0%  
 Number of laboratories in calculation: 76 Range of tolerance: 2028 - 8694 cells/Litre (|Z-Score| <= 2.0)



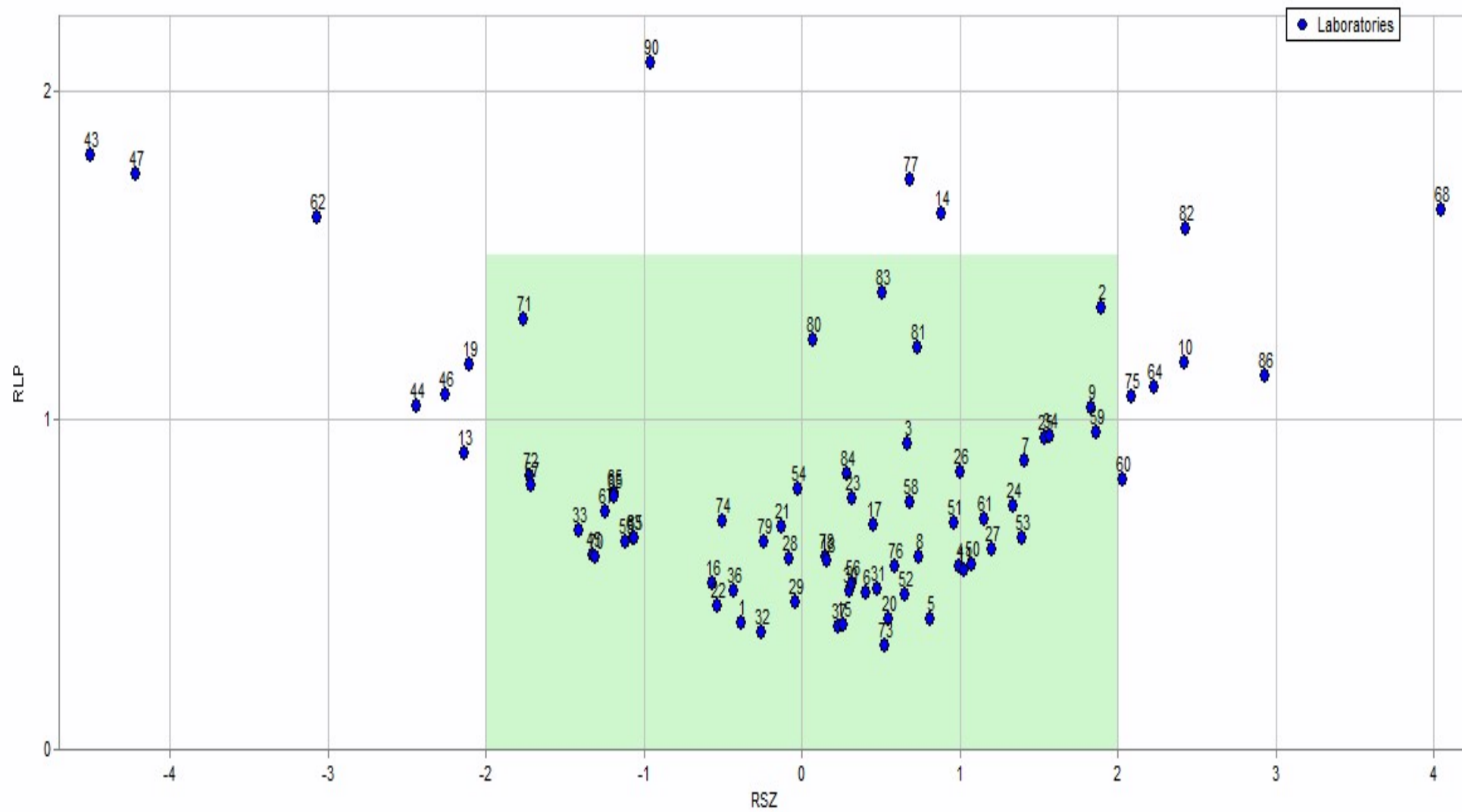


ANNEX XIII: Graphical summary of *Synedropsis sp.* results by analyst

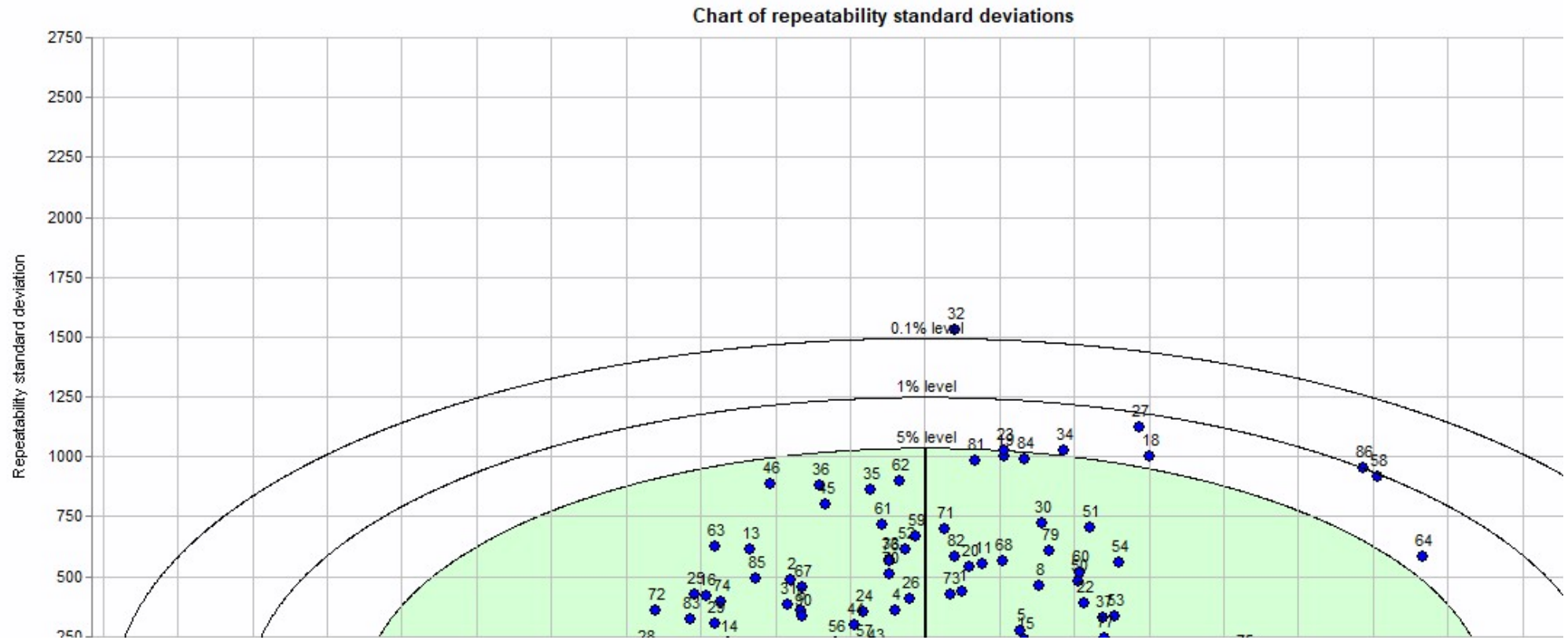
Sample: 022 Assigned value: 3501 cells/Litre (Reference value)  
 Measurand: *Synedropsis sp.* Rel. SDPA: 24.6% (Empirical value)  
 Statistical method: Q/Hampel Rel. repeatability s.d.: 17.5%  
 Number of laboratories in calculation: 77 Range of tolerance: 1777 - 5225 cells/Litre (|Z-Score| <= 2.0)



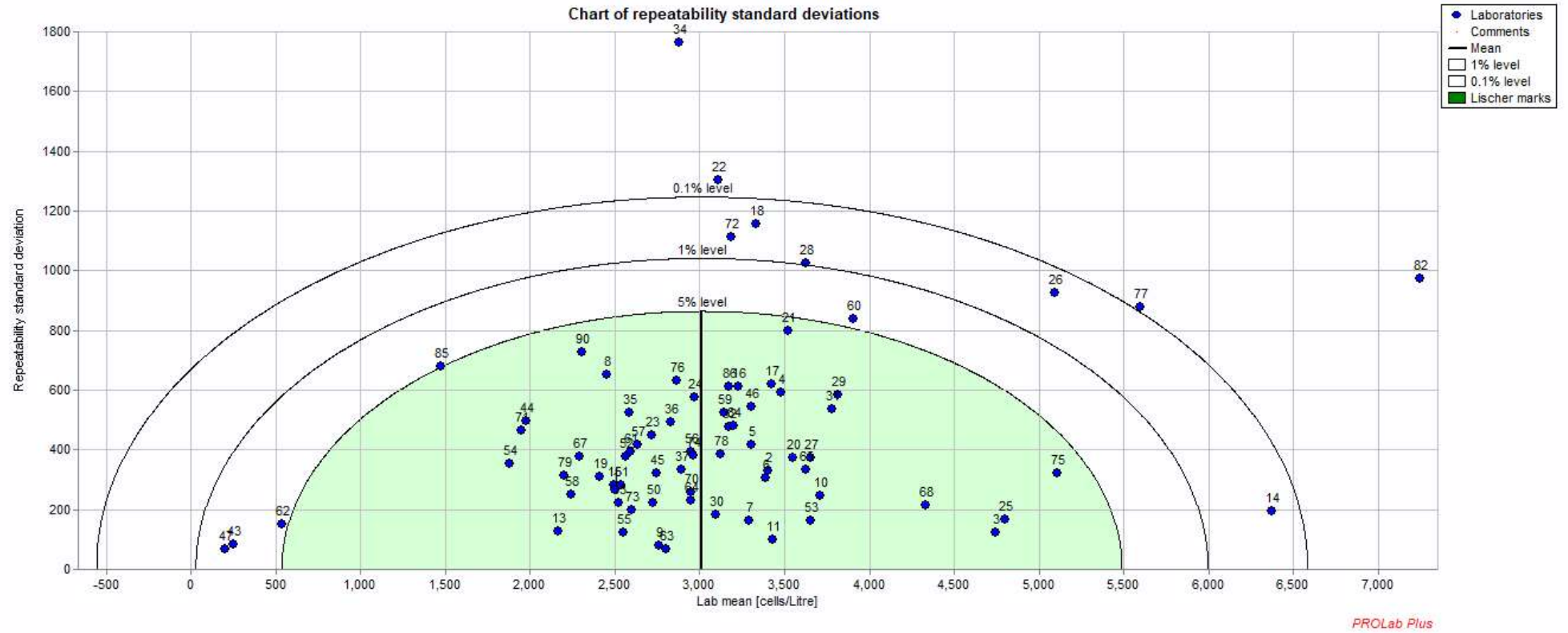
ANNEX XIV: RLP and RSZ for all measurands IPI2022



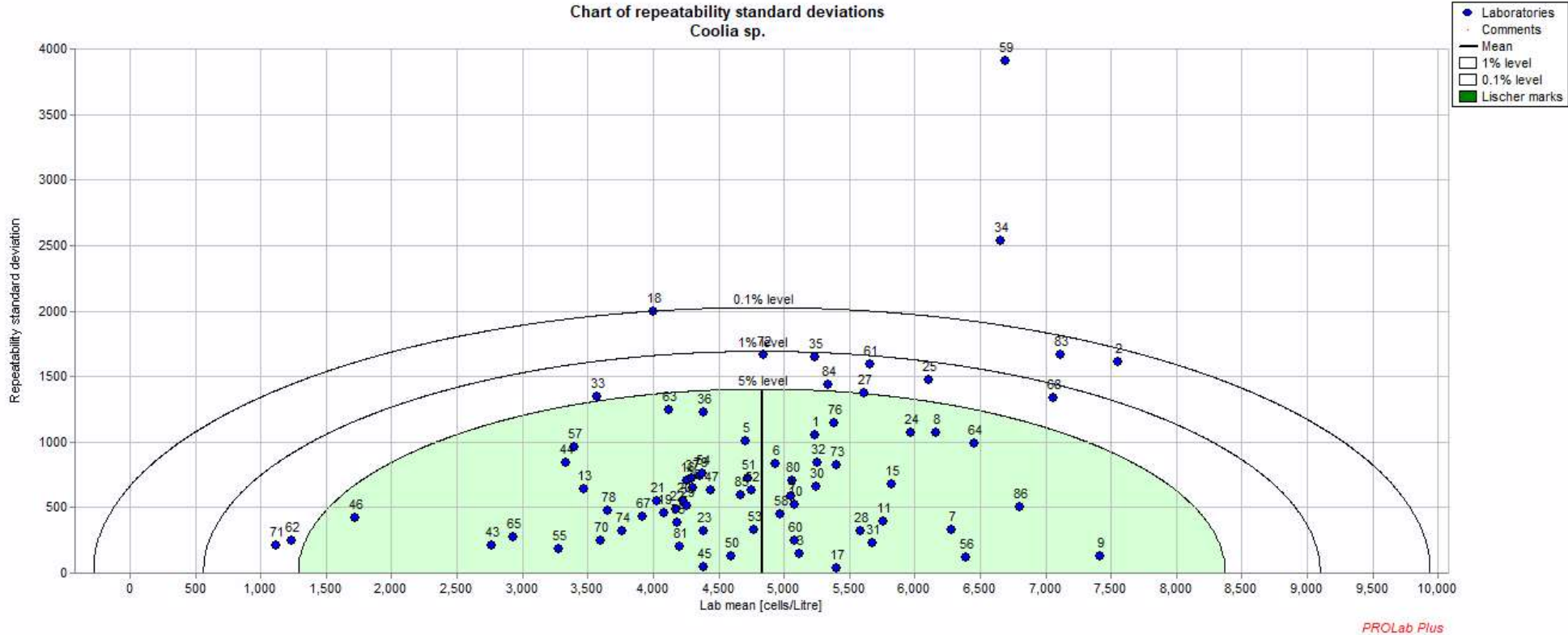
ANNEX XV: Lischer plot *Actinoptychus splendens*



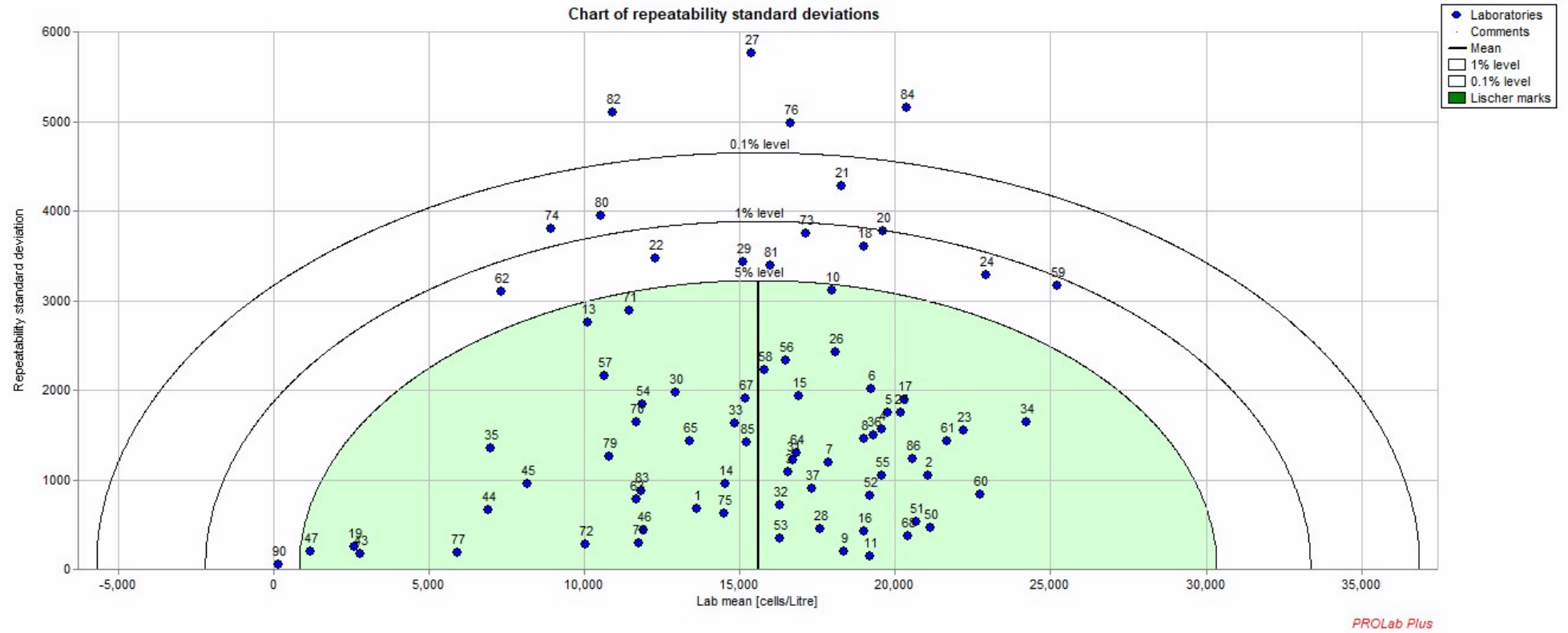
# ANNEX XV: Lischer plot *Alexandrium pacificum*



ANNEX XV: Lischer plot *Coolia monotis*

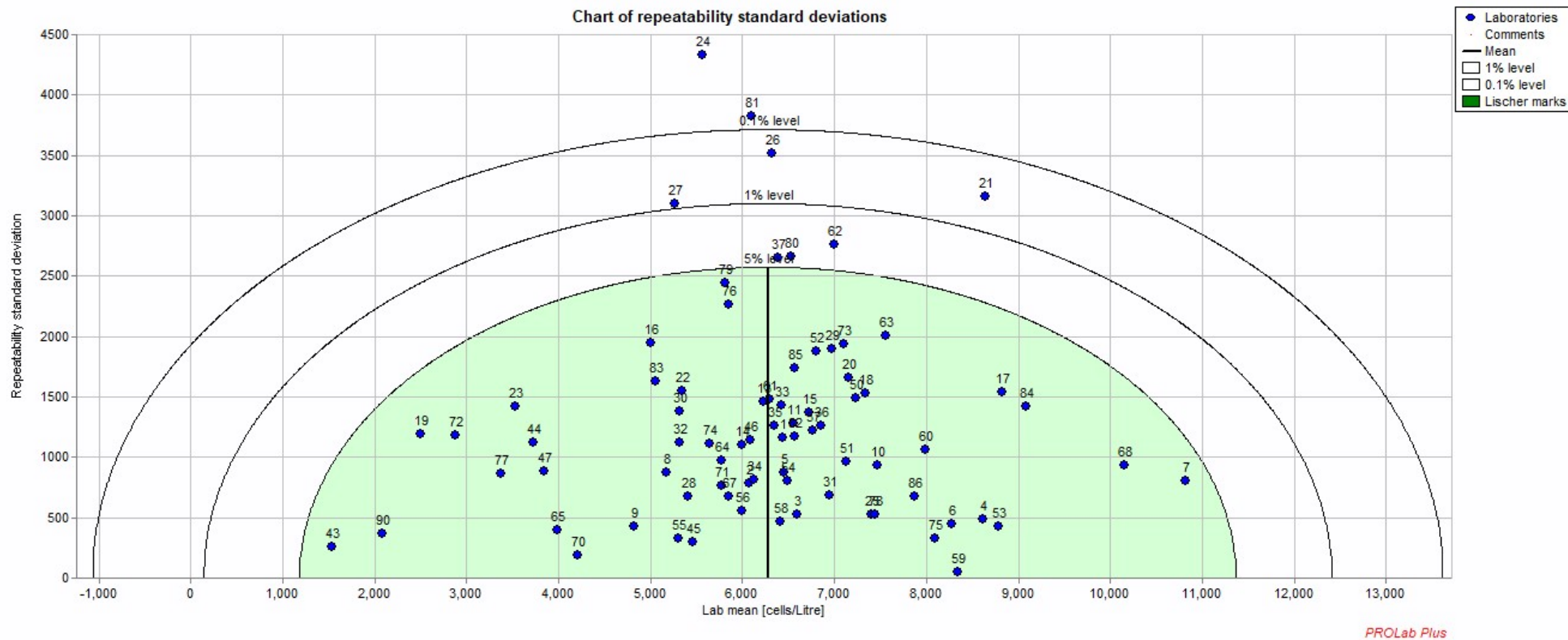


# ANNEX XV: Lischer plot *Chaetoceros peruvianus*



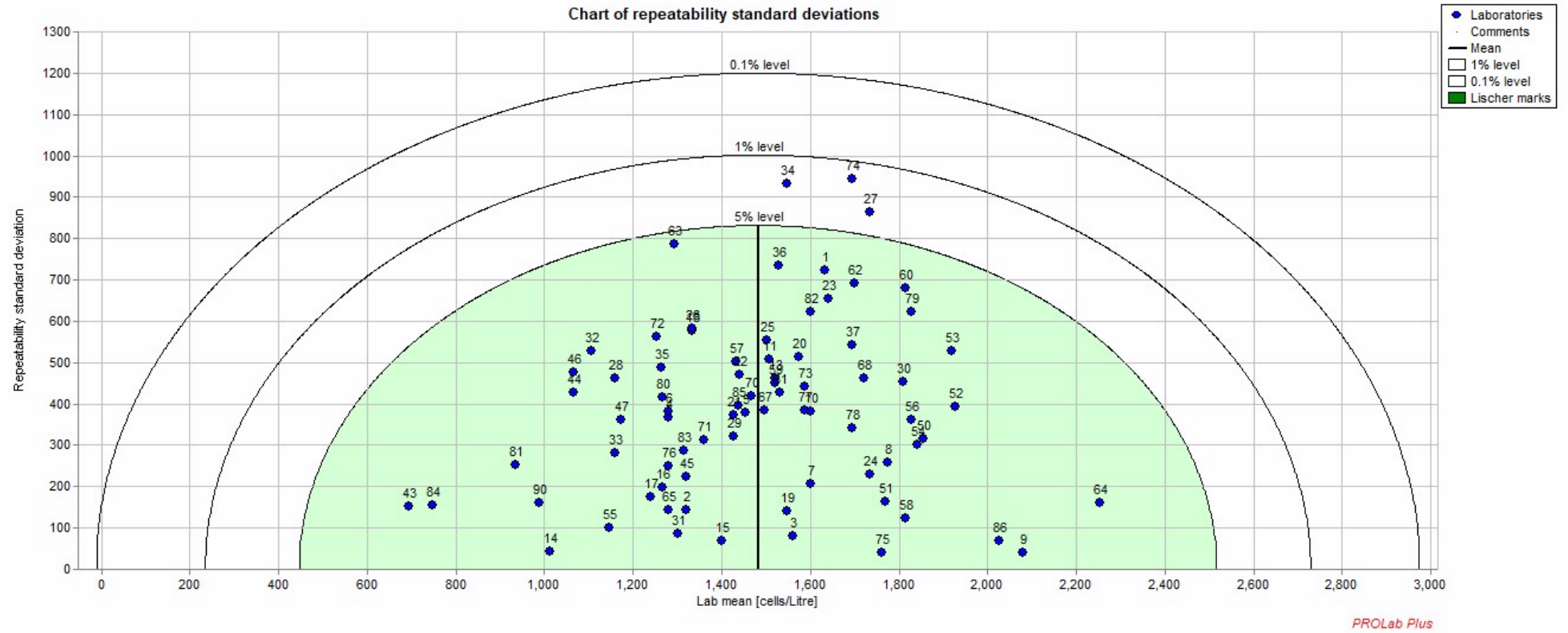


### ANNEX XV: Lischer plot *Guinardia striata*

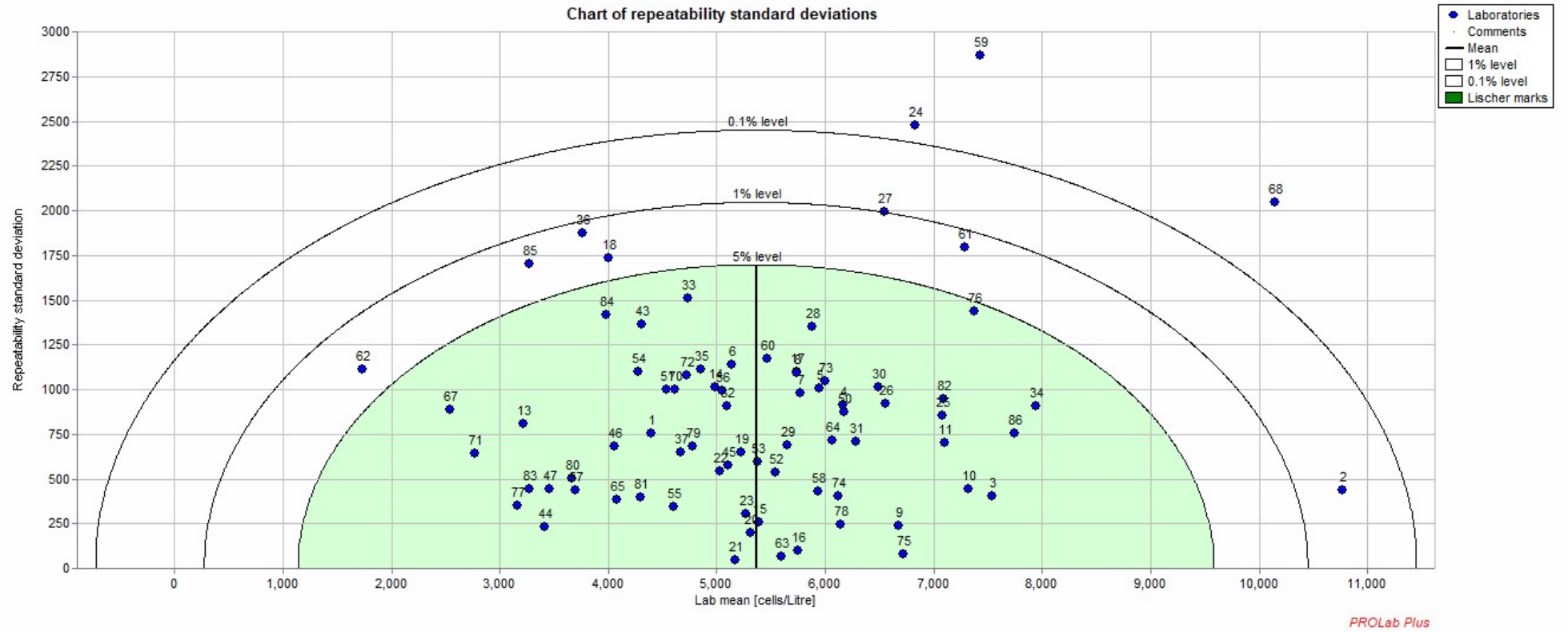




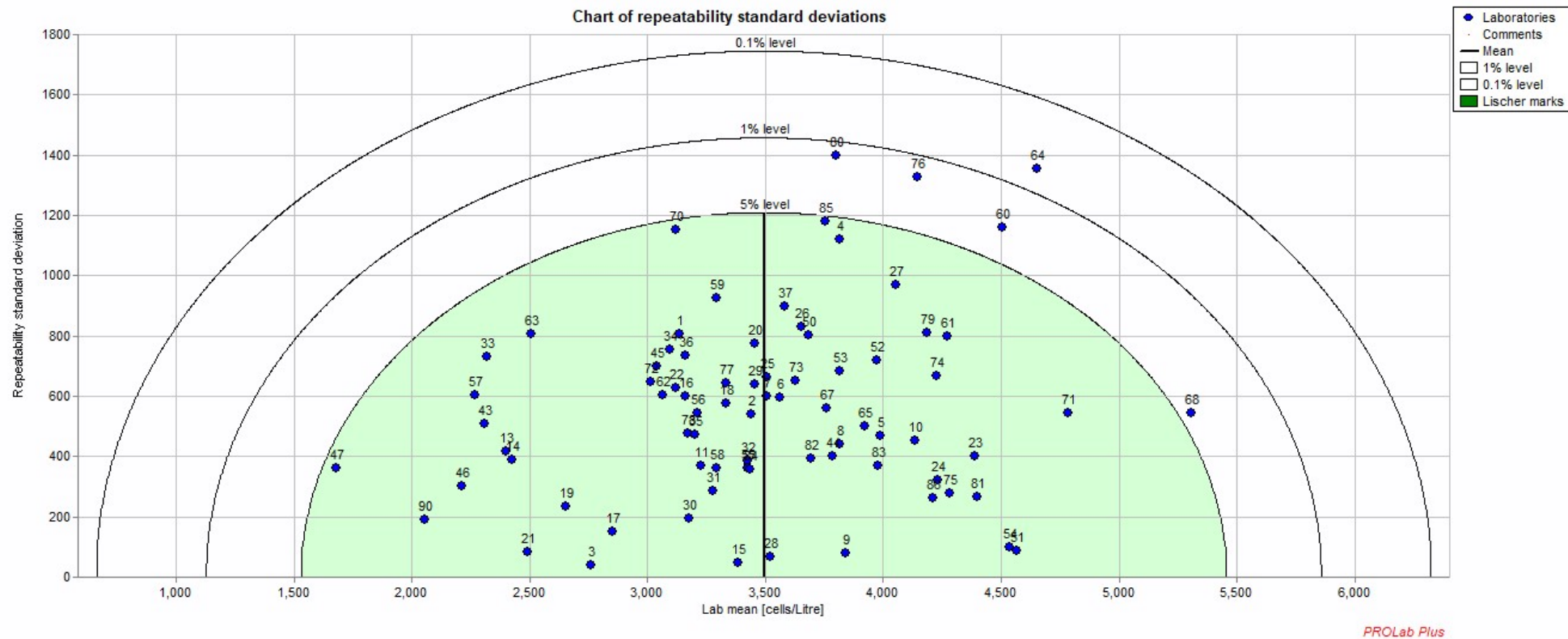
# ANNEX XV: Lischer plot *lampriscus sp.*



# ANNEX XV: Lischer plot *Prorocentrum rhathymum*



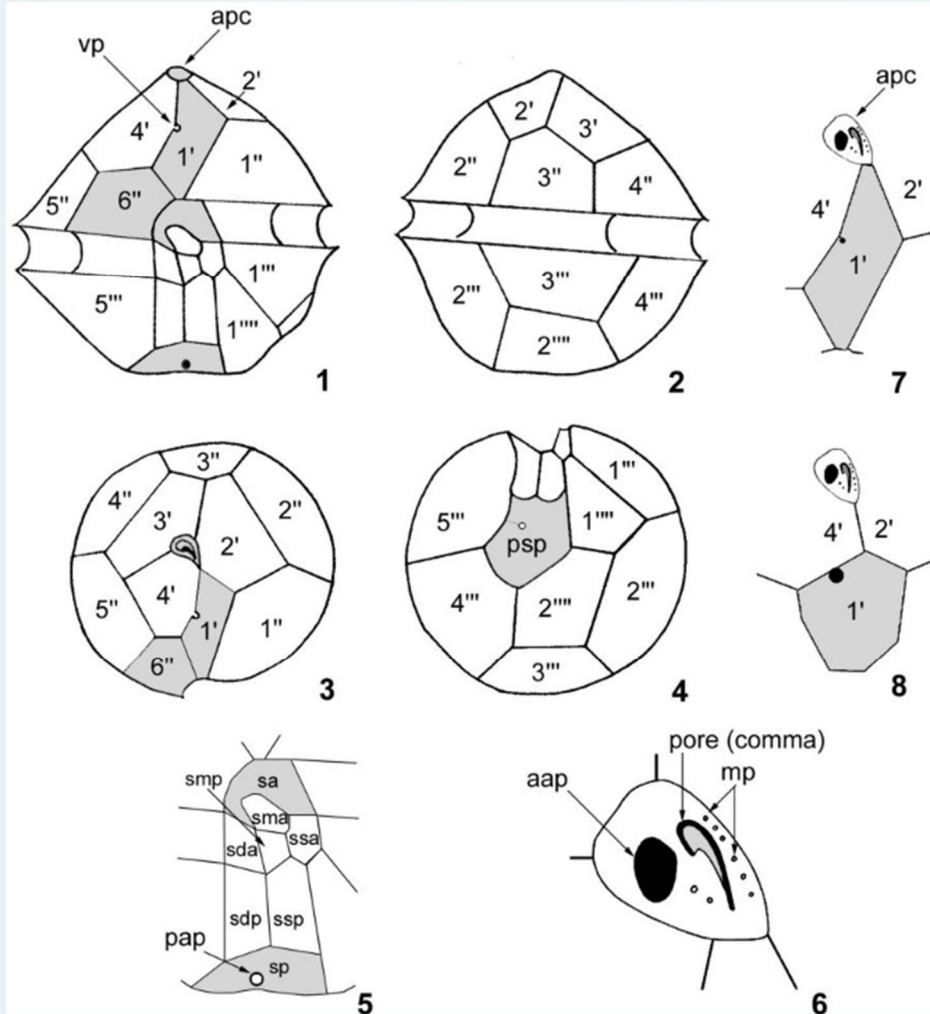
### ANNEX XV: Lischer plot *Synedropsis sp.*



ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q1

Question 1  
Not yet answered  
Marked out of 1.00  
Flag question  
Edit question

The following diagram shows the plate tabulation of *Alexandrium*. There are several important diagnostic features to identify and differentiate this important genera from other dinoflagellates. Choose the right answer from the drop down menu.



The vp arrow in image 1 points to

Ventral pore

The APC arrow in images 1 and 7 points to

Apical Pore Complex

The smp arrow in image 5 points to

Sulcal median posterior

The psp greyed out plate in image 4 points to

Posterior sulcal plate

The sa greyed out plate in image 5 points to

Sulcal anterior

The aap arrow in image 6 points to

Anterior attachment pore

The 1' plate greyed out in images 1, 3, 7 & 8 points to

First apical plate

The pap arrow in image 5 points to

Posterior attachment pore

**ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q1 answers**

<b>Q1</b>	<b>Model response</b>	<b>Actual response</b>	<b>Partial credit</b>	<b>Count</b>	<b>Frequency</b>
3016	The APC arrow in images 1 and 7 points to: Apical Pore Complex	Apical Pore Complex	12.50%	83	98.81%
3016	The APC arrow in images 1 and 7 points to: Anterior apical pore	Anterior apical pore	0.00%	1	1.19%
3017	The vp arrow in image 1 points to: Ventral pore	Ventral pore	12.50%	84	100.00%
3018	The pap arrow in image 5 points to: Posterior attachment pore	Posterior attachment pore	12.50%	72	85.71%
3018	The pap arrow in image 5 points to: Posterior sulcal pore	Posterior sulcal pore	0.00%	1	1.19%
3018	The pap arrow in image 5 points to: Posterior apical pore	Posterior apical pore	0.00%	1	1.19%
3018	The pap arrow in image 5 points to: Posterior antapical pore	Posterior antapical pore	0.00%	10	11.90%
3019	The aap arrow in image 6 points to: Anterior attachment pore	Anterior attachment pore	12.50%	78	92.86%
3019	The aap arrow in image 6 points to: Anterior antapical pore	Anterior antapical pore	0.00%	3	3.57%
3019	The aap arrow in image 6 points to: Anterior apical pore	Anterior apical pore	0.00%	3	3.57%
3020	The smp arrow in image 5 points to: Sulcal median posterior	Sulcal median posterior	12.50%	78	92.86%
3020	The smp arrow in image 5 points to: Posterior sulcal plate	Posterior sulcal plate	0.00%	1	1.19%
3020	The smp arrow in image 5 points to: Sulcal median pore	Sulcal median pore	0.00%	1	1.19%
3020	The smp arrow in image 5 points to: Sinister median plate	Sinister median plate	0.00%	4	4.76%
3021	The psp greyed out plate in image 4 points to: Posterior sulcal plate	Posterior sulcal plate	12.50%	79	94.05%
3021	The psp greyed out plate in image 4 points to: Posterior sulcal pore	Posterior sulcal pore	0.00%	5	5.95%
3022	The 1&#039; plate greyed out in images 1, 3, 7 & 8 points to: Ventral pore	Ventral pore	0.00%	1	1.19%
3022	The 1&#039; plate greyed out in images 1, 3, 7 & 8 points to: First apical plate	First apical plate	12.50%	82	97.62%
3022	The 1&#039; plate greyed out in images 1, 3, 7 & 8 points to: First preapical plate	First preapical plate	0.00%	1	1.19%
3023	The sa greyed out plate in image 5 points to: Posterior sulcal plate	Posterior sulcal plate	0.00%	1	1.19%
3023	The sa greyed out plate in image 5 points to: Sulcal anterior	Sulcal anterior	12.50%	78	92.86%
3023	The sa greyed out plate in image 5 points to: Sulcal antapical	Sulcal antapical	0.00%	5	5.95%



ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q2

Question 2

Not yet answered

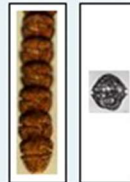
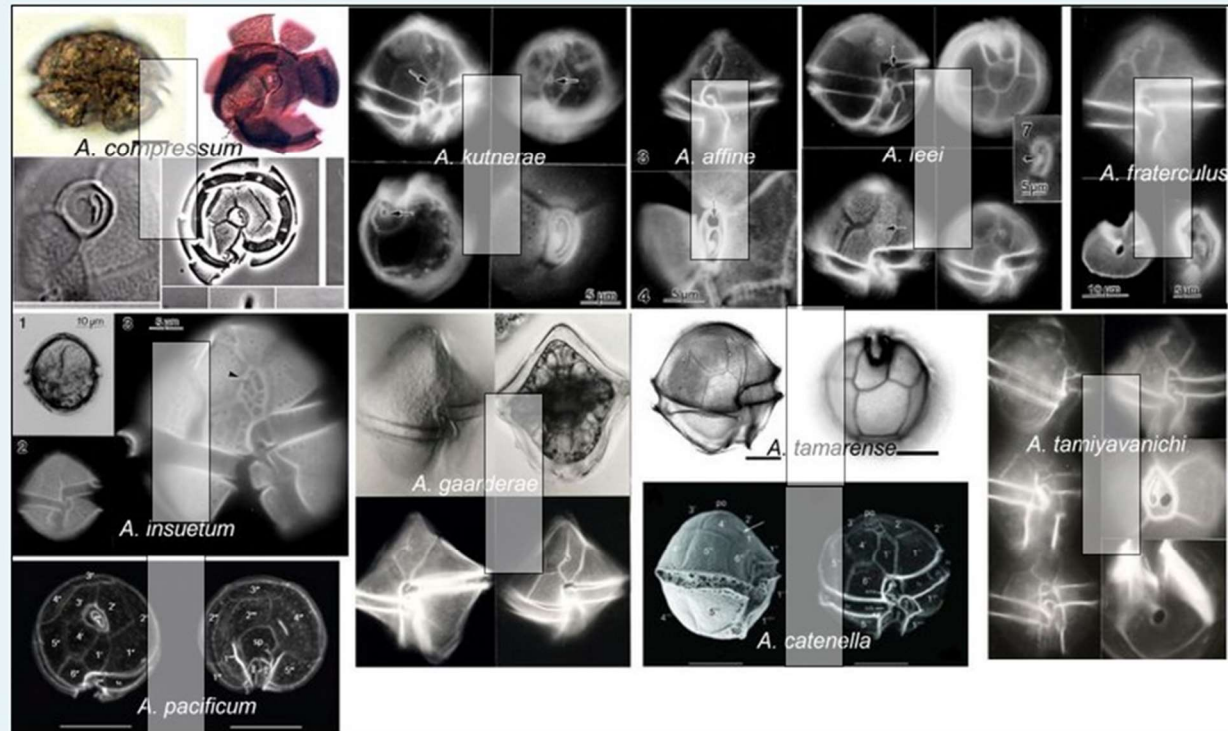
Marked out of 1.00

Flag question

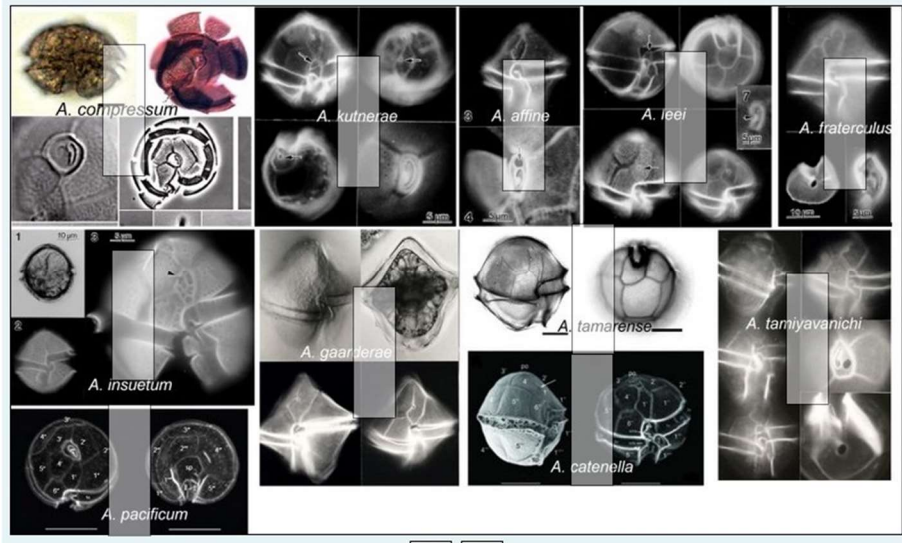
Edit question

The *Alexandrium* genus is known to have several species that produce toxins but also it has the peculiarity that some of its members are able to form large chains.

Drag and drop the chain forming image to the species you consider to be chain-formers and do the same for the non-chain formers.



ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q2 answers



Q2	Species	Model response	Partial credit	Count	Frequency
1	A.compressum	Chain former	100.00%	83	98.81%
1		Non-Chain former	0.00%	1	1.19%
2	A.affine	Chain former	100.00%	84	100.00%
3	A.fraterculus	Chain former	100.00%	81	96.43%
3		Non-Chain former	0.00%	3	3.57%
4	A.tamiyavanichi	Chain former	100.00%	83	98.81%
4		Non-Chain former	0.00%	1	1.19%
5	A.catenella	Chain former	100.00%	81	96.43%
5		Non-Chain former	0.00%	3	3.57%
6		Chain former	0.00%	39	46.43%
6	A.kutnerae	Non-Chain former	100.00%	45	53.57%
7		Chain former	0.00%	3	3.57%
7	A.leei	Non-Chain former	100.00%	81	96.43%
8	A.insuetum	Non-Chain former	100.00%	84	100.00%
9		Chain former	0.00%	1	1.19%
9	A.gaardnerae	Non-Chain former	100.00%	83	98.81%
10		Chain former	0.00%	2	2.38%
10	A.tamarensis	Non-Chain former	100.00%	82	97.62%
11	A.pacificum	Chain former	100.00%	71	84.52%
11		Non-Chain former	0.00%	13	15.48%



ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q3

Question 3

Not yet answered

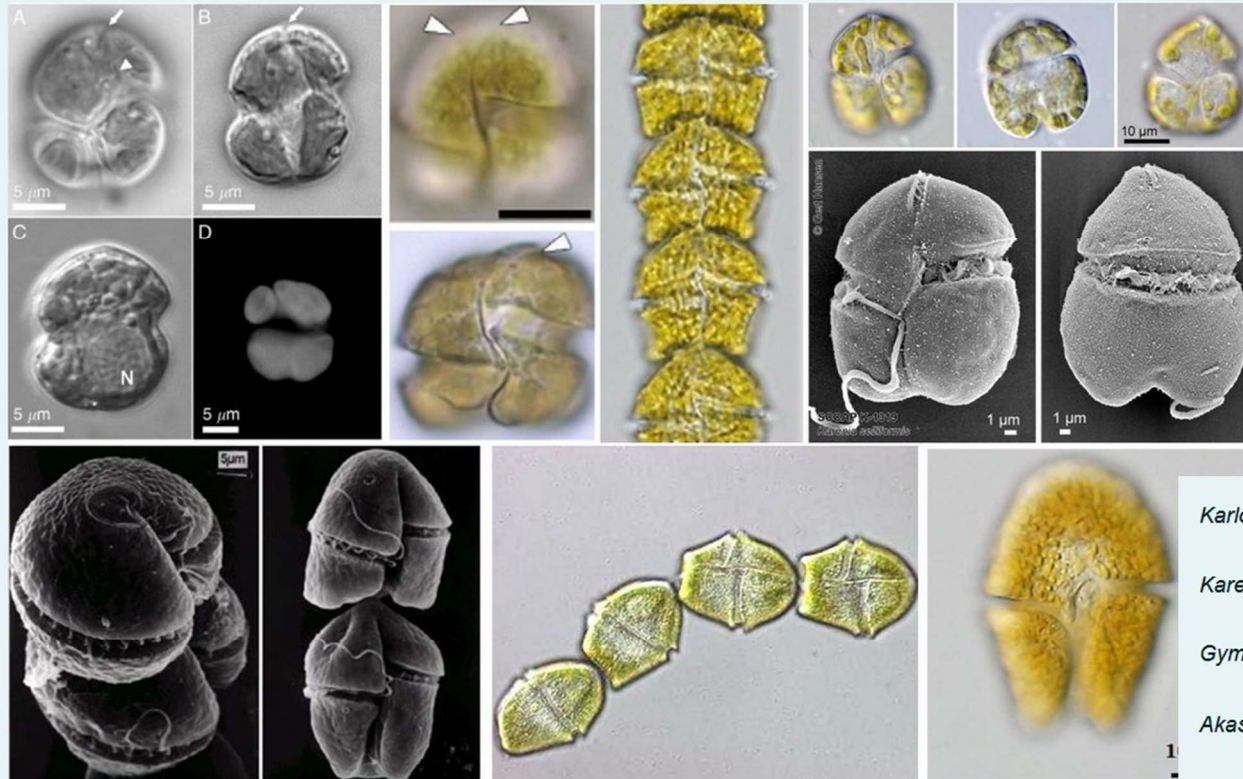
Marked out of 1.00

Flag question

Edit question

The Gymnodiniales is a group of 'naked dinoflagellates' which are generally quite difficult to identify. One important diagnostic feature of this group is the 'Acrobase' otherwise called 'Apical groove'. Some genera can be differentiated based on this feature.

Match the correct statement to each genus



<i>Karlodinium</i>	Apical groove is straight	↕
<i>Karenia</i>	Apical groove is straight	↕
<i>Gymnodinium</i>	Apical groove is horse shoe shaped	↕
<i>Akashiwo</i>	Apical groove goes around the apex of the cell	↕
<i>Takayama</i>	Apical groove is sigmoid	↕

**ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q3 answers**

<b>Q3</b>	<b>Model response</b>	<b>Actual response</b>	<b>Partial credit</b>	<b>Count</b>	<b>Frequency</b>
3037	_Karlodinium_: Apical groove is straight	Apical groove is straight	20.00%	53	63.10%
3037	_Karlodinium_: Apical groove goes around the apex of the cell	Apical groove goes around the apex of the cell	0.00%	1	1.19%
3037	_Karlodinium_: Apical groove descends dorsally	Apical groove descends dorsally	20.00%	11	13.10%
3037	_Karlodinium_: Apical groove is narrow and short	Apical groove is narrow and short	0.00%	19	22.62%
3038	_Takayama_: Apical groove is sigmoid	Apical groove is sigmoid	20.00%	82	97.62%
3038	_Takayama_: Apical groove is horse shoe shaped	Apical groove is horse shoe shaped	0.00%	1	1.19%
3038	_Takayama_: Apical groove is arranged around the cingulum	Apical groove is arranged around the cingulum	0.00%	1	1.19%
3039	_Karenia_: Apical groove is straight	Apical groove is straight	20.00%	77	91.67%
3039	_Karenia_: Apical groove is ellipsoid	Apical groove is ellipsoid	0.00%	1	1.19%
3039	_Karenia_: Apical groove descends dorsally	Apical groove descends dorsally	0.00%	6	7.14%
3040	_Gymnodinium_: Apical groove is sigmoid	Apical groove is sigmoid	0.00%	1	1.19%
3040	_Gymnodinium_: Apical groove is horse shoe shaped	Apical groove is horse shoe shaped	20.00%	78	92.86%
3040	_Gymnodinium_: Apical groove goes around the apex of the cell	Apical groove goes around the apex of the cell	0.00%	3	3.57%
3040	_Gymnodinium_: Apical groove reaches the antapex	Apical groove reaches the antapex	0.00%	2	2.38%
3041	_Akashiwo_: Apical groove is horse shoe shaped	Apical groove is horse shoe shaped	0.00%	2	2.38%
3041	_Akashiwo_: Apical groove goes around the apex of the cell	Apical groove goes around the apex of the cell	20.00%	78	92.86%
3041	_Akashiwo_: Apical groove descends dorsally	Apical groove descends dorsally	0.00%	1	1.19%
3041	_Akashiwo_: Apical groove reaches the antapex	Apical groove reaches the antapex	0.00%	1	1.19%
3041	_Akashiwo_: Apical groove is arranged around the cingulum	Apical groove is arranged around the cingulum	0.00%	2	2.38%

ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q4

Question 4

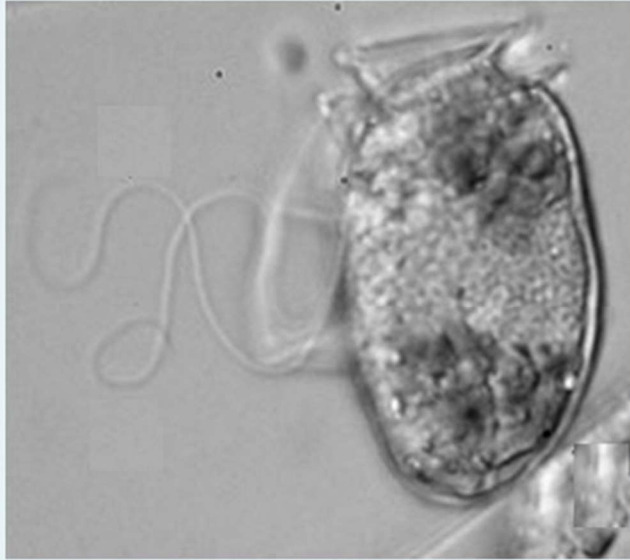
Not yet answered

Marked out of 1.00

Flag question

Edit question

The image shows a.....



Select one:

- a. Planozygote
- b. Hypnozygote
- c. Vegetative cell
- d. Haploid gamete
- e. Cyst

Q4	Partial credit	Count	Frequency
Planozygote	100.00%	75	89.29%
Hypnozygote	0.00%	1	1.19%
Vegetative cell	0.00%	8	9.52%
Haploid gamete	0.00%	0	0.00%
Cyst	0.00%	0	0.00%

ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q5

Question 5

Not yet answered

Marked out of 1.00

Flag question

Edit question

The image shows dinoflagellate gametes fusing during sexual reproduction. When the gamete pair are the same size, this is called.....

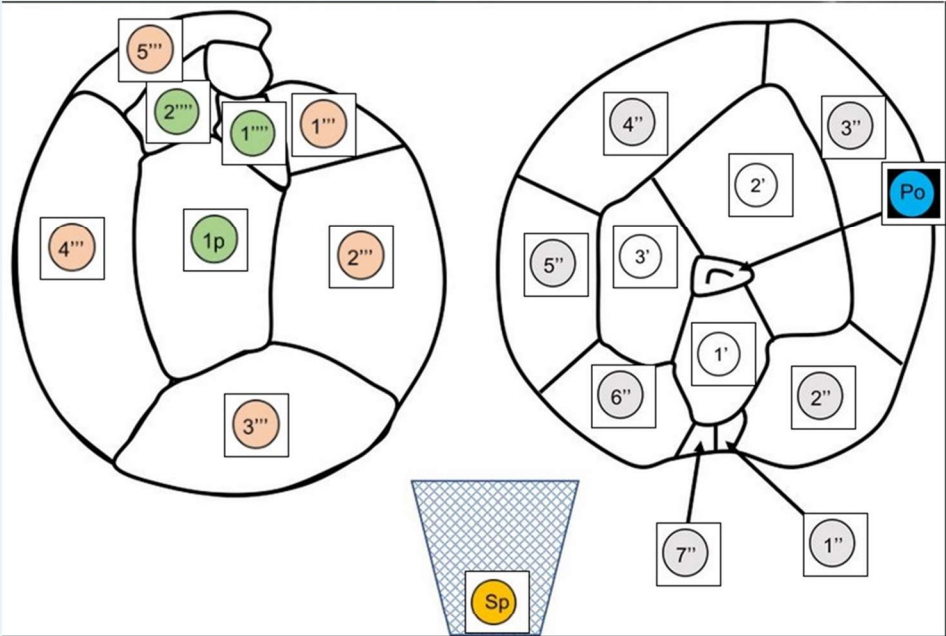
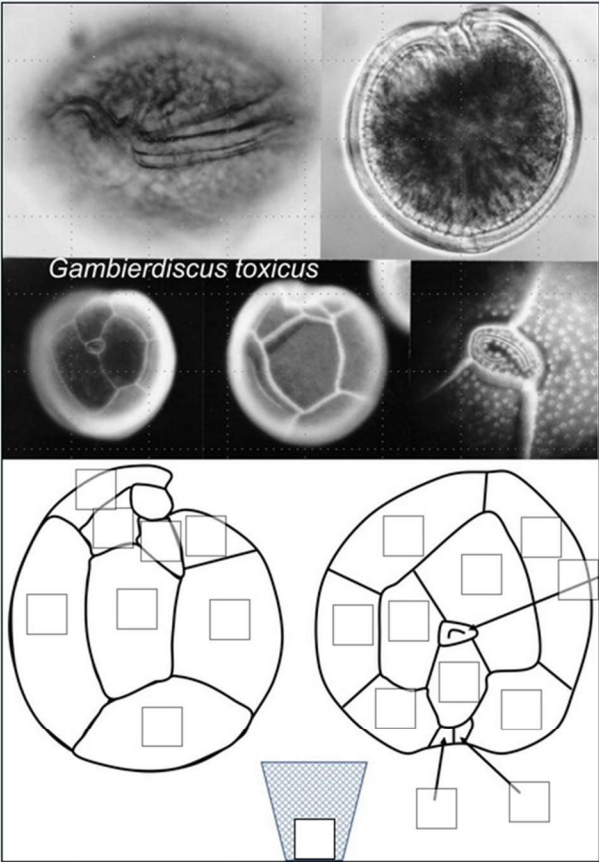


Select one:

- a. Isogamy
- b. Anisogamy
- c. Homothally
- d. Heterothally
- e. Oogamy

Q5	Partial credit	Count	Frequency
Isogamy	100.00%	82	97.62%
Anisogamy	0.00%	2	2.38%
Homothally	0.00%	0	0.00%
Heterothally	0.00%	0	0.00%
Oogamy	0.00%	0	0.00%
[No response]	0.00%	0	0.00%

ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q6



- 1'
- 1''
- 1'''
- 1''''
- 1p
- 2'
- 2''
- 2'''
- 2''''
- 3'
- 3''
- 3'''
- 3''''
- 4'
- 4''
- 4'''
- 4''''
- 5'
- 5''
- 6''
- 7''
- 7'''
- Po
- Sp

ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q6

Q6	Model response	Partial credit	Count	Frequency	Q6	Model response	Partial credit	Count	Frequency
24	Po	100.00%	84	100.00%	3	1'''	100.00%	83	98.81%
1	1'	100.00%	82	97.62%	3	1''''	0.00%	1	1.19%
1	4'	0.00%	1	1.19%	9	2'''	100.00%	84	100.00%
1	7''	0.00%	1	1.19%	12	3'''	100.00%	84	100.00%
6	2'	100.00%	84	100.00%	17	4'''	100.00%	84	100.00%
10	3'	100.00%	84	100.00%	19	5'''	100.00%	84	100.00%
2	1''	100.00%	82	97.62%	4	1''''	100.00%	83	98.81%
2	1'	0.00%	2	2.38%	4	2''''	0.00%	1	1.19%
7	2''	100.00%	82	97.62%	8	2''''	100.00%	73	86.90%
7	1''	0.00%	2	2.38%	8	1'''	0.00%	1	1.19%
11	3''	100.00%	82	97.62%	8	Sp	0.00%	10	11.90%
11	2''	0.00%	2	2.38%	5	1p	100.00%	74	88.10%
15	4''	100.00%	82	97.62%	5	2''''	0.00%	10	11.90%
15	3''	0.00%	2	2.38%					
18	5''	100.00%	82	97.62%					
18	4''	0.00%	2	2.38%					
20	6''	100.00%	82	97.62%					
20	5''	0.00%	2	2.38%					
22	7''	100.00%	82	97.62%					
22	6''	0.00%	2	2.38%					



## ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q7

Question 7  
Not yet answered  
Marked out of 1.00  
Flag question  
Edit question

Benthic dinoflagellates of the genus *Ostreopsis* are one of several causative agents of Ciguatera. The Kofoidian tabulation of this genus differs depending on which authors you follow, which complicates matters.

Following the tabulation according to Besada *et al.* 1982, Fraga *et al.* 2011 place the draggable items and place them in their right drop zone in the diagram supplied. There are more items than place holders, the items that you don't need you must place them in the bin provided.

**PLEASE NOTE: DO NOT PLACE ITEMS IN THE BIN UNTIL YOU ARE SURE YOU DON'T NEED THEM OTHERWISE YOU WON'T BE ABLE TO RETRIEVE THEM FROM THE BIN. BIN YOUR ITEMS ONLY AFTER YOU ANSWERED YOUR QUESTION**

From Larsen & Nguyen 2004



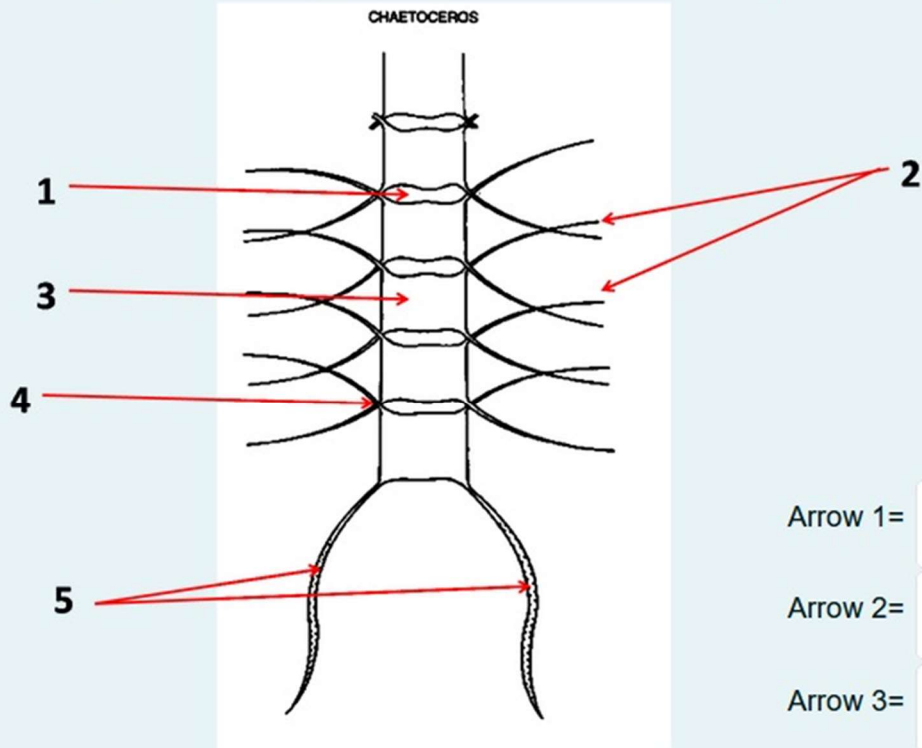
ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q7 answers

Q7	Model response	Partial credit	Count	Frequency		Q7	Model response	Partial credit	Count	Frequency
23	Po	100.00%	84	100.00%		3	1'''	100.00%	84	100.00%
1	1'	100.00%	57	67.86%		7	2'''	100.00%	84	100.00%
1	1''	0.00%	27	32.14%		11	3'''	100.00%	84	100.00%
5	2'	100.00%	84	100.00%		15	4'''	100.00%	84	100.00%
9	3'	100.00%	83	98.81%		18	5'''	100.00%	84	100.00%
9	7'''	0.00%	1	1.19%		4	1''''	100.00%	80	95.24%
13	4'	100.00%	57	67.86%		4	Sp	0.00%	4	4.76%
13	1'	0.00%	27	32.14%		8	2''''	100.00%	81	96.43%
						8	Sp	0.00%	3	3.57%
2	1''	100.00%	57	67.86%						
2	2''	0.00%	27	32.14%						
6	2''	100.00%	57	67.86%						
6	3''	0.00%	27	32.14%						
10	3''	100.00%	57	67.86%						
10	4'	0.00%	1	1.19%						
10	4''	0.00%	26	30.95%						
14	4''	100.00%	57	67.86%						
14	5''	0.00%	27	32.14%						
17	5''	100.00%	57	67.86%						
17	6''	0.00%	27	32.14%						
19	6''	100.00%	57	67.86%						
19	7''	0.00%	27	32.14%						

ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q8

Question 8  
Not yet answered  
Marked out of 1.00  
Flag question  
Edit question

Choose the right taxonomic terminology from the drop down menu to describe the different parts of a Chaetoceros chain.



- Arrow 1= Choose... ▾
- Arrow 2= Choose... ▾
- Arrow 3= Choose... ▾
- Arrow 4= Choose... ▾
- Arrow 5= Choose... ▾

- Arrow 1= Foramen ▾
- Arrow 2= Intercalary setae ▾
- Arrow 3= Valve mantle in girdle view ▾
- Arrow 4= Point of fusion of sibling setae ▾
- Arrow 5= Terminal setae ▾

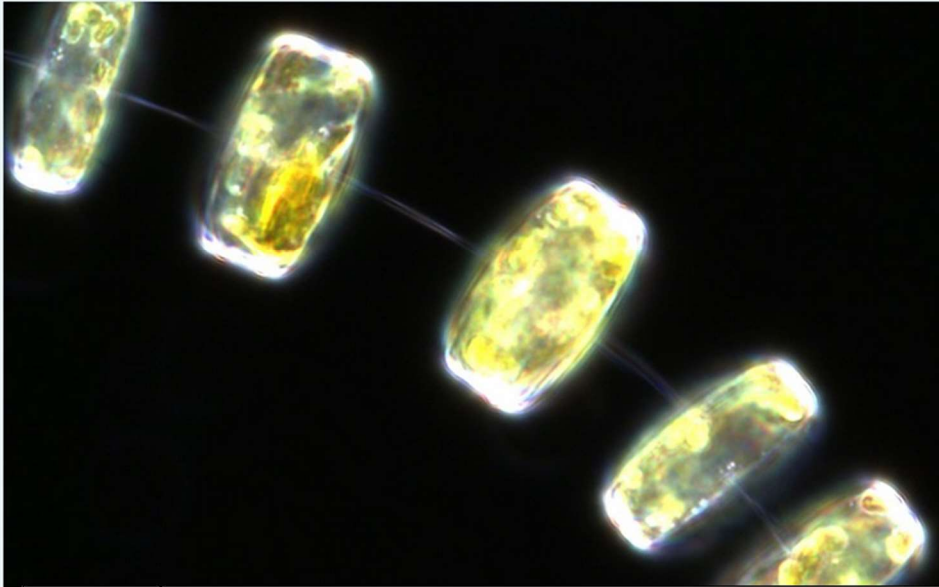
ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q8 answers

<b>Q8</b>	<b>Model response</b>	<b>Actual response</b>	<b>Partial credit</b>	<b>Count</b>	<b>Frequency</b>
<b>955</b>	Arrow 1=: Foramen	Foramen	20.00%	80	95.24%
<b>955</b>	Arrow 1=: Valve view	Valve view	0.00%	1	1.19%
<b>955</b>	Arrow 1=: Central inflation	Central inflation	0.00%	2	2.38%
<b>955</b>	Arrow 1=: Suture	Suture	0.00%	1	1.19%
<b>956</b>	Arrow 2=: Intercalary setae	Intercalary setae	20.00%	83	98.81%
<b>956</b>	Arrow 2=: Terminal setae	Terminal setae	0.00%	1	1.19%
<b>957</b>	Arrow 3=: Valve mantle in girdle view	Valve mantle in girdle view	20.00%	80	95.24%
<b>957</b>	Arrow 3=: Valve view	Valve view	0.00%	2	2.38%
<b>957</b>	Arrow 3=: Suture	Suture	0.00%	1	1.19%
<b>957</b>	Arrow 3=: Valvar plane	Valvar plane	0.00%	1	1.19%
<b>958</b>	Arrow 4=: Point of fusion of sibling setae	Point of fusion of sibling setae	20.00%	84	100.00%
<b>959</b>	Arrow 5=: Intercalary setae	Intercalary setae	0.00%	1	1.19%
<b>959</b>	Arrow 5=: Terminal setae	Terminal setae	20.00%	83	98.81%

ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q9

Question 9  
Not yet answered  
Marked out of 1.00  
Flag question  
Edit question

**Choose which statements are true about species belonging to the family Thalassiosiraceae?  
Each wrong answer will deduct 20% from your mark.**



Select one or more:

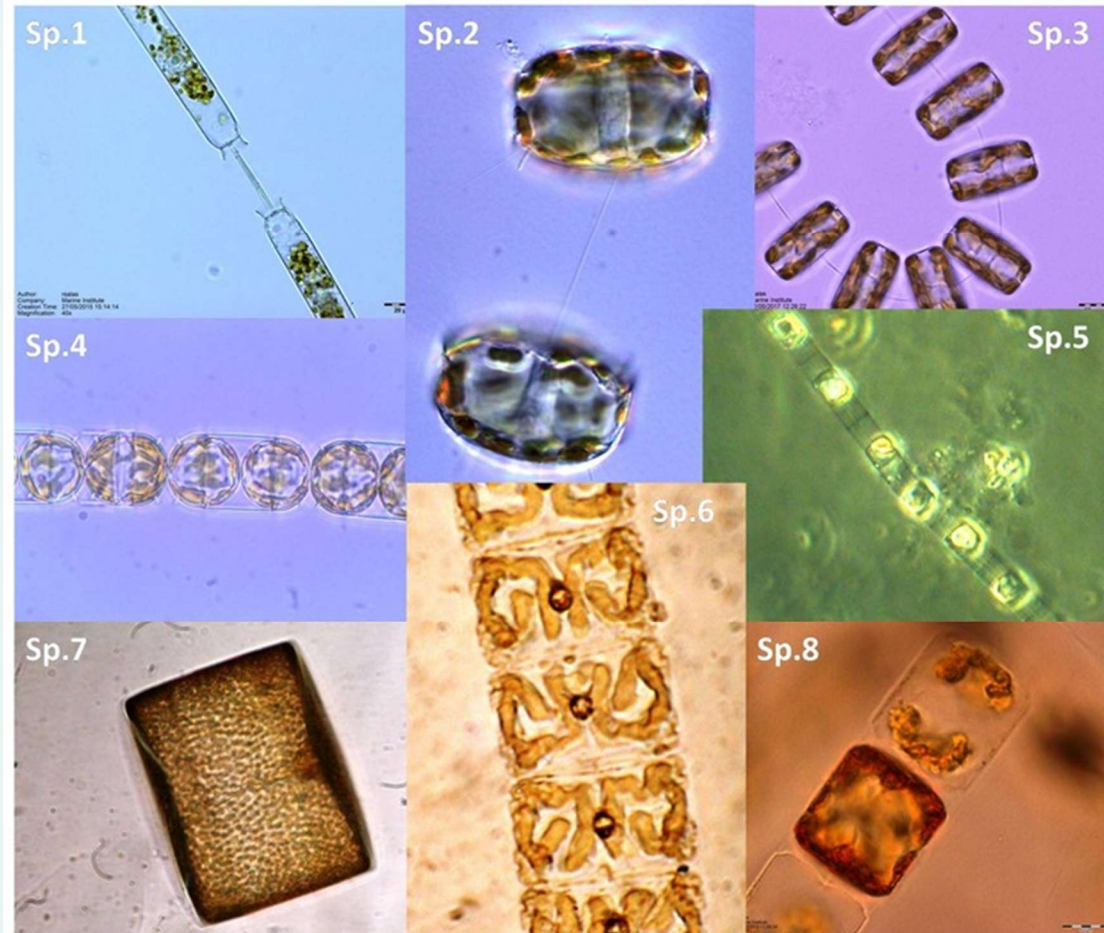
- a. Cells are linked by threads of organic matter from strutted processes
- b. They are unipolar centrics
- c. Their cell wall is known for having internal cribra and external foramina
- d. They have no labiate processes
- e. Their cell wall have a marginal ring of smaller labiate processes
- f. Their cell wall is known for having internal foramina and external cribra
- g. They have two larger marginal labiate processes
- h. They can have one or more labiate processes

Q9	Response	Partial credit	Count	Frequency
2588	Cells are linked by threads of organic matter from strutted processes	25.00%	81	96.43%
2589	They are unipolar centrics	-20.00%	10	11.90%
2590	Their cell wall is known for having internal cribra and external foramina	25.00%	74	88.10%
2591	They have no labiate processes	-20.00%	1	1.19%
2592	Their cell wall have a marginal ring of smaller labiate processes	25.00%	53	63.10%
2593	Their cell wall is known for having internal foramina and external cribra	-20.00%	6	7.14%
2594	They have two larger marginal labiate processes	-20.00%	1	1.19%
2595	They can have one or more labiate processes	25.00%	81	96.43%

ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q10

Question 10  
Not yet answered  
Marked out of 1.00  
Flag question  
Edit question

**Choose which of these species belong to the order Thalassiosirales? There are four families in this order: Thalassiosiraceae, Skeletonemataceae, Lauderiaceae and Stephanodiscaceae. Each wrong answer will deduct 20% from your mark.**



ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q10 answers

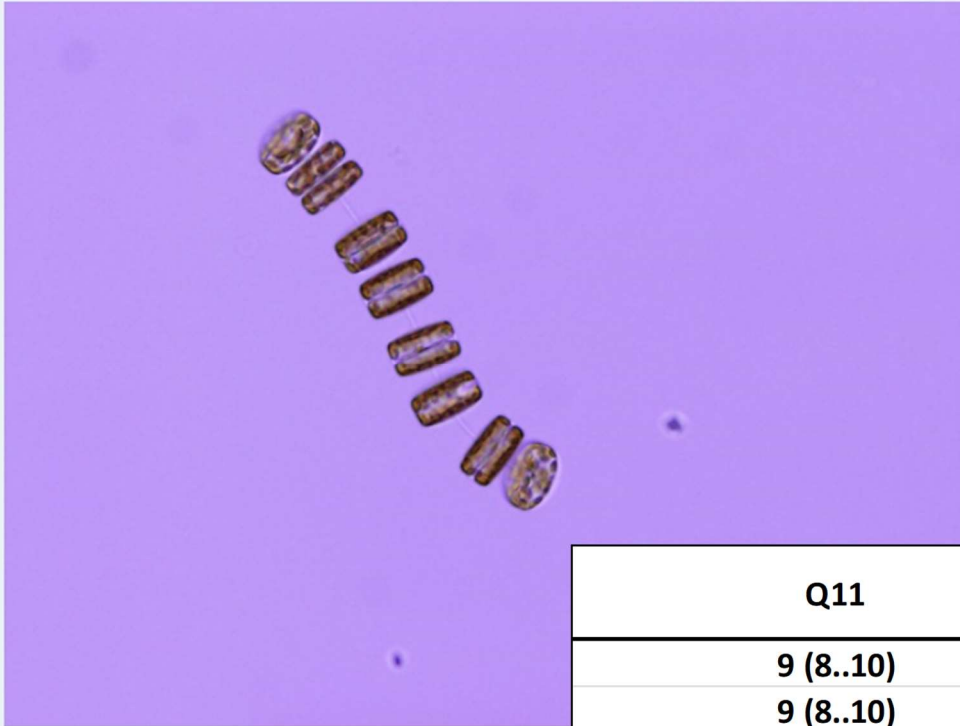
<b>Q10</b>	<b>Response</b>	<b>Order</b>	<b>Partial credit</b>	<b>Count</b>	<b>Frequency</b>
<b>2596</b>	Ditylum sp.	Lithodesmiales	-20.00%	2	2.38%
<b>2597</b>	Thalassiosira sp.	Thalassiosirales	25.00%	82	97.62%
<b>2598</b>	Thalassiosira sp.	Thalassiosirales	25.00%	82	97.62%
<b>2599</b>	Melosira sp.	Melosirales	-20.00%	2	2.38%
<b>2600</b>	Skeletonema sp.	Thalassiosirales	25.00%	77	91.67%
<b>2601</b>	Meuneria sp.	Naviculales	-20.00%	3	3.57%
<b>2602</b>	Coscinodiscus sp.	Coscinodiscales	-20.00%	3	3.57%
<b>2603</b>	Detonula sp.	Thalassiosirales	25.00%	76	90.48%



ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q11

Question **11**  
 Not yet answered  
 Marked out of 1.00  
 Flag question  
 Edit question

How many cells of *Thalassosira* are shown (numeric answers, 1,2,3 etc.)



Answer:

Q11	Actual response	Partial credit	Count	Frequency
9 (8..10)	9	100.00%	52	61.90%
9 (8..10)	8	100.00%	2	2.38%
9 (8..10)	09	100.00%	1	1.19%
9 (8..10)	10	100.00%	1	1.19%
[Did not match any answer]	13	0.00%	24	28.57%
[Did not match any answer]	12	0.00%	2	2.38%
[Did not match any answer]	14	0.00%	1	1.19%
[Did not match any answer]	16	0.00%	1	1.19%



ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q12

Question **12**

Not yet answered

Marked out of 1.00

Flag question

Edit question

Count all visible cells, even partial cells (numeric answers 1,2,3 etc.)



Answer: 26

Model response	Actual response	Partial credit	Count	Frequency
<b>26 (25..27)</b>	26	100.00%	77	91.67%
<b>26 (25..27)</b>	25	100.00%	6	7.14%
<b>[Did not match any answer]</b>	24	0.00%	1	1.19%

ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q13

Question 13

Not yet answered

Marked out of 1.00

Flag question

Edit question

The following dinoflagellates produce a large variety of toxic compounds. Match the toxin to their producer by dragging the toxin items onto the images.

The collage contains the following elements:

- Microscopic images:**
  - A. spinosum* (top left)
  - K. veneficum* (top middle)
  - K. brevis* (middle)
  - Other dinoflagellates (bottom left and bottom middle)
- Chemical structures:**
  - Azaspiracids:** A complex polyether structure with multiple hydroxyl groups.
  - Ciguatoxins:** A long-chain polyether with a complex ring system.
  - Karlotoxins:** A long-chain polyether with a complex ring system.
  - Yessotoxins:** A long-chain polyether with a complex ring system.
  - Brevetoxin:** A complex polyether with a complex ring system.
  - Okadaic Acid:** A complex polyether with a complex ring system.
  - Pinnatoxins:** A complex polyether with a complex ring system.
  - Spirolides:** A complex polyether with a complex ring system.
  - Saxitoxins:** A complex polyether with a complex ring system.

The row contains the following chemical structures:

- Azaspiracids
- Brevetoxin
- Ciguatoxins
- Karlotoxins
- Okadaic Acid
- Pinnatoxins
- Saxitoxins
- Spirolides
- Yessotoxins

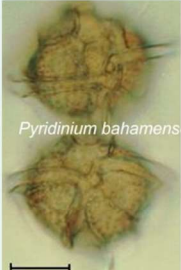
ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q13 answers

Part of question	Model response	Species	Partial credit	Count	Frequency
1	Azaspiracids	A.spinosum	100.00%	83	98.81%
1	Saxitoxins		0.00%	1	1.19%
2	Brevetoxins	K.brevis	100.00%	84	100.00%
3	Ciguatoxins	G.toxicus	100.00%	84	100.00%
4	Karlotoxins	K.veneficum	100.00%	84	100.00%
5	Okadaic Acid	D.acuta	100.00%	84	100.00%
6	Pinnatoxins	V.rugosum	100.00%	82	97.62%
6	Saxitoxins		0.00%	1	1.19%
6	Spirolides		0.00%	1	1.19%
7	Saxitoxins	P.bahamense	100.00%	84	100.00%
8	Saxitoxins		0.00%	7	8.33%
8	Spirolides	A.ostenfeldii	100.00%	77	91.67%
9	Saxitoxins		0.00%	1	1.19%
9	yessotoxins	L.polyedra	100.00%	83	98.81%
10	Saxitoxins	A.catenella	100.00%	84	100.00%

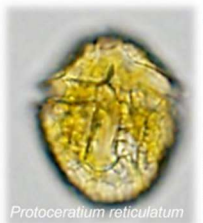
## ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q14

Question 14  
 Name:  
 Started on:  
 1.00  
 Flag  
 Question  
 Edit  
 Question


The following toxic/harmful dinoflagellate species belong to one of these two groups: Gonyaulales or Peridinales.  
 Do you know which one? Please drag and drop the images to their correct box.  
**PLEASE NOTE: ONCE YOU DROP THE ITEM ONTO THE BOX, YOU WON'T BE ABLE TO RETRIEVE IT, SO CHOOSE CAREFULLY YOUR OPTIONS BEFORE ANSWERING!!!**



*Pyridinium bahamensis*

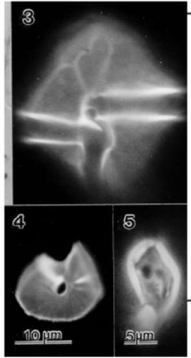


*Protoceratium reticulatum*

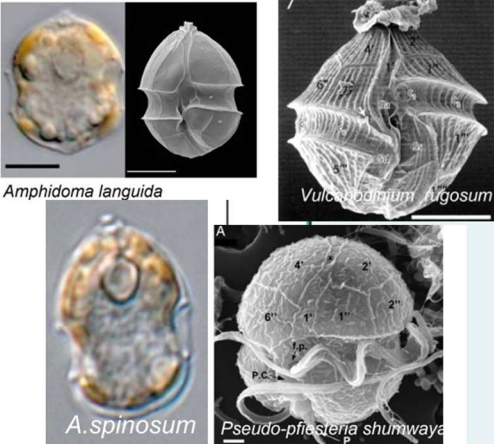


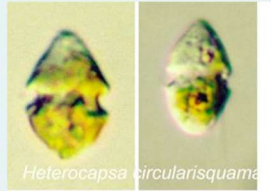
*Gonyaulax spinifera*

**Gonyaulales**

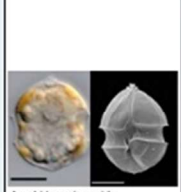


**Peridinales**






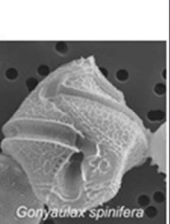
*Heterocapsa circularisquama*



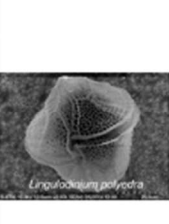
*Amphidoma languida*



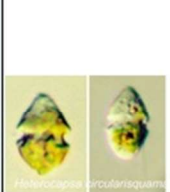
*A. spinosum*




*Gonyaulax spinifera*



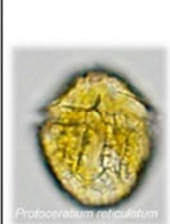
*Lingulodinium polyedra*



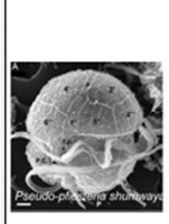
*Heterocapsa circularisquama*



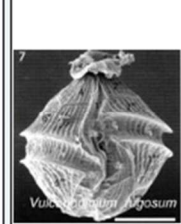
*Pyridinium bahamensis*



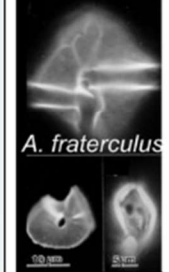
*Protoceratium reticulatum*



*Pseudo-pfiesteria shumwayae*



*Vulcanodinium rugosum*



*A. fraterculus*

ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q14 answers

Q14	Model response	Partial credit	Count	Frequency (%)
Peridinales	Am. Languida	100%	79	94.0
	Az. spinosum	100%	79	94.0
	H. circularis	100%	84	100.0
	P. shumwayae	100%	81	96.4
	V. rugosum	100%	80	95.2
	<b>Sub-total</b>		<b>81</b>	<b>96.0</b>
Gonyaulacales	G. spinifera	0%	0	0.0
	L. polyedra	0%	3	3.6
	P. bahamense	0%	3	3.5
	P. reticulatum	0%	1	1.2
	A. fraterculus	0%	4	4.7
	<b>Sub-total</b>		<b>2.2</b>	<b>2.6</b>

Q14	Model response	Partial credit	Count	Frequency (%)
Gonyaulacales	G. spinifera	100%	84	100.0
	L. polyedra	100%	81	96.4
	P. bahamense	100%	81	95.3
	P. reticulatum	100%	83	97.6
	A. fraterculus	100%	79	92.9
	<b>Sub-total</b>		<b>82</b>	<b>96.5</b>
Peridinales	Am. Languida	0%	4	4.8
	Az. spinosum	0%	3	3.6
	H. circularis	0%	0	0.0
	P. shumwayae	0%	1	1.2
	V. rugosum	0%	3	3.6
	<b>Sub-total</b>		<b>2.2</b>	<b>2.6</b>

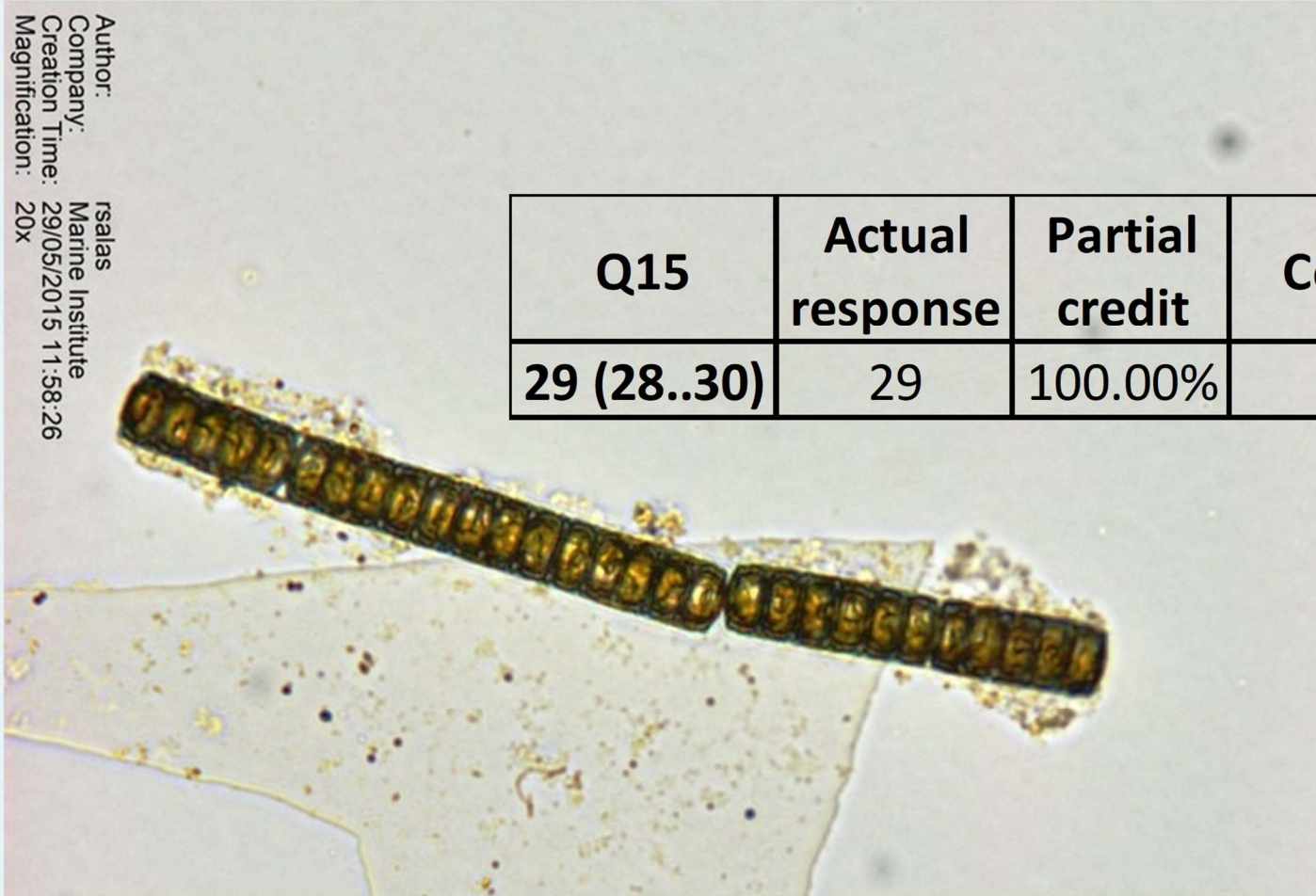


ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q15

Enumerate the Phytoplankton cells shown in this image: (Please use a numeral as the answer).

Answer: 29

Author: rsalas  
 Company: Marine Institute  
 Creation Time: 29/05/2015 11:58:26  
 Magnification: 20x



Q15	Actual response	Partial credit	Count	Frequency
29 (28..30)	29	100.00%	84	100.00%



ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q16

Question **16**

Not yet answered

Marked out of 1.00

Flag question

Edit question

Enumerate the Phytoplankton cells shown in this image including partially visible ones. (Please use a numeral as the answer).



Answer:

Q16	Actual response	Partial credit	Count	Frequency
9 (8..10)	9	100.00%	79	94.05%
9 (8..10)	8	100.00%	4	4.76%
[Did not match any answer]	4	0.00%	1	1.19%

## ANNEX XVI: Ocean Teacher HAB Quiz IPI202 Q17

Question 17

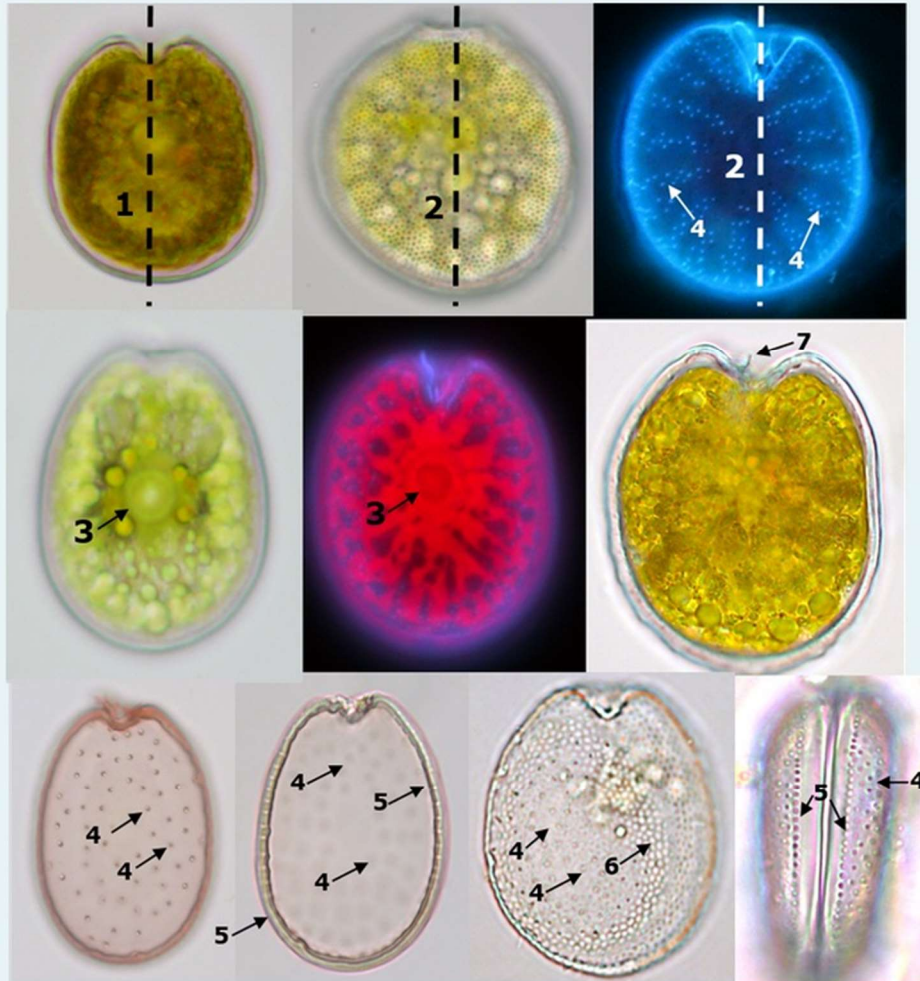
Not yet answered

Marked out of 1.00

Flag question

Edit question

Most benthic species of *Proocentrum* cannot be identified from only one photo. All diagnostic features are usually not visible and in focus at the same time, and a species is best illustrated by a series of photos. The most important diagnostic features are shown on the plate indicated by numbers - and the terminology used given in the text. Combine the numbers with the correct terminology.



- |                    |                       |   |
|--------------------|-----------------------|---|
| Number 7 indicates | An apical spine       | ⌵ |
| Number 5 indicates | Marginal pores        | ⌵ |
| Number 4 indicates | Valve pores           | ⌵ |
| Number 2 indicates | An asymmetric species | ⌵ |
| Number 6 indicates | Areolae               | ⌵ |
| Number 3 indicates | A pyrenoid            | ⌵ |
| Number 1 indicates | A symmetric species   | ⌵ |

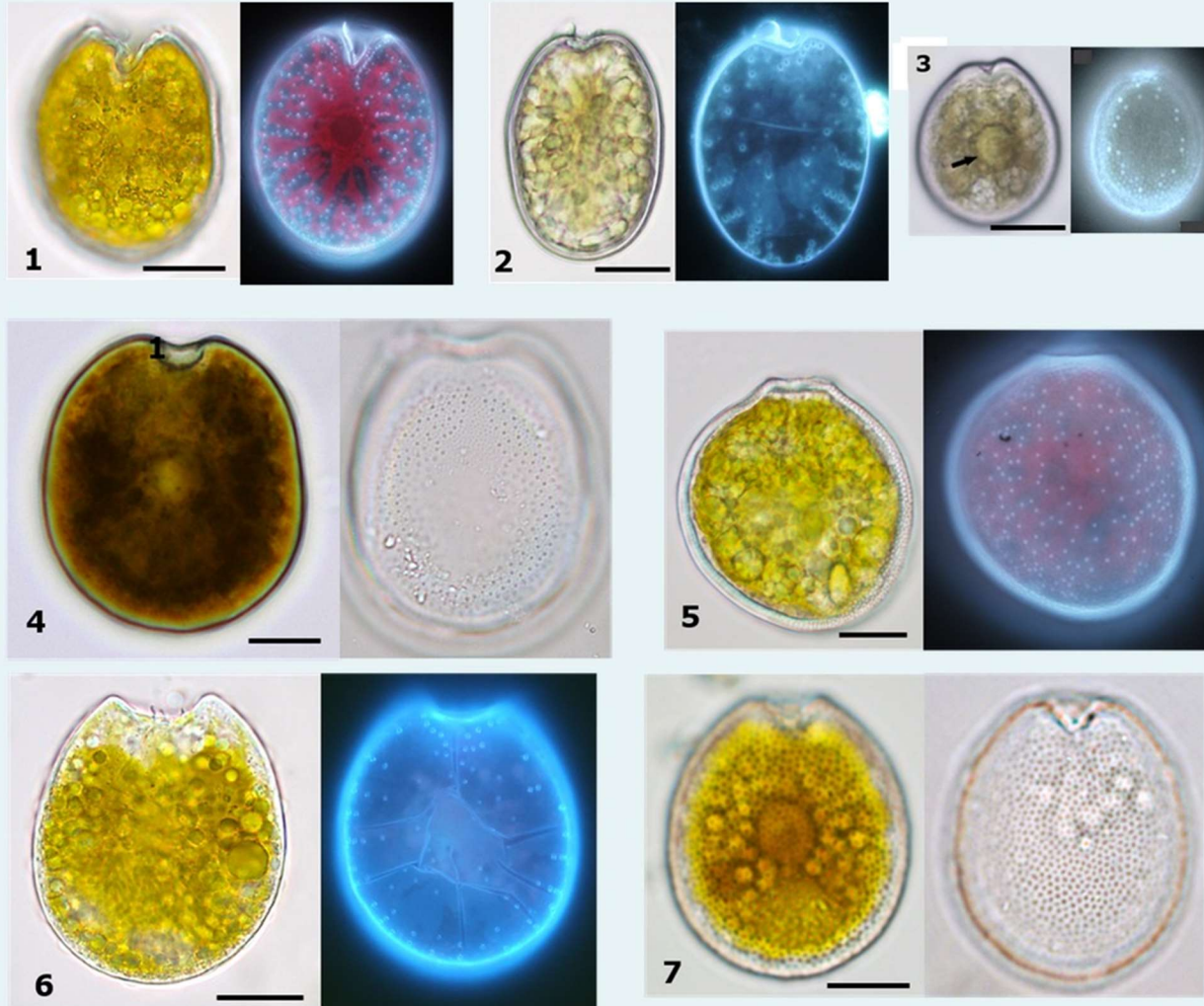
**ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q17 answers**

<b>Q17</b>	<b>Model response</b>	<b>Actual response</b>	<b>Partial credit</b>	<b>Count</b>	<b>Frequency</b>
<b>2963</b>	Number 1 indicates: A symmentric species	A symmentric species	14.29%	84	100.00%
<b>2964</b>	Number 2 indicates: An asymmetric species	An asymmetric species	14.29%	84	100.00%
<b>2965</b>	Number 3 indicates: A pyrenoid	A pyrenoid	14.29%	77	91.67%
<b>2965</b>	Number 3 indicates: Nucleus	Nucleus	0.00%	7	8.33%
<b>2966</b>	Number 4 indicates: Valve pores	Valve pores	14.29%	79	94.05%
<b>2966</b>	Number 4 indicates: Marginal pores	Marginal pores	0.00%	1	1.19%
<b>2966</b>	Number 4 indicates: Areolae	Areolae	0.00%	3	3.57%
<b>2966</b>	Number 4 indicates: Nucleus	Nucleus	0.00%	1	1.19%
<b>2967</b>	Number 5 indicates: Valve pores	Valve pores	0.00%	1	1.19%
<b>2967</b>	Number 5 indicates: Marginal pores	Marginal pores	14.29%	81	96.43%
<b>2967</b>	Number 5 indicates: Thecal plate	Thecal plate	0.00%	2	2.38%
<b>2968</b>	Number 6 indicates: Valve pores	Valve pores	0.00%	2	2.38%
<b>2968</b>	Number 6 indicates: Areolae	Areolae	14.29%	62	73.81%
<b>2968</b>	Number 6 indicates: Chloroplast	Chloroplast	0.00%	2	2.38%
<b>2968</b>	Number 6 indicates: Thecal plate	Thecal plate	0.00%	18	21.43%
<b>2969</b>	Number 7 indicates: An apical spine	An apical spine	14.29%	84	100.00%

## ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q18

Question **18**  
 Not yet answered  
 Marked out of 1.00  
 Flag question  
 Edit question

Identify the illustrated species - all seven species are illustrated by two photographs showing the diagnostic features of the species. All scale bars are 10 µm.



- Species 1
- Species 2
- Species 7
- Species 4
- Species 3
- Species 5
- Species 6

**ANNEX XVI: Ocean Teacher HAB Quiz IPI2022 Q18 Answers**

Q18	Model response	Actual response	Partial credit	Count	Frequency
2970	Species 1: Prorocentrum emerginatum	Prorocentrum emerginatum	14.29%	75	89.29%
2970	Species 1: Prorocentrum rhathymum	Prorocentrum rhathymum	0.00%	1	1.19%
2970	Species 1: Prorocentrum panamense	Prorocentrum panamense	0.00%	1	1.19%
2970	Species 1: Prorocentrum tsawwassenense	Prorocentrum tsawwassenense	0.00%	1	1.19%
2970	Species 1: Prorocentrum mexicanum	Prorocentrum mexicanum	0.00%	4	4.76%
2970	Species 1: Prorocentrum consutum	Prorocentrum consutum	0.00%	2	2.38%
2971	Species 2: Prorocentrum rhathymum	Prorocentrum rhathymum	14.29%	71	84.52%
2971	Species 2: Prorocentrum mexicanum	Prorocentrum mexicanum	14.29%	13	15.48%
2972	Species 3: Prorocentrum sipadanensis	Prorocentrum sipadanensis	14.29%	67	79.76%
2972	Species 3: Prorocentrum panamense	Prorocentrum panamense	0.00%	1	1.19%
2972	Species 3: Prorocentrum hoffmannianum	Prorocentrum hoffmannianum	0.00%	2	2.38%
2972	Species 3: Prorocentrum lima	Prorocentrum lima	0.00%	12	14.29%
2972	Species 3: Prorocentrum leve	Prorocentrum leve	0.00%	1	1.19%
2972	Species 3: Prorocentrum consutum	Prorocentrum consutum	0.00%	1	1.19%
2973	Species 4: Prorocentrum sipadanensis	Prorocentrum sipadanensis	0.00%	1	1.19%
2973	Species 4: Prorocentrum concavum	Prorocentrum concavum	14.29%	63	75.00%
2973	Species 4: Prorocentrum hoffmannianum	Prorocentrum hoffmannianum	0.00%	1	1.19%
2973	Species 4: Prorocentrum lima	Prorocentrum lima	0.00%	1	1.19%
2973	Species 4: Prorocentrum leve	Prorocentrum leve	0.00%	14	16.67%
2973	Species 4: Prorocentrum consutum	Prorocentrum consutum	0.00%	1	1.19%
2973	Species 4: Prorocentrum bimaculatum	Prorocentrum bimaculatum	0.00%	3	3.57%
2974	Species 5: Prorocentrum emerginatum	Prorocentrum emerginatum	0.00%	1	1.19%
2974	Species 5: Prorocentrum panamense	Prorocentrum panamense	14.29%	81	96.43%
2974	Species 5: Prorocentrum consutum	Prorocentrum consutum	0.00%	2	2.38%
2975	Species 6: Prorocentrum emerginatum	Prorocentrum emerginatum	0.00%	2	2.38%
2975	Species 6: Prorocentrum concavum	Prorocentrum concavum	0.00%	2	2.38%
2975	Species 6: Prorocentrum tsawwassenense	Prorocentrum tsawwassenense	14.29%	79	94.05%
2975	Species 6: Prorocentrum leve	Prorocentrum leve	0.00%	1	1.19%
2976	Species 7: Prorocentrum emerginatum	Prorocentrum emerginatum	0.00%	2	2.38%
2976	Species 7: Prorocentrum concavum	Prorocentrum concavum	0.00%	5	5.95%
2976	Species 7: Prorocentrum hoffmannianum	Prorocentrum hoffmannianum	14.29%	61	72.62%
2976	Species 7: Prorocentrum lima	Prorocentrum lima	0.00%	7	8.33%
2976	Species 7: Prorocentrum leve	Prorocentrum leve	0.00%	6	7.14%
2976	Species 7: Prorocentrum consutum	Prorocentrum consutum	0.00%	2	2.38%
2976	Species 7: Prorocentrum bimaculatum	Prorocentrum bimaculatum	0.00%	1	1.19%



**ANNEX XVII: Oceanteacher 2022 quiz results pg1**

<b>A. Code</b>	<b>Total</b>	<b>Q1</b>	<b>Q2</b>	<b>Q3</b>	<b>Q4</b>	<b>Q5</b>	<b>Q6</b>	<b>Q7</b>	<b>Q8</b>	<b>Q9</b>	<b>Q10</b>	<b>Q11</b>	<b>Q12</b>	<b>Q13</b>	<b>Q14</b>	<b>Q15</b>	<b>Q16</b>	<b>Q17</b>	<b>Q18</b>
<b>1</b>	<b>97.0</b>	100.0	91.1	100.0	100.0	100.0	100.0	100.0	78.6	100.0	100.0	100.0	100.0	89.3	100.0	100.0	100.0	100.0	85.7
<b>2</b>	<b>97.2</b>	87.5	91.1	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7	85.7
<b>3</b>	<b>95.9</b>	100.0	91.1	100.0	100.0	100.0	100.0	50.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7	100.0
<b>4</b>	<b>98.7</b>	100.0	91.1	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7	100.0
<b>5</b>	<b>79.7</b>	87.5	91.1	39.3	0.0	100.0	100.0	100.0	100.0	78.6	50.0	0.0	100.0	100.0	100.0	100.0	100.0	85.7	100.0
<b>6</b>	<b>94.8</b>	100.0	91.1	100.0	100.0	100.0	100.0	50.0	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7	100.0
<b>7</b>	<b>83.2</b>	37.5	91.1	100.0	100.0	100.0	100.0	100.0	78.6	100.0	100.0	0.0	100.0	100.0	58.9	100.0	100.0	71.4	57.1
<b>8</b>	<b>97.9</b>	100.0	71.4	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	89.3	100.0	100.0	100.0	100.0	100.0
<b>9</b>	<b>89.5</b>	100.0	100.0	78.6	100.0	100.0	100.0	55.4	100.0	100.0	75.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>10</b>	<b>99.5</b>	100.0	91.1	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>11</b>	<b>96.5</b>	100.0	100.0	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	57.1
<b>13</b>	<b>96.0</b>	100.0	100.0	100.0	100.0	100.0	100.0	55.4	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7	85.7
<b>14</b>	<b>90.5</b>	87.5	91.1	78.6	0.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7	85.7
<b>15</b>	<b>99.2</b>	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7
<b>16</b>	<b>99.5</b>	100.0	91.1	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>17</b>	<b>98.4</b>	100.0	91.1	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>18</b>	<b>91.2</b>	100.0	91.1	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	71.4
<b>19</b>	<b>90.2</b>	87.5	100.0	78.6	100.0	100.0	100.0	55.4	100.0	100.0	100.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>20</b>	<b>99.5</b>	100.0	91.1	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>21</b>	<b>91.6</b>	100.0	80.4	78.6	100.0	100.0	100.0	100.0	78.6	100.0	55.4	100.0	100.0	100.0	78.6	100.0	100.0	100.0	71.4
<b>22</b>	<b>83.5</b>	100.0	80.4	100.0	0.0	100.0	100.0	50.0	100.0	100.0	100.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	71.4
<b>23</b>	<b>87.3</b>	100.0	91.1	100.0	0.0	100.0	100.0	100.0	78.6	100.0	100.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>24</b>	<b>92.0</b>	100.0	100.0	100.0	100.0	100.0	100.0	55.4	100.0	100.0	100.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>25</b>	<b>100.0</b>	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>26</b>	<b>89.8</b>	100.0	100.0	58.9	100.0	100.0	100.0	55.4	100.0	100.0	100.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>27</b>	<b>99.2</b>	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7
<b>28</b>	<b>98.1</b>	100.0	100.0	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7
<b>29</b>	<b>96.6</b>	87.5	100.0	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	71.4
<b>30</b>	<b>99.2</b>	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7
<b>31</b>	<b>98.1</b>	100.0	100.0	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7
<b>32</b>	<b>97.3</b>	100.0	91.1	78.6	100.0	100.0	100.0	100.0	100.0	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>33</b>	<b>96.8</b>	100.0	80.4	78.6	100.0	100.0	100.0	100.0	100.0	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>34</b>	<b>95.5</b>	100.0	91.1	78.6	100.0	100.0	100.0	100.0	100.0	66.1	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	82.1
<b>35</b>	<b>99.5</b>	100.0	91.1	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>36</b>	<b>98.4</b>	100.0	91.1	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>37</b>	<b>98.9</b>	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	78.6	100.0	100.0	100.0	100.0
<b>38</b>	<b>93.6</b>	62.5	91.1	58.9	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7	85.7
<b>39</b>	<b>97.6</b>	100.0	91.1	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7
<b>40</b>	<b>96.8</b>	100.0	91.1	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7	85.7
<b>41</b>	<b>96.0</b>	100.0	91.1	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7	71.4
<b>42</b>	<b>96.0</b>	100.0	91.1	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7	71.4
<b>43</b>	<b>88.5</b>	100.0	100.0	100.0	100.0	100.0	83.9	55.4	100.0	55.4	100.0	100.0	100.0	89.3	78.6	100.0	100.0	85.7	42.9

**ANNEX XVII: Oceanteacher 2022 quiz results pg2**

<b>A. Code</b>	<b>Total</b>	<b>Q1</b>	<b>Q2</b>	<b>Q3</b>	<b>Q4</b>	<b>Q5</b>	<b>Q6</b>	<b>Q7</b>	<b>Q8</b>	<b>Q9</b>	<b>Q10</b>	<b>Q11</b>	<b>Q12</b>	<b>Q13</b>	<b>Q14</b>	<b>Q15</b>	<b>Q16</b>	<b>Q17</b>	<b>Q18</b>
<b>44</b>	<b>81.2</b>	87.5	80.4	78.6	0.0	100.0	100.0	55.4	100.0	78.6	100.0	0.0	100.0	89.3	100.0	100.0	100.0	100.0	85.7
<b>45</b>	<b>95.1</b>	75.0	91.1	78.6	100.0	100.0	100.0	100.0	100.0	66.1	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>46</b>	<b>89.8</b>	75.0	91.1	100.0	100.0	0.0	100.0	50.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>47</b>	<b>92.7</b>	87.5	91.1	100.0	100.0	100.0	100.0	100.0	58.9	100.0	50.0	100.0	100.0	100.0	78.6	100.0	100.0	100.0	100.0
<b>50</b>	<b>94.4</b>	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>51</b>	<b>94.4</b>	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>52</b>	<b>94.4</b>	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>53</b>	<b>99.2</b>	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7
<b>54</b>	<b>92.9</b>	100.0	100.0	100.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	71.4
<b>55</b>	<b>93.2</b>	100.0	91.1	78.6	100.0	100.0	100.0	55.4	100.0	50.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>56</b>	<b>75.8</b>	87.5	91.1	78.6	100.0	100.0	100.0	50.0	100.0	26.8	30.4	0.0	100.0	89.3	78.6	100.0	100.0	71.4	57.1
<b>57</b>	<b>84.6</b>	75.0	100.0	100.0	100.0	100.0	100.0	55.4	100.0	58.9	78.6	0.0	100.0	78.6	100.0	100.0	100.0	85.7	85.7
<b>58</b>	<b>98.2</b>	100.0	80.4	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7
<b>59</b>	<b>90.4</b>	100.0	91.1	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	0.0	100.0	100.0	78.6	100.0	100.0	71.4	85.7
<b>60</b>	<b>85.8</b>	87.5	91.1	100.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	0.0	100.0	100.0	78.6	100.0	100.0	100.0	85.7
<b>61</b>	<b>91.7</b>	100.0	100.0	100.0	100.0	100.0	94.6	55.4	100.0	100.0	100.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>62</b>	<b>75.9</b>	75.0	80.4	19.6	100.0	100.0	87.5	55.4	58.9	55.4	100.0	0.0	100.0	100.0	100.0	100.0	100.0	57.1	71.4
<b>63</b>	<b>94.7</b>	100.0	80.4	100.0	100.0	100.0	100.0	55.4	100.0	66.1	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>64</b>	<b>95.9</b>	75.0	100.0	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	71.4
<b>65</b>	<b>95.8</b>	87.5	100.0	100.0	100.0	100.0	100.0	100.0	100.0	78.6	100.0	100.0	100.0	100.0	100.0	100.0	100.0	71.4	85.7
<b>66</b>	<b>85.1</b>	75.0	100.0	100.0	100.0	100.0	87.5	100.0	100.0	58.9	78.6	0.0	100.0	100.0	100.0	100.0	100.0	85.7	42.9
<b>67</b>	<b>100.0</b>	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>68</b>	<b>92.0</b>	100.0	100.0	100.0	100.0	100.0	100.0	55.4	100.0	100.0	100.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>70</b>	<b>100.0</b>	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>71</b>	<b>93.1</b>	100.0	91.1	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	0.0	100.0	100.0	100.0	100.0	100.0	85.7	100.0
<b>72</b>	<b>93.2</b>	100.0	80.4	100.0	100.0	100.0	100.0	50.0	100.0	58.9	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7	100.0
<b>73</b>	<b>89.9</b>	100.0	91.1	100.0	100.0	100.0	87.5	55.4	100.0	100.0	100.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	82.1
<b>74</b>	<b>93.7</b>	100.0	91.1	100.0	100.0	100.0	100.0	50.0	100.0	58.9	100.0	100.0	100.0	100.0	100.0	100.0	100.0	85.7	100.0
<b>75</b>	<b>99.4</b>	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	89.3	100.0	100.0	100.0	100.0
<b>76</b>	<b>84.8</b>	100.0	91.1	58.9	100.0	100.0	87.5	55.4	100.0	55.4	75.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>77</b>	<b>96.9</b>	100.0	100.0	100.0	100.0	100.0	87.5	55.4	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>78</b>	<b>84.4</b>	100.0	100.0	100.0	0.0	100.0	100.0	100.0	100.0	100.0	100.0	0.0	100.0	89.3	100.0	100.0	100.0	85.7	42.9
<b>79</b>	<b>77.7</b>	100.0	71.4	58.9	100.0	0.0	87.5	50.0	100.0	100.0	78.6	0.0	100.0	89.3	100.0	100.0	100.0	71.4	85.7
<b>80</b>	<b>81.3</b>	100.0	80.4	100.0	100.0	100.0	87.5	100.0	100.0	0.0	0.0	100.0	100.0	100.0	50.0	100.0	100.0	85.7	57.1
<b>81</b>	<b>68.7</b>	100.0	80.4	100.0	100.0	100.0	87.5	100.0	100.0	0.0	0.0	0.0	0.0	100.0	78.6	100.0	100.0	71.4	14.3
<b>82</b>	<b>75.7</b>	50.0	91.1	39.3	0.0	100.0	100.0	55.4	100.0	100.0	55.4	0.0	100.0	100.0	100.0	100.0	100.0	85.7	85.7
<b>83</b>	<b>92.5</b>	100.0	91.1	100.0	100.0	100.0	46.4	55.4	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	71.4
<b>84</b>	<b>100.0</b>	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
<b>85</b>	<b>97.9</b>	100.0	91.1	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	71.4
<b>86</b>	<b>99.4</b>	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0	89.3	100.0	100.0	100.0	100.0	100.0
<b>90</b>	<b>77.4</b>	50.0	71.4	100.0	100.0	100.0	100.0	55.4	78.6	75.0	75.0	100.0	100.0	89.3	78.6	100.0	0.0	71.4	42.9
<b>Total</b>	<b>92.6</b>	<b>92.9</b>	<b>92.9</b>	<b>89.3</b>	<b>89.3</b>	<b>96.4</b>	<b>96.4</b>	<b>83.9</b>	<b>96.4</b>	<b>89.3</b>	<b>92.9</b>	<b>66.1</b>	<b>98.2</b>	<b>98.2</b>	<b>96.4</b>	<b>100.0</b>	<b>98.2</b>	<b>92.9</b>	<b>85.7</b>